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### INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 5:	1 42	(11) International Publication Number:	WO 94/03075		
A23L 1/305		(43) International Publication Date:	17 February 1994 (17.02.94)		

(21) International Application Number: PCT/US93/07190

(22) International Filing Date: 29 July 1993 (29.07.93)

 (30) Priority data:
 923,780
 31 July 1992 (31.07.92)
 US

 946,235
 16 September 1992 (16.09.92)
 US

 029,335
 4 March 1993 (04.03.93)
 US

 040,510
 31 March 1993 (31.03.93)
 US

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(81) Designated States: AT, AU, BB, BG, BR, CA, CH, CZ, DE, DK, ES, FI, GB, HU, JP, KP, KR, LK, LU, MG, MN, MW, NL, NO, NZ, PL, PT, RO, RU, SD, SE, SK, UA, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).

#### Published

Without international search report and to be republished upon receipt of that report.

(54) Title: MORPHOGEN-ENRICHED DIETARY COMPOSITION

#### (57) Abstract

Disclosed are methods and compositions useful in dietary applications and capable of enhancing tissue morphogenesis, including tissue development and viability in a mammal, particularly a human. The methods and compositions include a morphogen which, when provided to an individual as a food formulation or supplement, is capable of enhancing tissue development and viability in the individual.

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### MORPHOGEN-ENRICHED DIETARY COMPOSITION

## Field of the Invention

This invention relates generally to the field of dietary compositions and supplements.

# Background of the Invention

The present invention relates to compositions 10 useful as mammalian dietary compositions and supplements. In particular, the invention relates to food additives and dietary supplements capable of enhancing tissue morphogenesis and development, particularly in individuals at risk for normal tissue 15 development and viability. Examples of such individuals include infants, particularly prematurelyborn ("preterm") and low birth weight infants, and juveniles; aged individuals; and individuals experiencing altered metabolic function and/or 20 suffering from metabolic dysfunctions and other disorders that threaten organ or tissue function or viability, such as can result from malnutrition or starvation, autoimmune diseases, organ cirrhosis and other tissue necrotizing dysfunctions, or disorders 25 associated with aging cells (cell senescence.)

Mammalian infants are nourished by mother's milk until such time as they can digest food solids. Infant formulas now exist for humans and other mammals which can supplant or supplement mother's milk. The formulas may be milk based (e.g., cow milk) or non-milk-based

Particularly at risk are prematurely (e.g., soy). born infants whose tissues and organs are at an earlier stage of development, and whose nutritional requirements may differ from those of full term 5 infants. Formula development is an ongoing endeavor to more accurately mimic the beneficial aspects of mother's milk. Nevertheless, despite the efforts of many researchers, infant formulas still differ in a number of significant ways from human milk. 10 this is due because human milk has many substances, such as immunoglobulins, free amino acids, polyamines, nucleotides and polyunsaturated fatty acids not In addition, present, for example, in cow's milk. while infant formulas try to mimic the protein quantity 15 found in human milk, the foreign proteins typically are present in the formula as hydrolysates to avoid rejection or reaction by the infant's digestive system. The proteins are present primarily as amino acid sources rather than as functional proteins as might normally be transmitted by the nursing mother to the 20 infant. In addition, human milk may contain unidentified growth and differentiation factors that are important for overall tissue and skeletal development.

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Another group of individuals with potentially unique nutritional requirements are individuals undergoing metabolic changes which may result from periods of intense growth or stress, including, for example, pregnant women and drowning victims. Other sources of stress to the body may result from

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malnutrition or starvation, or from metabolic disorders that affect organ viability, such as autoimmune disease and organ cirrhosis. Aged individuals, and postmenopausal women also have altered or slower metabolic function. All of these individuals are at risk for tissue damage or loss of tissue function due to altered metabolic function.

Reduced or lost tissue function due to malnutrition 10 also is found in many patients admitted to hospitals (protein energy malnutrition, "PEM"). nutritional support for such patients, while not a primary mode of treatment is, nevertheless, an important factor for therapy and recovery. 15 therefore important to administer a nutritionally balanced diet given orally, enterally or parenterally, adequate to the needs of the patient. This is especially true for those patients where conventional feeding is contraindicated (e.g., in dehydrated or gastroenterological patients) or is insufficient (e.g., 20 in hypercatabolic patients). The enteral or oral mode of administration of foods typically is preferable to parenteral modes because of the lower morbidity, trophic effect upon the intestinal mucosa, reduced 25 dependency on instrumentation and lower costs.

It is an object of this invention to provide dietary compositions and supplements for enhancing tissue morphogenesis, including tissue growth,

30 development, maintenance and viability in a mammal, particularly a human. Another object of the invention is to provide an infant formula capable of enhancing tissue development in an infant or juvenile. Still another object is to provide an an infant formula that

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more closely mimics a nursing mother's milk. Another object of the invention is to provide dietary supplements for individuals at risk for normal tissue development, growth, maintenance and viability, including premature infants, aged individuals and individuals with altered metabolic function and/or suffering from disorders or metabolic dysfunctions which threaten organ viability and function. These and other objects and features of the invention will be apparent from the description, drawings, and claims which follow.

### Summary of the Invention

The present invention provides compositions and methods useful in dietary applications and capable of enhancing tissue morphogenesis, including tissue growth, development, maintenance and viability in a mammal, particularly a human. The dietary compositions and supplements of this invention comprise a morphogenic protein ("morphogen"), as described herein, which, when provided to an individual as a food formulation or supplement, is capable of enhancing tissue development, growth, maintenance and/or viability in the individual. The compositions and processes provided herein are suitable for both infants and adults, and as part of clinical nutrition.

As used herein, "enhancing tissue viability" is understood to mean protecting tissue from lost or reduced tissue function due to cell damage or cell senescence, including inducing cells to maintain their differentiated phenotype, inducing regeneration of damaged tissue, and/or inhibiting additional damage

thereto. "Morphogenically effective concentration" is understood to mean a concentration sufficient to enhance tissue development and tissue viability in an individual at risk for tissue damage and/or reduced or 5 lost tissue function due to insufficient nutritional considerations, tissue damage associated therewith, and/or incomplete tissue development, regardless of etiology. The ability of morphogens to repair, regenerate and protect various disparate tissues, including but not limited to, tissues of the 10 gastrointestinal tract, including the oral mucosa, liver tissue, dentin tissue, periodontal tissue, nerve tissue, bone tissue, and any tissue at risk of damage due to immune response-mediated tissue destruction, 15 including ischemia-reperfusion related tissue damage are disclosed in international applications US 92/01968 (WO 92/15323), US 92/07358 (WO 93/04692) and US 92/07232 (WO 93/05751) respectively, the disclosures of which are incorporated herein by reference. "Morphogen-solubilizing molecule" is understood to mean 20 a molecule capable of maintaining a morphogen in soluble form in physiologically buffered solutions. "Food formulation" is understood to mean a dietary composition normally ingested by an individual to satisfy the body's fundamental nutritional requirements; "dietary supplement" is understood to mean supplemental compositions ingested by an individual in addition to the food formulations ingested to satisfy the fundamental nutritional 30 requirements. Multivitamin and iron tablets are examples of commmon dietary supplements. composition" is understood to include both food formulations and dietary supplements. As used herein,

the term "infant formula" is understood to refer to the

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well established infant compositions as defined by the American Academy of Pediatrics (AAP) and the AAP Committee on Nutrition ((1985) Pediatrics 75:976, the European Society of Pediatric Gastroenterology and Nutrition (ESPGAN) and the ESPGAN Committee on Nutrition ((1987) Acta Paed Scan Suppl:330), including recent updates published by these committees on infant formula nutritional guidelines.

The dietary composition or supplement preferably is administered orally, and may be provided in liquid form or as a powder to be dissolved in a beverage.

Alternatively, the dietary supplement may be provided as a solid, e.g., in a capsular, tablet, troche or lozenge form; or, the supplement may be provided as an aerosol, for oral or nasal administration. Where oral administration is not possible or desirable, other administration routes are envisioned. For example, for some premature infants, or for intibated patients, parenteral administration may be required, e.g., via an enteral feeding tube.

The morphogen may be provided alone or in association with one or more suitable excipients or carriers, and/or in combination with other beneficial molecules such as vitamins, minerals, lipids, fiber sources and the like. The dietary supplements also may include pharmaceutically acceptable inert materials for use as binders or stabilizers, including magnesium stearate or calcium carbonate. The morphogen may be

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formulated together with one or more normal food ingredients, e.g., as part of a food formulation.

Alternatively or, in addition, the morphogen may be provided as a dietary supplement in, for example, tablet or syrup form.

The mature form of the morphogen, or active truncated forms thereof which may be formulated in the composition, further may be provided in association 10 with a morphogen precursor "pro" domain, which is known to enhance the solubility of the protein in physiologically buffered solutions. Other useful molecules known to enhance protein solubility include casein, including derivatives, salts and analogs 15 thereof, as well as other milk components, and various serum and milk serum proteins. Additional useful molecules which may be associated with the morphogen include tissue targeting molecules capable of directing the morphogen to a desired target tissue. 20 targeting molecules envisioned to be useful in the treatment protocols of this invention include antibodies, antibody fragments or other binding proteins which interact specifically with surface molecules on the target tissue cells.

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Still another useful tissue targeting molecule may be part or all of a morphogen precursor "pro" domain. Morphogens may be synthesized in one tissue and secreted and transported to another tissue. For example, while the protein has been shown to be active in bone tissue, the primary source of OP-1 synthesis appears to be the tissue of the urogenic system (e.g., renal and bladder tissue), with secondary expression levels occurring in the brain, heart, lungs and

gastrointestinal tract (GI tract, see below.)

Moreover, the protein has been identified in serum,

saliva and various milk forms. In addition, the

secreted form of the protein comprises the mature dimer

in association with the pro domain of the intact

morphogen sequence. Accordingly, the associated

morphogen pro domains may act to target specific

morphogens to different tissues in vivo. As described

below, morphogen species comprising the pro domain may

be obtained from the culture medium of morphogen
secreting mammalian cells. Alternatively, a tissue
targeting species may be formulated by complexing the

mature dimer (or an active fragment thereof) with part

or all of a pro domain.

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Associated tissue targeting or solubility-enhancing molecules also may be covalently linked to the morphogen using standard chemical means.

In one preferred embodiment, the morphogen comprises part of an infant formula. The infant formula may be milk-based or nonmilk-based, e.g., soy-based. A typical ready-to-feed morphogen-enriched formulation for infants, when diluted to feeding concentrations, comprises, in addition to the morphogen added to the formula, from about 1-5% by weight fat, from about 0.01 to about 0.5% by weight immunoglobulins as appropriate, from about 4-10% by weight carbohydrate in a quantity substantially to mimic the carbohydrate content of human mother's milk, from about 0.5 to 4% by weight

protein in a quantity substantially to mimic the protein content of human mother's milk, optional vitamins and minerals as required, a total solids content of from about 8 to 17% by weight, and the remainder water.

In another preferred embodiment, the dietary composition is formulated for individuals at risk for reduced or lost tissue function, such as postmenopausal 10 women, elderly individuals, undernourished or malnourished individuals, dehydrated individuals, drowning victims, individuals suffering from metabolic disorders including an endocrine imbalance, gastrointestinal disorders, or immune-compromised 15 individuals. Undernourished or malnourished individuals include those suffering from a lack of food (starvation) and/or eating disorders (e.g., anorexia nervosa), and/or suffering from a maladsorption syndrome (e.g., individuals afflicted with digestive or intestinal fistulas, shortened bowel, or 20 hypercatabolism.) Individuals receiving a medical therapy, including radiotherapy, chemotherapy or a surgical procedure also are at risk for reduced or lost tissue function as a result of a therapy-related 25 malabsorption-malnutrition dysfunction. embodiment, the dietary supplement is formulated for individuals undergoing periods of increased growth or stress, such as infants and juveniles, or pregnant or lactating women. In another embodiment, the dietary 30 supplement is formulated for individuals at risk for reduced or lost organ function as results from tissue cirrhosis or an autoimmune disease.

Morphogen-enriched nutritional products, particularly clinical nutrition products for use in hospital or other clinical settings, in addition to comprising a morphogen preferably are based on the utilization of diverse other protein sources (casein, sodium and calcium caseinate, isolated soy protein, protein hydrolyzates and/or crystalline amino acids) mixtures of vegetable and animal fats, carbohydrates (basically glucose polymers), vitamins and minerals to meet, at least, the dietary intakes recommended for healthy individuals (see, for example, Committee on Dietary Allowances, Food and Nutrition Board, Nat Acad Sci, 9th Ed, 1980).

Among the morphogens useful in this invention are 15 proteins originally identified as osteogenic proteins, such as the OP-1, OP-2 and CBMP2 proteins, as well as amino acid sequence-related proteins such as DPP (from Drosophila), Vgl (from Xenopus), Vgr-1 (from mouse, see 20 U.S. 5,011,691 to Oppermann et al.), GDF-1 (from mouse, see Lee (1991) PNAS 88:4250-4254), all of which are presented in Table II and Seq. ID Nos.5-14), and the recently identified 60A protein (from Drosophila, Seq. ID No. 24, see Wharton et al. (1991) PNAS 88:9214-9218.) The members of this family, which 25 include members of the  $TGF-\beta$  super-family of proteins, share substantial amino acid sequence homology in their The proteins are translated as a C-terminal regions. precursor, having an N-terminal signal peptide 30 sequence, typically less than about 30 residues, followed by a "pro" domain that is cleaved to yield the mature sequence. The "pro" form of the protein includes the pro domain and the mature domain, and forms a soluble species that appears to be the primary

form secreted from cultured mammalian cells. The signal peptide is cleaved rapidly upon translation, at a cleavage site that can be predicted in a given sequence using the method of Von Heijne ((1986) Nucleic Acids Research 14:4683-4691.) Table I, below, describes the various morphogens identified to date, including their nomenclature as used herein, their Seq. ID references, and publication sources for the amino acid sequences for the full length proteins not included in the Seq. Listing. The disclosure of these publications is incorporated herein by reference.

#### TABLE I

Refers generically to the group of "OP-1" 15 morphogenically active proteins expressed from part or all of a DNA sequence encoding OP-1 protein, including allelic and species variants thereof, e.g., human OP-1 ("hOP-1", Seq. ID No. 5, mature 20 protein amino acid sequence), or mouse OP-1 ("mOP-1", Seq. ID No. 6, mature protein amino acid sequence.) conserved seven cysteine skeleton is defined by residues 38 to 139 of Seq. ID 25 Nos. 5 and 6. The cDNA sequences and the amino acids encoding the full length proteins are provided in Seq. Id Nos. 16 and 17 (hOP1) and Seq. ID Nos. 18 and 19 (mOP1.) The mature proteins are defined 30

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by residues 293-431 (hOP1) and 292-430 (mOP1). The "pro" regions of the proteins, cleaved to yield the mature, morphogenically active proteins are defined essentially by residues 30-292 (hOP1) and residues 30-291 (mOP1).

"OP-2"

refers generically to the group of active proteins expressed from part or all of a DNA sequence encoding OP-2 protein, including allelic and species variants thereof, e.g., human OP-2 ("hOP-2", Seq. ID No. 7, mature protein amino acid sequence) or mouse OP-2 ("mOP-2", Seq. ID No. 8, mature protein amino acid sequence). The conserved seven cysteine skeleton is defined by residues 38 to 139 of Seq. ID Nos. 7 and 8. The cDNA sequences and the amino acids encoding the full length proteins are provided in Seq. ID Nos. 20 and 21 (hOP2) and Seq. ID Nos.

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full length proteins are provided in Seq. ID Nos. 20 and 21 (hOP2) and Seq. ID Nos. 22 and 23 (mOP2.) The mature proteins are defined essentially by residues 264-402 (hOP2) and 261-399 (mOP2). The "pro" regions of the proteins, cleaved to yield the mature, morphogenically active proteins likely are defined essentially by residues 18-263 (hOP2) and residues 18-260 (mOP2). (Another cleavage site also

occurs 21 residues upstream for both OP-2

proteins.)

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refers generically to the morphogenically "CBMP2" active proteins expressed from a DNA sequence encoding the CBMP2 proteins, including allelic and species variants thereof, e.g., human CBMP2A ("CBMP2A(fx)", 5 Seq ID No. 9) or human CBMP2B DNA ("CBMP2B(fx)", Seq. ID No. 10). The amino acid sequence for the full length proteins, referred to in the literature as BMP2A and BMP2B, or BMP2 and BMP4, appear 10 in Wozney, et al. (1988) Science 242:1528-1534. The pro domain for BMP2 (BMP2A) likely includes residues 25-248 or 25-282; the mature protein, residues 249-396 or The pro domain for BMP4 (BMP2B) 15 likely includes residues 25-256 or 25-292; the mature protein, residues 257-408 or 293-408. refers to protein sequences encoded by the "DPP(fx)"

20 "DPP(fx)" refers to protein sequences encoded by the Drosophila DPP gene and defining the conserved seven cysteine skeleton (Seq. ID No. 11). The amino acid sequence for the full length protein appears in Padgett, et al (1987) Nature 325: 81-84. The prodomain likely extends from the signal peptide cleavage site to residue 456; the

residues 457-588.

"Vgl(fx)" refers to protein sequences encoded by the Xenopus Vgl gene and defining the conserved seven cysteine skeleton (Seq. ID No. 12). The amino acid sequence for the

mature protein likely is defined by

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full length protein appears in Weeks (1987) <u>Cell</u> <u>51</u>: 861-867. The prodomain likely extends from the signal peptide cleavage site to residue 246; the mature protein likely is defined by residues 247-360.

"Vgr-1(fx)" refers to protein sequences encoded by the murine Vgr-1 gene and defining the conserved seven cysteine skeleton (Seq. ID No. 13). The amino acid sequence for the full length protein appears in Lyons, et al, (1989) PNAS 86: 4554-4558. The prodomain likely extends from the signal peptide cleavage site to residue 299; the mature protein likely is defined by residues 300-438.

"GDF-1(fx)" refers to protein sequences encoded by the human GDF-1 gene and defining the conserved seven cysteine skeleton (Seq. ID No. 14). The cDNA and encoded amino sequence for the full length protein is provided in Seq. ID. No. 32. The prodomain likely extends from the signal peptide clavage site to residue 214; the mature protein likely is defined by residues 215-372.

refers generically to the morphogenically active proteins expressed from part or all of a DNA sequence (from the Drosophila 60A gene) encoding the 60A proteins (see Seq. ID No. 24 wherein the cDNA and encoded

amino acid sequence for the full length protein is provided). "60A(fx)" refers to the protein sequences defining the conserved seven cysteine skeleton (residues 354 to 455 of Seq. ID No. 24.) The prodomain likely extends from the signal peptide cleavage site to residue 324; the mature protein likely is defined by residues 325-455.

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"BMP3(fx)"

refers to protein sequences encoded by the human BMP3 gene and defining the conserved seven cysteine skeleton (Seq. ID No. 26). The amino acid sequence for the full length protein appears in Wozney et al. (1988) Science 242: 1528-1534. The prodomain likely extends from the signal peptide cleavage site to residue 290; the mature protein likely is defined by

residues 291-472.

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"BMP5(fx)"

refers to protein sequences encoded by the human BMP5 gene and defining the conserved seven cysteine skeleton (Seq. ID No. 27). The amino acid sequence for the full length protein appears in Celeste, et al. (1991) PNAS 87: 9843-9847. The pro domain likely extends from the signal peptide cleavage site to residue 316; the mature protein likely is defined by residues 317-454.

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"BMP6(fx)" refers to protein sequences encoded by the human BMP6 gene and defining the conserved seven cysteine skeleton (Seq. ID No. 28).

The amino acid sequence for the full length protein appears in Celeste, et al.

(1990) PNAS 87: 9843-5847. The pro domain likely includes extends from the signal peptide cleavage site to residue 374; the mature sequence likely includes residues 375-513.

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The OP-2 proteins have an additional cysteine residue in this region (e.g., see residue 41 of Seq. ID Nos. 7 and 8), in addition to the conserved cysteine skeleton in common with the other proteins in this family. The GDF-1 protein has a four amino acid insert within the conserved skeleton (residues 44-47 of Seq. ID No. 14) but this insert likely does not interfere with the relationship of the cysteines in the folded structure. In addition, the CBMP2 proteins are missing one amino acid residue within the cysteine skeleton.

The morphogens are inactive when reduced, but are
active as oxidized homodimers and when oxidized in
combination with other morphogens of this invention.
Thus, as defined herein, a morphogen is a dimeric
protein comprising a pair of polypeptide chains,
wherein each polypeptide chain comprises at least the
C-terminal six cysteine skeleton defined by residues
43-139 of Seq. ID No. 5, including functionally
equivalent arrangements of these cysteines (e.g., amino
acid insertions or deletions which alter the linear
arrangement of the cysteines in the sequence but not

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their relationship in the folded structure), such that, when the polypeptide chains are folded, the dimeric protein species comprising the pair of polypeptide chains has the appropriate three-dimensional structure, 5 including the appropriate intra- or inter-chain disulfide bonds such that the protein is capable of acting as a morphogen as defined herein. Specifically, the morphogens generally are capable of all of the following biological functions in a morphogenically 10 permissive environment: stimulating proliferation of progenitor cells; stimulating the differentiation of progenitor cells; stimulating the proliferation of differentiated cells, and supporting the growth and maintenance of differentiated cells. In addition, it 15 is also anticipated that these morphogens are capable of inducing redifferentiation of committed cells under appropriate environmental conditions.

In one preferred aspect, the morphogens of
this invention comprise one of two species of generic
amino acid sequences: Generic Sequence 1 (Seq. ID
No. 1) or Generic Sequence 2 (Seq. ID No. 2); where
each Xaa indicates one of the 20 naturally-occurring
L-isomer, α-amino acids or a derivative thereof.

Generic Sequence 1 comprises the conserved six cysteine
skeleton and Generic Sequence 2 comprises the conserved
six cysteine skeleton plus the additional cysteine
identified in OP-2 (see residue 36, Seq. ID No. 2). In
another preferred aspect, these sequences further
comprise the following additional sequence at their Nterminus:

Cys Xaa Xaa Xaa Xaa (Seq. ID No. 15)

Preferred amino acid sequences within the foregoing generic sequences include: Generic Sequence 3 (Seq. ID No. 3), Generic Sequence 4 (Seq. ID No. 4), Generic Sequence 5 (Seq. ID No. 30) and Generic Sequence 6 5 (Seq. ID No. 31), listed below. These Generic Sequences accommodate the homologies shared among the various preferred members of this morphogen family identified in Table II, as well as the amino acid sequence variation among them. Specifically, Generic 10 Sequences 3 and 4 are composite amino acid sequences of the following proteins presented in Table II and identified in Seq. ID Nos. 5-14: human OP-1 (hOP-1, Seq. ID Nos. 5 and 16-17), mouse OP-1 (mOP-1, Seq. ID Nos. 6 and 18-19), human and mouse OP-2 (Seq. ID Nos. 7, 8, and 20-22), CBMP2A (Seq. ID No. 9), CBMP2B (Seq. ID No. 10), DPP (from Drosophila, Seq. ID No. 11), Vgl, (from Xenopus, Seq. ID No. 12), Vgr-1 (from mouse, Seq. ID No. 13), and GDF-1 (from mouse, Seq. ID No. 14.) The generic sequences include both 20 the amino acid identity shared by the sequences in Table II, as well as alternative residues for the variable positions within the sequence. Note that these generic sequences allow for an additional cysteine at position 41 or 46 in Generic Sequences 3 or 25 4, respectively, providing an appropriate cysteine skeleton where inter- or intramolecular disulfide bonds can form, and contain certain critical amino acids which influence the tertiary structure of the proteins.

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### Generic Sequence 3

Leu Tyr Val Xaa Phe

Xaa Xaa Xaa Gly Trp Xaa Xaa Trp Xaa

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Xaa Ala Pro Xaa Gly Xaa Xaa Ala

15 20

Xaa Tyr Cys Xaa Gly Xaa Cys Xaa

25 30

10 Xaa Pro Xaa Xaa Xaa Xaa

35

Xaa Xaa Xaa Asn His Ala Xaa Xaa

40 45

Xaa Xaa Leu Xaa Xaa Xaa Xaa

15 50

Xaa Xaa Xaa Xaa Xaa Cys

55 60

Cys Xaa Pro Xaa Xaa Xaa Xaa

65

20 Xaa Xaa Xaa Leu Xaa Xaa Xaa

70 75

Xaa Xaa Xaa Xaa Val Xaa Leu Xaa

80

Xaa Xaa Xaa Met Xaa Val Xaa

25 85 90

### Xaa Cys Gly Cys Xaa

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wherein each Xaa is independently selected from a group of one or more specified amino acids defined as follows: "Res." means "residue" and Xaa at res.4 = (Ser, Asp or Glu); Xaa at res.6 = (Arg, Gln, Ser or Lys); Xaa at res.7 = (Asp or Glu); Xaa at res.8 = (Leu or Val); Xaa at res.11 = (Gln, Leu, Asp, His or Asn); Xaa at res.12 = (Asp, Arg or Asn); Xaa at res.14 = (Ile 10 or Val); Xaa at res.15 = (Ile or Val); Xaa at res.18 = (Glu, Gln, Leu, Lys, Pro or Arg); Xaa at res.20 = (Tyr or Phe); Xaa at res.21 = (Ala, Ser, Asp, Met, His, Leu or Gln); Xaa at res.23 = (Tyr, Asn or Phe); Xaa at res.26 = (Glu, His, Tyr, Asp or Gln); Xaa at res.28 = (Glu, Lys, Asp or Gln); Xaa at res.30 = (Ala, Ser, Pro 15 or Gln); Xaa at res.31 = (Phe, Leu or Tyr); Xaa at res.33 = (Leu or Val); Xaa at res.34 = (Asn, Asp, Ala or Thr); Xaa at res.35 = (Ser, Asp, Glu, Leu or Ala); Xaa at res.36 = (Tyr, Cys, His, Ser or Ile); Xaa at res.37 = (Met, Phe, Gly or Leu); Xaa at res.38 = (Asn 20 or Ser); Xaa at res.39 = (Ala, Ser or Gly); Xaa at res.40 = (Thr, Leu or Ser); Xaa at res.44 = (Ile or Val); Xaa at res.45 = (Val or Leu); Xaa at res.46 = (Gln or Arg); Xaa at res.47 = (Thr, Ala or Ser); Xaa at res.49 = (Val or Met); Xaa at res.50 = (His or Asn); Xaa at res.51 = (Phe, Leu, Asn, Ser, Ala or Val); Xaa at res.52 = (Ile, Met, Asn, Ala or Val); Xaa at res.53 = (Asn, Lys, Ala or Glu); Xaa at res.54 = (Pro or Ser); Xaa at res.55 = (Glu, Asp, Asn, or Gly); Xaa at res.56 30 = (Thr, Ala, Val, Lys, Asp, Tyr, Ser or Ala); Xaa at res.57 = (Val, Ala or Ile); Xaa at res.58 = (Pro or Asp); Xaa at res.59 = (Lys or Leu); Xaa at res.60 = (Pro or Ala); Xaa at res.63 = (Ala or Val); Xaa at

res.65 = (Thr or Ala); Xaa at res.66 = (Gln, Lys, Arg or Glu); Xaa at res.67 = (Leu, Met or Val); Xaa at res.68 = (Asn, Ser or Asp); Xaa at res.69 = (Ala, Pro or Ser); Xaa at res.70 = (Ile, Thr or Val); Xaa at 5 res.71 = (Ser or Ala); Xaa at res.72 = (Val or Met); Xaa at res.74 = (Tyr or Phe); Xaa at res.75 = (Phe, Tyr or Leu); Xaa at res.76 = (Asp or Asn); Xaa at res.77 = (Asp, Glu, Asn or Ser); Xaa at res.78 = (Ser, Gln, Asn or Tyr); Xaa at res.79 = (Ser, Asn, Asp or Glu); Xaa at 10 res.80 = (Asn, Thr or Lys); Xaa at res.82 = (Ile or Val); Xaa at res.84 = (Lys or Arg); Xaa at res.85 = (Lys, Asn, Gln or His); Xaa at res.86 = (Tyr or His); Xaa at res.87 = (Arg, Gln or Glu); Xaa at res.88 = (Asn, Glu or Asp); Xaa at res.90 = (Val, Thr or Ala); Xaa at res.92 = (Arg, Lys, Val, Asp or Glu); Xaa at res.93 = (Ala, Gly or Glu); and Xaa at res.97 = (His or Arg);

### Generic Sequence 4

20

25

 Cys
 Xaa
 Xaa
 Xaa
 Leu
 Tyr
 Val
 Xaa
 Phe

 1
 5
 5
 10

 Xaa
 Xaa
 Gly
 Trp
 Xaa
 Trp
 Xaa

 Xaa
 Ala
 25
 25

 Xaa
 Tyr
 Cys
 Xaa
 Gly
 Xaa
 Cys
 Xaa

 Xaa
 Pro
 Xaa
 Xaa
 Xaa
 Xaa
 Xaa

30 40

5

10

 Xaa
 Xaa
 Asn
 His
 Ala
 Xaa
 Xaa
 50

 Xaa
 Xaa
 Leu
 Xaa
 Xaa
 Xaa
 Xaa
 Xaa
 Xaa
 Xaa
 Cys

 Xaa
 Xaa
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 Xaa
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15 Xaa Cys Gly Cys Xaa 100

wherein each Xaa is independently selected from a group of one or more specified amino acids as defined by the following: "Res." means "residue" and Xaa at res.2 = (Lys or Arg); Xaa at res.3 = (Lys or Arg); Xaa at res.4 20 = (His or Arg); Xaa at res.5 = (Glu, Ser, His, Gly, Arg or Pro); Xaa at res.9 = (Ser, Asp or Glu); Xaa at res.11 = (Arg, Gln, Ser or Lys); Xaa at res.12 = (Asp or Glu); Xaa at res.13 = (Leu or Val); Xaa at res.16 = (Gln, Leu, Asp, His or Asn); Xaa at res.17 = (Asp, Arg, 25 or Asn); Xaa at res.19 = (Ile or Val); Xaa at res.20 = (Ile or Val); Xaa at res.23 = (Glu, Gln, Leu, Lys, Pro or Arg); Xaa at res.25 = (Tyr or Phe); Xaa at res.26 = (Ala, Ser, Asp, Met, His, Leu, or Gln); Xaa at res.28 = (Tyr, Asn or Phe); Xaa at res.31 = (Glu, His, Tyr, Asp 30 or Gln); Xaa at res.33 = Glu, Lys, Asp or Gln); Xaa at res.35 = (Ala, Ser or Pro); Xaa at res.36 = (Phe, Leu or Tyr); Xaa at res.38 = (Leu or Val); Xaa at res.39 = (Asn, Asp, Ala or Thr); Xaa at res.40 = (Ser, Asp, Glu,

Leu or Ala); Xaa at res.41 = (Tyr, Cys, His, Ser or Ile); Xaa at res.42 = (Met, Phe, Gly or Leu); Xaa at res.44 = (Ala, Ser or Gly); Xaa at res.45 = (Thr, Leu or Ser); Xaa at res.49 = (Ile or Val); Xaa at res.50 = 5 (Val or Leu); Xaa at res.51 = (Gln or Arg); Xaa at res.52 = (Thr, Ala or Ser); Xaa at res.54 = (Val or Met); Xaa at res.55 = (His or Asn); Xaa at res.56 = (Phe, Leu, Asn, Ser, Ala or Val); Xaa at res.57 = (Ile, Met, Asn, Ala or Val); Xaa at res.58 = (Asn, Lys, Ala 10 or Glu); Xaa at res.59 = (Pro or Ser); Xaa at res.60 = (Glu, Asp, or Gly); Xaa at res.61 = (Thr, Ala, Val, Lys, Asp, Tyr, Ser or Ala); Xaa at res.62 = (Val, Ala or Ile); Xaa at res.63 = (Pro or Asp); Xaa at res.64 = (Lys or Leu); Xaa at res.65 = (Pro or Ala); Xaa at 15 res.68 = (Ala or Val); Xaa at res.70 = (Thr or Ala); Xaa at res.71 = (Gln, Lys, Arg or Glu); Xaa at res.72 = (Leu, Met or Val); Xaa at res.73 = (Asn, Ser or Asp); Xaa at res.74 = (Ala, Pro or Ser); Xaa at res.75 = (Ile, Thr or Val); Xaa at res.76 = (Ser or Ala); Xaa at 20 res.77 = (Val or Met); Xaa at res.79 = (Tyr or Phe); Xaa at res.80 = (Phe, Tyr or Leu); Xaa at res.81 = (Asp or Asn); Xaa at res.82 = (Asp, Glu, Asn or Ser); Xaa at res.83 = (Ser, Gln, Asn or Tyr); Xaa at res.84 = (Ser, Asn, Asp or Glu); Xaa at res.85 = (Asn, Thr or Lys); Xaa at res.87 = (Ile or Val); Xaa at res.89 = (Lys or 25 Arg); Xaa at res.90 = (Lys, Asn, Gln or His); Xaa at res.91 = (Tyr or His); Xaa at res.92 = (Arg, Gln or Glu); Xaa at res.93 = (Asn, Glu or Asp); Xaa at res.95 = (Val, Thr or Ala); Xaa at res.97 = (Arg, Lys, Val, 30 Asp or Glu); Xaa at res.98 = (Ala, Gly or Glu); and Xaa at res. 102 = (His or Arg).

Similarly, Generic Sequence 5 (Seq. ID No. 30) and Generic Sequence 6 (Seq. ID No. 31) accommodate the homologies shared among all the morphogen protein family members identified in Table II. Specifically, 5 Generic Sequences 5 and 6 are composite amino acid sequences of human OP-1 (hOP-1, Seq. ID Nos. 5 and 16-17), mouse OP-1 (mOP-1, Seq. ID Nos. 6 and 18-19), human and mouse OP-2 (Seq. ID Nos. 7, 8, and 20-22), CBMP2A (Seq. ID No. 9), CBMP2B (Seq. ID No. 10), DPP 10 (from Drosophila, Seq. ID No. 11), Vgl, (from Xenopus, Seq. ID No. 12), Vgr-1 (from mouse, Seq. ID No. 13), and GDF-1 (from mouse, Seq. ID No. 14), human BMP3 (Seq. ID No. 26), human BMP5 (Seq. ID No. 27), human BMP6 (Seq. ID No. 28) and 60(A) (from Drosophila, Seq. 15 ID Nos. 24-25). The generic sequences include both the amino acid identity shared by these sequences in the C-terminal domain, defined by the six and seven cysteine skeletons (Generic Sequences 5 and 6, respectively), as well as alternative residues for the 20 variable positions within the sequence. As for Generic Sequences 3 and 4, Generic Sequences 5 and 6 allow for an additional cysteine at position 41 (Generic Sequence 5) or position 46 (Generic Sequence 6), providing an appropriate cysteine skeleton where inter- or 25 intramolecular disulfide bonds can form, and containing certain critical amino acids which influence the tertiary structure of the proteins.

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## Generic Sequence 5

_				<b>51</b>
T.em	Xaa	Xaa	xaa	Pne

1 5

5 Xaa Xaa Xaa Gly Trp Xaa Xaa Trp Xaa

10

Xaa Xaa Pro Xaa Xaa Xaa Ala

15 20

Xaa Tyr Cys Xaa Gly Xaa Cys Xaa

10 25 30

Xaa Pro Xaa Xaa Xaa Xaa

35

Xaa Xaa Xaa Asn His Ala Xaa Xaa

40 45

15 Xaa Xaa Xaa Xaa Xaa Xaa Xaa

50

Xaa Xaa Xaa Xaa Xaa Xaa Cys

55 60

Cys Xaa Pro Xaa Xaa Xaa Xaa

20 65

Xaa Xaa Xaa Leu Xaa Xaa Xaa

70 75

Xaa Xaa Xaa Xaa Val Xaa Leu Xaa

80

Xaa Xaa Xaa Xaa Met Xaa Val Xaa

85 90

5 . Xaa Cys Xaa Cys Xaa

95

wherein each Xaa is independently selected from a group of one or more specified amino acids defined as follows: "Res." means "residue" and Xaa at res.2 = (Tyr or Lys); Xaa at res.3 = Val or Ile); Xaa at res.4 = (Ser, Asp or Glu); Xaa at res.6 = (Arg, Gln, Ser, Lys or Ala); Xaa at res.7 = (Asp, Glu or Lys); Xaa at res.8 = (Leu, Val or Ile); Xaa at res.11 = (Gln, Leu, Asp, His, Asn or Ser); Xaa at res.12 = (Asp, Arg, Asn or 15 Glu); Xaa at res.14 = (Ile or Val); Xaa at res.15 = (Ile or Val); Xaa at res.16 (Ala or Ser); Xaa at res.18 = (Glu, Gln, Leu, Lys, Pro or Arg); Xaa at res.19 = (Gly or Ser); Xaa at res.20 = (Tyr or Phe); Xaa at res.21 = (Ala, Ser, Asp, Met, His, Gln, Leu or Gly); 20 Xaa at res.23 = (Tyr, Asn or Phe); Xaa at res.26 = (Glu, His, Tyr, Asp, Gln or Ser); Xaa at res.28 = (Glu, Lys, Asp, Gln or Ala); Xaa at res.30 = (Ala, Ser, Pro, Gln or Asn); Xaa at res.31 = (Phe, Leu or Tyr); Xaa at res.33 = (Leu, Val or Met); Xaa at res.34 = (Asn, Asp, Ala, Thr or Pro); Xaa at res.35 = (Ser, Asp, Glu, Leu, 25 Ala or Lys); Xaa at res.36 = (Tyr, Cys, His, Ser or Ile); Xaa at res.37 = (Met, Phe, Gly or Leu); Xaa at res.38 = (Asn, Ser or Lys); Xaa at res.39 = (Ala, Ser, Gly or Pro); Xaa at res.40 = (Thr, Leu or Ser); Xaa at 30 res.44 = (Ile, Val or Thr); Xaa at res.45 = (Val, Leu or Ile); Xaa at res.46 = (Gln or Arg); Xaa at res.47 = (Thr, Ala or Ser); Xaa at res.48 = (Leu or Ile); Xaa at

res.49 = (Val or Met); Xaa at res.50 = (His, Asn or Arg); Xaa at res.51 = (Phe, Leu, Asn, Ser, Ala or Val); Xaa at res.52 = (Ile, Met, Asn, Ala, Val or Leu); Xaa at res.53 = (Asn, Lys, Ala, Glu, Gly or Phe); Xaa at 5 res.54 = (Pro, Ser or Val); Xaa at res.55 = (Glu, Asp, Asn, Gly, Val or Lys); Xaa at res.56 = (Thr, Ala, Val, Lys, Asp, Tyr, Ser, Ala, Pro or His); Xaa at res.57 = (Val, Ala or Ile); Xaa at res.58 = (Pro or Asp); Xaa at res.59 = (Lys, Leu or Glu); Xaa at res.60 = (Pro or 10 Ala); Xaa at res.63 = (Ala or Val); Xaa at res.65 = (Thr, Ala or Glu); Xaa at res.66 = (Gln, Lys, Arg or Glu); Xaa at res.67 = (Leu, Met or Val); Xaa at res.68 = (Asn, Ser, Asp or Gly); Xaa at res.69 = (Ala, Pro or Ser); Xaa at res.70 = (Ile, Thr, Val or Leu); Xaa at 15 res.71 = (Ser, Ala or Pro); Xaa at res.72 = (Val, Met or Ile); Xaa at res.74 = (Tyr or Phe); Xaa at res.75 = (Phe, Tyr, Leu or His); Xaa at res.76 = (Asp, Asn or Leu); Xaa at res.77 = (Asp, Glu, Asn or Ser); Xaa at res.78 = (Ser, Gln, Asn, Tyr or Asp); Xaa at res.79 = (Ser, Asn, Asp, Glu or Lys); Xaa at res.80 = (Asn, Thr 20 or Lys); Xaa at res.82 = (Ile, Val or Asn); Xaa at res.84 = (Lys or Arg); Xaa at res.85 = (Lys, Asn, Gln, His or Val); Xaa at res.86 = (Tyr or His); Xaa at res.87 = (Arg, Gln, Glu or Pro); Xaa at res.88 = (Asn, 25 Glu or Asp); Xaa at res.90 = (Val, Thr, Ala or Ile); Xaa at res.92 = (Arg, Lys, Val, Asp or Glu); Xaa at res.93 = (Ala, Gly, Glu or Ser); Xaa at res.95 = (Gly or Ala) and Xaa at res.97 = (His or Arg).

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# Generic Sequence 6

	_	Xaa	Xaa	Xaa		Leu	Xaa	Xaa	Xaa	
	1				5					10
5	Xaa	Xaa	Xaa	Gly	Trp	Xaa	Xaa	Trp	Xaa	
					15					
	Xaa	Xaa	Pro	Xaa	Xaa	Xaa	Xaa	Ala		
	20					25				
	Xaa	Tyr	Cys	Xaa	Gly	Xaa	Cys	Xaa		
10			30					35		
	Xaa	Pro	Xaa	Xaa	Xaa	Xaa	Xaa			
					40					
	Xaa	Xaa	Xaa	Asn	His	Ala	Xaa	Xaa		
			45					50		
15	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa		
					55					
	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Cys		
		60					65			
	Cys	Xaa	Pro	Xaa	Xaa	Xaa	Xaa	Xaa		
20				70						
	Xaa	Xaa	Xaa	Leu	Xaa	Xaa	Xaa			
	75					80				
	Xaa	Xaa	Xaa	Xaa	Val	Xaa	Leu	Xaa		
				85						
25	Xaa	Xaa	Xaa		Met	Xaa	Val	Xaa		
	90					95	•			
		Cys	Xaa	Cvs	Xaa					
		~ <sub>J</sub> 5	100	-10						

wherein each Xaa is independently selected from a group of one or more specified amino acids as defined by the following: "Res." means "residue" and Xaa at res.2 = (Lys, Arg, Ala or Gln); Xaa at res.3 = (Lys, Arg or

Met); Xaa at res.4 = (His, Arg or Gln); Xaa at res.5 = (Glu, Ser, His, Gly, Arg, Pro, Thr, or Tyr); Xaa at res.7 = (Tyr or Lys); Xaa at res.8 = (Val or Ile); Xaa at res.9 = (Ser, Asp or Glu); Xaa at res.11 = (Arg, 5 Gln, Ser, Lys or Ala); Xaa at res.12 = (Asp, Glu, or Lys); Xaa at res.13 = (Leu, Val or Ile); Xaa at res.16 = (Gln, Leu, Asp, His, Asn or Ser); Xaa at res.17 = (Asp, Arg, Asn or Glu); Xaa at res.19 = (Ile or Val); Xaa at res.20 = (Ile or Val); Xaa at res.21 = (Ala or 10 Ser); Xaa at res.23 = (Glu, Gln, Leu, Lys, Pro or Arg); Xaa at res.24 = (Gly or Ser); Xaa at res.25 = (Tyr or Phe); Xaa at res.26 = (Ala, Ser, Asp, Met, His, Gln, Leu, or Gly); Xaa at res.28 = (Tyr, Asn or Phe); Xaa at res.31 = (Glu, His, Tyr, Asp, Gln or Ser); Xaa at 15 res.33 = Glu, Lys, Asp, Gln or Ala); Xaa at res.35 = (Ala, Ser, Pro, Gln or Asn); Xaa at res.36 = (Phe, Leu or Tyr); Xaa at res.38 = (Leu, Val or Met); Xaa at res.39 = (Asn, Asp, Ala, Thr or Pro); Xaa at res.40 = (Ser, Asp, Glu, Leu, Ala or Lys); Xaa at res.41 = (Tyr, 20 Cys, His, Ser or Ile); Xaa at res.42 = (Met, Phe, Gly or Leu); Xaa at res.43 = (Asn, Ser or Lys); Xaa at res.44 = (Ala, Ser, Gly or Pro); Xaa at res.45 = (Thr, Leu or Ser); Xaa at res.49 = (Ile, Val or Thr); Xaa at res.50 = (Val, Leu or Ile); Xaa at res.51 = (Gln or Arg); Xaa at res.52 = (Thr, Ala or Ser); Xaa at res.53 = (Leu or Ile); Xaa at res.54 = (Val or Met); Xaa at res.55 = (His, Asn or Arg); Xaa at res.56 = (Phe, Leu, Asn, Ser, Ala or Val); Xaa at res.57 = (Ile, Met, Asn, Ala, Val or Leu); Xaa at res.58 = (Asn, Lys, Ala, Glu, 30 Gly or Phe); Xaa at res.59 = (Pro, Ser or Val); Xaa at res.60 = (Glu, Asp, Gly, Val or Lys); Xaa at res.61 = (Thr, Ala, Val, Lys, Asp, Tyr, Ser, Ala, Pro or His); Xaa at res.62 = (Val, Ala or Ile); Xaa at res.63 = (Pro or Asp); Xaa at res.64 = (Lys, Leu or Glu); Xaa at

res.65 = (Pro or Ala); Xaa at res.68 = (Ala or Val); Xaa at res.70 = (Thr, Ala or Glu); Xaa at res.71 = (Gln, Lys, Arg or Glu); Xaa at res.72 = (Leu, Met or Val); Xaa at res.73 = (Asn, Ser, Asp or Gly); Xaa at 5 res.74 = (Ala, Pro or Ser); Xaa at res.75 = (Ile, Thr, Val or Leu); Xaa at res.76 = (Ser, Ala or Pro); Xaa at res.77 = (Val, Met or Ile); Xaa at res.79 = (Tyr or Phe); Xaa at res.80 = (Phe, Tyr, Leu or His); Xaa at res.81 = (Asp, Asn or Leu); Xaa at res.82 = (Asp, Glu, 10 Asn or Ser); Xaa at res.83 = (Ser, Gln, Asn, Tyr or Asp); Xaa at res.84 = (Ser, Asn, Asp, Glu or Lys); Xaa at res.85 = (Asn, Thr or Lys); Xaa at res.87 = (Ile, Val or Asn); Xaa at res.89 = (Lys or Arg); Xaa at res.90 = (Lys, Asn, Gln, His or Val); Xaa at res.91 = (Tyr or His); Xaa at res.92 = (Arg, Gln, Glu or Pro); 15 Xaa at res.93 = (Asn, Glu or Asp); Xaa at res.95 = (Val, Thr, Ala or Ile); Xaa at res.97 = (Arg, Lys, Val, Asp or Glu); Xaa at res.98 = (Ala, Gly, Glu or Ser); Xaa at res.100 = (Gly or Ala); and Xaa at res.102 = (His or Arg). 20

Particularly useful sequences for use as morphogens in this invention include the C-terminal domains, e.g., the C-terminal 96-102 amino acid residues of Vgl,

25 Vgr-1, DPP, OP-1, OP-2, CBMP-2A, CBMP-2B, GDF-1 (see Table II, below, and Seq. ID Nos. 5-14), as well as proteins comprising the C-terminal domains of 60A, BMP3, BMP5 and BMP6 (see Seq. ID Nos. 24-28), all of which include at least the conserved six or seven cysteine skeleton. In addition, biosynthetic

constructs designed from the generic sequences, such as COP-1, 3-5, 7, 16, disclosed in U.S. Pat. No. 5,011,691, also are useful. Other sequences include the inhibins/activin proteins (see, for example, U.S. 5 Pat. Nos. 4,968,590 and 5,011,691). Accordingly, other useful sequences are those sharing at least 70% amino acid sequence homology or "similarity", and preferably 80% homology or similarity with any of the sequences above. These are anticipated to include allelic, species variants and other sequence variants (e.g., including "muteins" or "mutant proteins"), whether naturally-occurring or biosynthetically produced, as well as novel members of this morphogenic family of proteins. As used herein, "amino acid sequence 15 homology" is understood to mean amino acid sequence similarity, and homologous sequences share identical or similar amino acids, where similar amino acids are conserved amino acids as defined by Dayoff et al., Atlas of Protein Sequence and Structure; vol.5, 20 Suppl.3, pp.345-362 (M.O. Dayoff, ed., Nat'l BioMed. Research Fdn., Washington D.C. 1978.) Thus, a candidate sequence sharing 70% amino acid homology with a reference sequence requires that, following alignment of the candidate sequence with the reference sequence, 70% of the amino acids in the candidate sequence are 25 identical to the corresponding amino acid in the reference sequence, or constitute a conserved amino acid change thereto. "Amino acid sequence identity" is understood to require identical amino acids between two aligned sequences. Thus, a candidate sequence sharing 30

60% amino acid identity with a reference sequence requires that, following alignment of the candidate sequence with the reference sequence, 60% of the amino acids in the candidate sequence are identical to the corresponding amino acid in the reference sequence.

As used herein, all homologies and identities calculated use OP-1 as the reference sequence. Also as used herein, sequences are aligned for homology and identity calculations using the method of Needleman et al. (1970) <u>J.Mol. Biol.</u> 48:443-453 and identities calculated by the Align program (DNAstar, Inc.) In all cases, internal gaps and amino acid insertions in the candidate sequence as aligned are ignored when making the homology/identity calculation.

The currently most preferred protein sequences useful as morphogens in this invention include those having greater than 60% identity, preferably greater 20 than 65% identity, with the amino acid sequence defining the conserved six cysteine skeleton of hOP1 (e.g., residues 43-139 of Seq. ID No. 5). These most preferred sequences include both allelic and species variants of the OP-1 and OP-2 proteins, including the 25 Drosophila 60A protein. Accordingly, in another preferred aspect of the invention, useful morphogens include active proteins comprising species of polypeptide chains having the generic amino acid sequence herein referred to as "OPX", which accommodates the homologies between the various 30 identified species of OP1 and OP2 (Seq. ID No. 29).

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In still another preferred aspect of the invention, useful morphogens include dimeric proteins comprising amino acid sequences encoded by nucleic acids that hybridize to DNA or RNA sequences encoding the C
5 terminal sequences defining the conserved seven cysteine domain of OP1 or OP2, e.g., nucleotides 10361341 and nucleotides 1390-1695 of Seq. ID No. 16 and 20, respectively, under stringent hybridization conditions. As used herein, stringent hybridization 10 conditions are defined as hybridization in 40% formamide, 5 X SSPE, 5 X Denhardt's Solution, and 0.1% SDS at 37°C overnight, and washing in 0.1 X SSPE, 0.1% SDS at 50°C.

The morphogens useful in the methods, composition 15 and devices of this invention include proteins comprising any of the polypeptide chains described above, whether isolated from naturally-occurring sources, or produced by recombinant DNA or other synthetic techniques, and includes allelic and species variants of these proteins, naturally-occurring or biosynthetic mutants thereof, as well as various truncated and fusion constructs. Deletion or addition mutants also are envisioned to be active, including 25 those which may alter the conserved C-terminal cysteine skeleton, provided that the alteration does not functionally disrupt the relationship of these cysteines in the folded structure. Accordingly, such active forms are considered the equivalent of the 30 specifically described constructs disclosed herein.

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The proteins may include forms having varying glycosylation patterns, varying N-termini, a family of related proteins having regions of amino acid sequence homology, and active truncated or mutated forms of native or biosynthetic proteins, produced by expression of recombinant DNA in host cells.

The morphogenic proteins can be expressed from intact or truncated cDNA or from synthetic DNAs in procaryotic or eucaryotic host cells, and purified, cleaved, refolded, and dimerized to form morphogenically active compositions. Currently preferred host cells include <a href="Eccolic">E. colic</a> or mammalian cells, such as CHO, COS or BSC cells. A detailed description of the morphogens useful in the methods, compositions and devices of this invention is disclosed in international application US92/01968 (WO 92/15323) the disclosure of which is incorporated herein by reference.

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Thus, in view of this disclosure, skilled genetic engineers can isolate genes from cDNA or genomic libraries of various different species which encode appropriate amino acid sequences, or construct DNAs from oligonucleotides, and then can express them in various types of host cells, including both procaryotes and eucaryotes, to produce large quantities of active proteins useful as dietary compositions for enhancing tissue morphogenesis, including enhancing tissue development and tisssue viability in a variety of mammals, including humans.

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The foregoing and other objects, features and advantages of the present invention will be made more apparent from the following detailed description of the invention.

### Brief Description of the Drawings

The foregoing and other objects and features of
this invention, as well as the invention itself, may be
more fully understood from the following description,
when read together with the accompanying drawings, in
which:

present in mammary gland extract eluate fractions of a C-18 reverse phase chromatography column (A), and the corresponding results of a Western Blot (B);

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- FIG. 2A and B shows relative amounts of protein present in bovine colostrum eluate fractions from purification scheme A of a C-18 reverse phase chromatography column (A), and the corresponding results of a Western blot under reduced (1) and oxidized (2) conditions (B);
- present in bovine colostrum eluate fractions from
  purification scheme B of a C-18 reverse phase
  chromatography column (A), and the corresponding
  results of a Western Blot under reduced conditions (B);
- FIG. 4A and B shows relative amounts of protein
  present in bovine 57 day milk eluate fractions of a C18 reverse phase chromatography column (A), and the
  corresponding results of a Western Blot under reduced
  (1) and oxidized (2) conditions (B);

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- FIG. 5 shows Western Blot analysis of bovine colostrum using OP-1 and BMP2-specific antibodies;
- FIG. 6A and B show results of <u>in vivo</u> and <u>in vitro</u>

  5 activity assays, respectively, for the corresponding fractions shown in Fig. 1;
  - FIG. 7 is a photomicrograph of an immunoblot showing the presence of hOP-1 in serum; and
- FIG. 8A is a dose response curve for the induction of the 180 kDa and 140 kDa N-CAM isoforms in morphogentreated NG108-15 cells;
- 15 FIG. 8B is a photomicrograph of a Western blot of whole cell extracts from morphogen-treated NG108-15 cells with an N-CAM-specific antibody; and
- FIG. 9 (A and B) are photomicrographs showing the effect of morphogen-specific antibody on mouse development (9B) compared to untreated, control mice (9A).

#### Detailed Description of the Invention

It now has been discovered that the proteins

described herein are found in nursing mother's milk and are useful as components of a dietary composition for enhancing tissue morphogenesis in a mammal, particularly in an individual at risk for normal tissue development and vibility. As described herein, these proteins ("morphogens") are capable of enhancing tissue development in growing mammals, stimulating CAM expression and maintaining the normal tissue function in adult tissue.

Provided below are detailed descriptions of 15 suitable morphogens useful in the compositions and methods of this invention, as well as methods for their administration and application, and numerous, nonlimiting examples which demonstrate the suitability 20 of the morphogens described herein as active components of a dietary composition for a mammal; and 2) provide assays with which to test candidate morphogens for Specifically, examples are provided their efficacy. which (1) demonstrate the presence of endogenous 25 morphogen in milk and human serum (Examples 1 and 2), (2) demonstrate the ability of morphogens to induce CAM expression in a mammal (Example 3), (3) demonstrate the ability of morphogens to enhance tissue development in developing embryos (Example 4) and juveniles 30 (Example 5); (4) demonstrate the ability of morphogens to reduce an osteoporotic condition in a mammal (Example 6); (5) demonstrate the presence of morphogens in developing tissues and adult stomach and gut tissue, demonstrate the ability of parenterally provided

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morphogen to localize to stomach tissue, and describe protocols for identifying morphogen-synthesizing tissue (Example 7) and (6) describe protocols for obtaining morphogen-specific antibodies and measuring morphogens in solution (Example 8).

#### I. Useful Morphogens

WO 94/03075

As defined herein a protein is morphogenic if it is 10 capable of inducing the developmental cascade of cellular and molecular events that culminate in the formation of new, organ-specific tissue and comprises at least the conserved C-terminal six cysteine skeleton or its functional equivalent (see supra). Specifically, the morphogens generally are capable of 15 all of the following biological functions in a morphogenically permissive environment: stimulating proliferation of progenitor cells; stimulating the differentiation of progenitor cells; stimulating the 20 proliferation of differentiated cells; and supporting the growth and maintenance of differentiated cells. Details of how the morphogens useful in the method of this invention first were identified, as well as a description on how to make, use and test them for 25 morphogenic activity are disclosed in international application US92/01968 (WO 92/15323). As disclosed therein, the morphogens may be purified from naturallysourced material or recombinantly produced from procaryotic or eucaryotic host cells, using the genetic sequences disclosed therein. Alternatively, novel 30 morphogenic sequences may be identified following the procedures disclosed therein.

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Particularly useful proteins include those which comprise the naturally derived sequences disclosed in Table II. Other useful sequences include biosynthetic constructs such as those disclosed in U.S. Pat. 5,011,691, the disclosure of which is incorporated herein by reference (e.g., COP-1, COP-3, COP-4, COP-5, COP-7, and COP-16).

Accordingly, the morphogens useful in the methods
and compositions of this invention also may be
described by morphogenically active proteins having
amino acid sequences sharing 70% or, preferably, 80%
homology (similarity) with any of the sequences
described above, where "homology" is as defined herein
above.

The morphogens useful in the method of this invention also can be described by any of the 6 generic sequences described herein (Generic Sequences 1, 2, 3, 4, 5 and 6). Generic sequences 1 and 2 also may include, at their N-terminus, the sequence

Cys Xaa Xaa Xaa Xaa (Seq. ID No. 15)
1 5

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Table II, set forth below, compares the amino acid sequences of the active regions of native proteins that have been identified as morphogens, including human OP-1 (hOP-1, Seq. ID Nos. 5 and 16-17), mouse OP-1 (mOP-1, Seq. ID Nos. 6 and 18-19), human and mouse OP-2 (Seq. ID Nos. 7, 8, and 20-23), CBMP2A (Seq. ID No. 9), CBMP2B (Seq. ID No. 10), BMP3 (Seq. ID No. 26), DPP (from Drosophila, Seq. ID No. 11), Vgl, (from Xenopus, Seq. ID No. 12), Vgr-1 (from mouse, Seq. ID No. 13),

GDF-1 (from mouse, Seq. ID Nos. 14, 32 and 33), 60A protein (from Drosophila, Seq. ID Nos. 24 and 25), BMP5 (Seq. ID No. 27) and BMP6 (Seq. ID No. 28). sequences are aligned essentially following the method 5 of Needleman et al. (1970) <u>J. Mol. Biol.</u>, <u>48</u>:443-453, calculated using the Align Program (DNAstar, Inc.) In the table, three dots indicates that the amino acid in that position is the same as the amino acid in hOP-1. Three dashes indicates that no amino acid is present in 10 that position, and are included for purposes of illustrating homologies. For example, amino acid residue 60 of CBMP-2A and CBMP-2B is "missing". Of course, both these amino acid sequences in this region comprise Asn-Ser (residues 58, 59), with CBMP-2A then 15 comprising Lys and Ile, whereas CBMP-2B comprises Ser and Ile.

#### TABLE II

20									
	hOP-1	Cys	Lys	Lys	His	Glu	Leu	Tyr	Val
	mOP-1	• • •	• • •	• • •	• • •	•••	• • •	• • •	• • •
	hOP-2	• • •	Arg	Arg	•••	• • •	•••	• • •	• • •
	mOP-2	• • •	Arg	Arg	• • •	• • •	• • •	• • •	• • •
25	DPP	• • •	Arg	Arg	• • •	Ser	•••	• • •	• • •
	Vgl	•••	• • •	Lys	Arg	His	• • •	• • •	• • •
	Vgr-1	• • •	• • •	•••	• • •	Gly	• • •	•••	• • •
	CBMP-2A	• • •	• • •	Arg	• • •	Pro	•••	• • •	• • •
	CBMP-2B	• • •	Arg	Arg	• • •	Ser	• • •	• • •	• • •

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	BMP3	• • •	Ala	Arg	Arg	Tyr	• • •	Lys	• • •	
	GDF-1	• • •	Arg	Ala	Arg	Arg	• • •	• • •	• • •	
	60A	•••	Gln	Met	Glu	Thr	• • •	• • •	• • •	
	BMP5	• • •	• • •	• • •	• • •	• • •	• • •	• • •	• • •	
5	BMP6	• • •	Arg	• • •	•••	• • •	• • •	• • •	• • •	
		1				5				
	hOP-1	Ser	Phe	Arg	Asp	Leu	Gly	Trp	Gln	Asp
10	mOP-1	•••	• • •	• • •	• • •	• • •	• • •	• • •	• • •	• • •
	hOP-2	• • •		Gln	• • •	• • •	• • •	• • •	Leu	• • •
	mOP-2	Ser	• • •	•••	• • •	• • •	• • •	• • •	Leu	• • •
	DPP	Asp	• • •	Ser	• • •	Val	• • •	• • •	Asp	• • •
	Vgl	Glu	• • •	Lys	• • •	Val	• • •	• • •	• • •	Asn
15	Vgr-1	• • •	• • •	Gln	•••	Val	•••	• • •	• • •	• • •
	CBMP-2A	Asp	•••	Ser	• • •	Val	•••	• • •	Asn	• • •
	CBMP-2B	Asp	•••	Ser	• • •	Val	• • •	• • •	Asn	• • •
	BMP3	Asp	•••	Ala	• • •	Ile	• • •	• • •	Ser	Glu
	GDF-1	• • •	• • •	• • • .	Glu	Val	• • •	• • •	His	Arg
20	60A	Asp	• • •	Lys	• • •	• • •	•••	• • •	His	• • •
	BMP5	• • •		• • •	• • •	• • •	• • •	• • •	• • •	• • •
	BMP6	• • •	• • •	${\tt Gln}$	• • •	• • •	• • •	• • •	• • • •	• • •
			10					15		
25	hOP-1	Trp	Ile	Ile	Ala	Pro	Glu	Gly	Tyr	Ala
	mOP-1	• • •	• • •	•••	• • •	• • •	• • •	• • •	• • •	
	hOP-2	• • •	Val	• • •	• • •	• • •	Gln	• • •	• • •	Ser
	mOP-2	• • •	Val	• • •	• • •		Gln	• • •	•••	Ser
	DPP	• • •	•••	Val	• • •	• • •	Leu	• • •	• • •	Asp
30	Vgl	• • •	Val	• • •	• • •	• • •	Gln	• • •	• • •	Met
	Vgr-1	• • •		• • •		• • •	Lys	• • •	• • •	• • •
	CBMP-2A	• • •	• • •	Val	• • •		Pro	• • •	• • •	His

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	CBMP-2B	• • •	•••	Val	• • •	• • •	Pro	• • •	• • •	Gln
	BMP3	• • •	• • •	• • •	Ser	•••	Lys	Ser	Phe	Asp
	GDF-1	• • •	Val	• • •	• • •	• • •	Arg	• • •	Phe	Leu
	60A	• • •	• • •	• • •	• • •	• • •	•••	•••	• • •	Gly
5	BMP5	•••	• • •	•••	• • •	• • •	• • •	• • •	• • •	
	BMP6	• • •	• • •	• • •	• • •	•••	Lys	• • •	• • •	• • •
				20					25	
10	hOP-1	Ala	Tyr	Tyr	Cys	Glu	Gly	Glu	Cys	Ala
	mOP-1	•••	• • •	•••	• • •	• • •	• • •	• • •	• • •	• • •
	hOP-2	•••	• • •	• • •	• • •	• • •	• • •	• • •	• • •	Ser
	mOP-2	•••	• • •	•••	•••	• • •	• • •	• • •	• • •	• • •
	DPP	• • •	• • •	• • •		His	• • •	Lys	• • •	Pro
15	Vgl	• • •	Asn	• • •	• • •	Tyr	• • •	• • •		Pro
	Vgr-1	•••	Asn	• • •	• • •	Asp	• • •	• • •	• • •	Ser
	CBMP-2A	• • •	Phe	• • •	• • •	His	• • •	Glu		Pro
	CBMP-2B	• • •	Phe	• • •	• • •	His	• • •	Asp	• • •	Pro
	BMP3	•••	• • •	• • •	• • •	Ser	• • •	Ala		Gln
20	GDF-1	•••	Asn	•••	•••	Gln	• • •	Gln		• •,•
	60A	• • •	Phe	• • •	• • •	Ser	• • •	• • •		Asn
	BMP5	•••	Phe	•••	• • •	Asp	• • •	• • •		Ser
	BMP6	•••	Asn	• • •	• • •	Asp	•••	• • •	•	Ser
					30					35
<b>25</b>										
	hOP-1	Phe	Pro	Leu	Asn	Ser	Tyr	Met	Asn	Ala
	mOP-1	• • •	• • •	• • •	• • •	• • •	• • •	•••	• • •	• • •
	hOP-2	• • •	• • • •	•••	Asp	• • •	Cys	•••	• • •	• • •
	mOP-2	• • •	• • •	• • •	Asp	•••	Cys	•••	• • •	• • •
30	DPP	•••	• • •	•••	Ala	Asp	His	Phe	• • •	Ser
	Vgl	Tyr	•••	• • •	Thr	Glu	Ile	Leu	• • •	Gly
	Vgr-1	•••	• • •		•••	Ala	His	•••	• • •	• • •
	CBMP-2A	• • •	• • •	•••	Ala	Asp	His	Leu	• • •	Ser
	CBMP-2B	•••	•••	• • •	Ala	Asp	His	Leu	• • •	Ser
						_				

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	GDF-1	Leu	•••	Val	Ala	Leu	Ser	Gly	Ser**	• • •
	BMP3	• • •	•••	Met	Pro	Lys	Ser	Leu	Lys	Pro
	60A	•••	• • •	• • •	• • •	Ala	His	• • •	• • •	• • •
	BMP5	•••	•••	•••	• • •	Ala	His	Met	• • •	• • •
5	BMP6	• • •	•••	• • •	• • •	Ala	His	Met	• • •	• • •
						40				
	•									
	hOP-1	Thr	Asn	His	Ala	Ile	Val	${\tt Gln}$	Thr	Leu
	mOP-1	• • •	• • •	• • •	• • •		•••	• • •	• • •	• • •
10	hOP-2	• • •	• • •	• • •	• • •	• • •	Leu	• • •	Ser	• • •
	mOP-2	• • •	• • •	•••	• • •		Leu	• • •	Ser	• • •
	DPP	• • •	• • •	• • •	• • •	Val	• • •	• • •	• • •	• • •
	Vgl	Ser	• • •	•••	• • •	• • •	Leu	• • •	• • •	• • •
	Vgr-1	• • •	• • •	• • •	•••	• • •	• • •	• • •	• • •	• • •
15	CBMP-2A	• • •	•••	• • •	•••	• • •	• • •	• • •	• • •	• • •
	CBMP-2B	• • •	•••	• • •	• • •	• • •	• • •	• • •	• • •	• • •
	BMP3	Ser	• • •	•••	• • •	Thr	Ile		Ser	Ile
	GDF-1	Leu	• • •	• • •	• • •	Val	Leu	Arg	Ala	• • •
	60A	• • •	• • •	• • •	•••	• • •	• • •	• • •	• • •	• • •
20	BMP5	• • •	• • •	• • •	•••	• • •	• • •	•••	•••	• • •
	BMP6	•••	•••	• • •	• • •	• • •	• • •	• • •	• • •	• • •
		45					50			
25	hOP-1	Val	His	Phe	Ile	Asn	Pro	Glu	Thr	Val
	mOP-1	• • •	• • •	• • •	• • •	• • •	• • •	Asp	• • •	• • •
	hOP-2	• • •	His	Leu	Met	Lys	•••	Asn	Ala	• • •
	mOP-2	•••	His	Leu	Met	Lys	• • •	Asp	Val	
	DPP	• • •	Asn	Asn	Asn	• • •	• • •	Gly	Lys	• • •
30	Vgl	•••	• • •	Ser	• • •	Glu	• • •	•••	Asp	Ile
	Vgr-1	• • •	•••	Val	Met	• • •	• • •	•••	Tyr	• • •
	CBMP-2A	•••	Asn	Ser	Val	• • •	Ser		Lys	Ile
	CBMP-2B	•••	Asn	Ser	Val	• • •	Ser		Ser	Ile
	BMP3	•••	Arg	Ala**		Val	Val	Pro	Gly ·	
		,	- 0	-	•				-	

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	GDF-1	Met	•••	Ala	Ala	Ala	• • •	Gly	Ala	Ala
	60A	• • •	• • •	Leu	Leu	Glu	• • •	Lys	Lys	• • •
	BMP5	• • •	• • •	Leu	Met	Phe	• • •	Asp	His	• • •
	BMP6	• • •	• • •	Leu	Met	• • •	• • •	• • •	Tyr	• • •
5			55					60		
										4
	hOP-1	Pro	Lys	Pro	Cys	Cys	Ala	Pro	Thr	Gln
	mOP-1	• • •	• • •	• • •	• • •	• • •	• • •	•••	• • •	• • •
10	hOP-2	• • •	• • •	Ala	• • •	• • •	• • •	• • •	• • •	Lys
	mOP-2	• • •	• • •	Ala	• • •	• • •	• • •	• • •	• • •	Lys
	DPP	• • •	• • •	Ala		• • •	Val	• • •	• • •	• • •
	Vgl	• • •	Leu	• • •	•••	• • •	Val	• • •	• • •	Lys
	Vgr-1	• • •	• • •	•••	• • •	• • •	• • •	• • •	• • •	Lys
15	CBMP-2A	• • •	• • •	Ala	• • •		Val	• • •	• • •	Glu
	CBMP-2B	• • •	•••	Ala	•••	• • •	Val	• • •	• • •	Glu
	BMP3	• • •	Glu	• • •	• • •	• • •	Val	• • •	Glu	Lys
	GDF-1	Asp	Leu	• • •	• • •	• • •	Val	• • •	Ala	Arg
	60A	• • •	• • •	• • •	• • •	• • •	• • •	• • •	• • •	Arg
20	BMP5	• • •	• • •	•••	• • •	• • •		• • •	• • •	Lys
	BMP6	• • •	• • •	• • •	• • •	• • •	• • •		• • •	Lys
				65					70	
	hOP-1	Leu	Asn	Ala	Ile	Ser	Val	Leu	Tyr	Phe
25	mOP-1	• • •	• • •	• • •	• • •	• • •	• • •	• • •	•••	• • •
	hOP-2		Ser	• • •	Thr	•••		• • •	•••	Tyr
	mOP-2	• • •	Ser	• • •	Thr	•••	•••.	•••	•••	Tyr
	Vgl	Met	Ser	Pro	• • •	• • •	Met	• • •	Phe	Tyr
	Vgr-1	Val	• • •	• • •	• • •	• • •	• • •	• • •	•••	• • •
30	DPP	• • •	Asp	Ser	Val	Ala	Met	• • •	• • •	Leu
	CBMP-2A	• • •	Ser	• • •	• • •	• • •	Met	• • •	• • •	Leu
	CBMP-2B	• • •	Ser	• • •	• • •	• • •	Met	•••	•••	Leu
	BMP3	Met	Ser	Ser	Leu	• • •	Ile	• • •	Phe	Tyr
	GDF-1	• • •	Ser	Pro	• • •	• • •	• • •	• • •	Phe	

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	60A	•••	Gly	•••	Leu	Pro	• • •	• • •	•••	His
	BMP5	•••	•••	•••	• • •	• • •	•••	• • •	•••	• • •
	BMP6	• • •	• • •	•••	• • •	• • •	• • •	• • •	•••	• • •
					75					80
5										
	hOP-1	Asp	Asp	Ser	Ser	Asn	Val	Ile	Leu	Lys
	mOP-1	• • •	•••	• • •	•••	• • •	•••	•••	• • •	• • •
	hOP-2	•••	Ser	•••	Asn	• • •	• • •	• • •	• • •	Arg
	mOP-2	•••	Ser	•••	Asn	• • •	• • •	•••	•••	Arg
10	DPP	Asn	• • •	Gln	•••	Thr	• • •	Val	• • •	• • •
	Vgl	•••	Asn	Asn	Asp	• • •	• • •	Val	• • •	Arg
	Vgr-1	• • •	• • •	Asn	• • •	• • •	•••	• • •	• • •	• • •
	CBMP-2A	•••	Glu	Asn	Glu	Lys	• • •	Val	• • •	• • •
	CBMP-2B	•••	Glu	Tyr	Asp	Lys	• • •	Val	•••	• • •
15	вирз	•••	Glu	Asn	Lys		• • •	Val	• • •	
	GDF-1	•••	Asn	• • •	Asp	•••		Val	• • •	Arg
	60A	Leu	Asn	Asp	Glu			Asn	• • •	• • •
	BMP5	•••	•••	•••	• • •	•••	•••		• • •	• • •
	BMP6	•••	•••	Asn	• • •	• • •	• • •		•••	
20						85				
	hOP-1	Lys	Tyr	Arg	Asn	Met	Val	Val	Arg	
	mOP-1	•••	•••	•••	• • •	• • •		•••	• • •	
25	hOP-2	• • •	His		•••	•••	• • •	• • •	Lys	
	mOP-2	• • •	His		•••	•••		• • •	Lys	
	DPP	Asn		Gln	Glu	• • •	Thr	• • •	Val	
	Vgl	His		Glu	• • •		Ala	•••	Asp	
	Vgr-1	•••	• • •	• • •	• • •		• • •	• • •	•••	
30	CBMP-2A	Asn	•••	Gln	Asp	•••	•••	• • •	Glu	
20	CBMP-2B	Asn	•••	Gln	Glu	•••	•••	•••	Glu	
		22211	• • •	~			- • •			

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	BMP3	Val	•••	Pro	• • •	• • •	Thr	• • •	Glu
	GDF-1	Gln	• • •	Glu	Asp	• • •	•••	• • •	Asp
	60A	•••	• • •	• • •	•••	•••	Ile	• • •	Lys
	BMP5	•••	• • •	• • •	• • •	•••	• • •	• • •	• • •
5	BMP6	• • •	• • •	• • •	Trp	•••	• • •	• • •	• • •
		· <b>9</b> 0					95		
	h0P-1	Ala	Cys	Gly	Cys	His			
10	mOP-1	• • •	•••	• • •	• • •	• • •			
	hOP-2	• • •	•••	• • •	• • •	• • •			
	mOP-2	•••	•••	• • •	•••	• • •			
	DPP	Gly	•••	• • •	• • •	Arg			
	Vgl	Glu	• • •	• • •	•••	Arg			
15	Vgr-1	• • •	• • •	• • •	• • •	• • •			
	CBMP-2A	Gly	• • •	•••	• • •	Arg			•
	CBMP-2B	Gly	•••	• • •	• • •	Arg			
	вмр3	Ser	• • •	Ala	• • •	Arg			
	GDF-1	Glu	• • •	• • •	• • •	Arg			
20	60A	Ser	•••	• • •	• • •	• • •			
	BMP5	Ser	• • •	• • •	•••	• • •			
	BMP6	• • • .	• • •	• • •	• • •	• • •			
				100					
	**Between	residues	56 an	d 57	of BMP3	is a	Val	residue;	

As is apparent from the foregoing amino acid
sequence comparisons, significant amino acid changes
can be made within the generic sequences while
retaining the morphogenic activity. For example, while

between residues 43 and 44 of GDF-1 lies

the amino acid sequence Gly-Gly-Pro-Pro.

the GDF-1 protein sequence depicted in Table II shares only about 50% amino acid identity with the hOP1 sequence described therein, the GDF-1 sequence shares greater than 70% amino acid sequence homology (or "similarity") with the hOP1 sequence, where "homology" or "similarity" includes allowed conservative amino acid changes within the sequence as defined by Dayoff, et al., Atlas of Protein Sequence and Structure vol.5, supp.3, pp.345-362, (M.O. Dayoff, ed., Nat'l BioMed.

The currently most preferred protein sequences useful as morphogens in this invention include those having greater than 60% identity, preferably greater 15 than 65% identity, with the amino acid sequence defining the conserved six cysteine skeleton of hOP1 (e.g., residues 43-139 of Seq. ID No. 5). These most preferred sequences include both allelic and species variants of the OP-1 and OP-2 proteins, including the 20 Drosophila 60A protein. Accordingly, in still another preferred aspect, the invention includes morphogens comprising species of polypeptide chains having the generic amino acid sequence referred to herein as "OPX", which defines the seven cysteine skeleton and 25 accommodates the identities between the various identified mouse and human OP1 and OP2 proteins. is presented in Seq. ID No. 29. As described therein, each Xaa at a given position independently is selected from the residues occurring at the corresponding 30 position in the C-terminal sequence of mouse or human OP1 or OP2 (see Seq. ID Nos. 5-8 and/or Seq. ID Nos. 16-23).

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# II. <u>Formulations and Methods for Administering</u> Therapeutic <u>Agents</u>

#### A. General Considerations

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The morphogens may be provided to an individual by any suitable means, most preferably orally, or, alternatively, parenterally. Where the morphogen is to be provided parenterally, such as intravenously or by enteral feeding tube, the morphogen preferably comprises part of an aqueous solution. The solution is physiologically acceptable so that in addition to delivery of the desired morphogen to the patient, the solution does not otherwise adversely affect the patient's electrolyte and volume balance. The aqueous medium for the morphogen thus may comprise normal physiologic saline (0.85% NaCl, 0.15M), pH 7-7.4. aqueous solution containing the morphogen can be made, for example, by dissolving the protein in 50% ethanol containing acetonitrile in 0.1% trifluoroacetic acid 20 (TFA) or 0.1% HCl, or equivalent solvents. One volume of the resultant solution then is added, for example, to ten volumes of phosphate buffered saline (PBS), which further may include 0.1-0.2% human serum albumin (HSA). The resultant solution preferably is vortexed 25 extensively. If desired, a given morphogen may be made more soluble by association with a suitable molecule. For example, the pro form of the morphogenic protein comprises a species that is soluble in physiologically 30 buffered solutions. In fact, the endogenous protein is thought to be transported (e.g., secreted and circulated) in this form. This soluble form of the protein may be obtained from the culture medium of morphogen-secreting mammalian cells. Alternatively, a

soluble species may be formulated by complexing (e.g., via non-covalent interaction) the mature dimer (or an active fragment thereof) with part or all of one or, preferably, two pro domain peptides (see Section A.1, below). Another molecule capable of enhancing solubility and particularly useful for oral administrations, is casein, including derivatives and analogs thereof. For example, addition of 0.2% casein increases solubility of the mature active form of OP-1 in physiologically buffered solutions by 80%. Other components found in milk and/or various serum proteins also may be useful.

Useful solutions for parenteral administration may

be prepared by any of the methods well known in the

pharmaceutical art, described, for example, in

Remington's Pharmaceutical Sciences (Gennaro, A., ed.),

Mack Pub., 1990. Formulations may include, for

example, polyalkylene glycols such as polyethylene

glycol, oils of vegetable origin, hydrogenated

naphthalenes, and the like. Biocompatible, preferably

bioresorbable, polymers, including, for example,

hyaluronic acid, collagen, polybutyrate, tricalcium

phosphate, lactide and lactide/glycolide copolymers,

may be useful excipients to control the release of the

morphogen in vivo.

As described above, the dietary supplements comprising the morphogens described herein preferably are provided orally. Oral administration of proteins as therapeutics generally is not practiced as most proteins are readily degraded by digestive enzymes and acids in the mammalian digestive system before they can be absorbed into the bloodstream. However, the

morphogens described herein typically are acid stable and protease-resistant (see, for example, U.S. Pat.No. 4,968,590.) In addition, at least one morphogen, OP-1, has been identified in mammary gland extract, colostrum 5 and 57-day milk (see Example 1, below). Moreover, the OP-1 purified from mammary gland extract is morphogenically active. Specifically, this protein induces endochondral bone formation in mammals when implanted subcutaneously in association with a suitable 10 matrix material, using a standard in vivo bone assay, such as is disclosed in U.S. Pat.No. 4,968,590. Moreover, the morphogen also is detected in the bloodstream (see Example 2, below). These findings indicate that oral and parenteral administration are 15 viable means for administering morphogens to an individual. In addition, while the mature forms of certain morphogens described herein typically are sparingly soluble, the morphogen form found in milk (and mammary gland extract and colostrum) is readily 20 soluble, probably by association of the mature, morphogenically active form with part or all of at least one pro domain peptide and/or by association with one or more milk components. Accordingly, the compounds provided herein also may be associated with 25 molecules capable of enhancing their solubility in vitro or in vivo.

The dietary compositions for oral administration may be formulated as a liquid, for example, as part of an aqueous medium as described above for parenteral administration, and which further may contain flavoring and coloring agents. The formulation also may be combined with a beverage or may be provided in a syrup. The dietary composition also may be provided as

an aerosol for oral or nasal administration.

Formulations for inhalation administration contain as excipients, for example, lactose, or may be aqueous solutions containing, for example, polyoxyethylene-9-lauryl ether, glycocholate and deoxycholate, or oily solutions for administration in the form of nasal drops, or as a gel to be applied intranasally.

Alternatively, the dietary composition may be provided as a solid, for example as a tablet, capsule or lozenge. As for parenteral administration, formulations for oral administration also may include molecules to enhance a controlled release of the morphogen in vivo.

As will be appreciated by those skilled in the art, 15 the concentration of the compounds described in a given dietary supplement composition will vary depending upon a number of factors, including the dosage number to be administered, the chemical characteristics (e.g., 20 hydrophobicity) of the compounds employed, and the The preferred dosage to be route of administration. administered also is likely to depend on such variables as the type and extent of tissue development enhancement desired, the type and extent of any tissue damage present to be repaired, the overall health status of the particular individual, the relative biological efficacy of the compound selected, the formulation of the compound excipients, and its route of administration. In general terms, the compounds of 30 this invention may be provided in a formulation containing about 0.001 to 10% w/v of morphogen to formulation. Typical dose ranges are from about 10 ng/kg to about 1 g/kg of body weight per day; a preferred dose range is from about 0.1  $\mu$ g/kg to

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100 mg/kg of body weight per day. Optimally, the morphogen dosage given is between 0.1-100  $\mu$ g of protein per kilogram weight of the individual. No obvious morphogen induced pathological lesions are induced when 5 mature morphogen (e.g., OP-1, 20  $\mu$ g) is administered daily to normal growing rats for 21 consecutive days. Moreover, 10  $\mu$ g systemic injections of morphogen (e.g., OP-1) injected daily for 10 days into normal newborn mice does not produce any gross abnormalities.

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In administering morphogens parenterally in the methods of the present invention, preferably a large volume loading dose is used at the start of the treatment. The treatment then is continued with a 15 maintenance dose. In all cases administration dosages then can be monitored by measuring at intervals the levels of the morphogen in the blood.

#### A.1 Soluble Morphogen Complexes

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A currently preferred form of the morphogen useful in therapeutic formulations, having improved solubility in aqueous solutions and consisting essentially of amino acids, is a dimeric morphogenic protein comprising at least the 100 amino acid peptide sequence having the pattern of seven or more cysteine residues characteristic of the morphogen family complexed with a peptide comprising part or all of a pro region of a member of the morphogen family, or an allelic, species 30 or other sequence variant thereof. Preferably, the dimeric morphogenic protein is complexed with two peptides. Also, the dimeric morphogenic protein preferably is noncovalently complexed with the pro region peptide or peptides. The pro region peptides

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also preferably comprise at least the N-terminal eighteen amino acids that define a given morphogen pro region. In a most preferred embodiment, peptides defining substantially the full length pro region are used.

Other soluble forms of morphogens include dimers of the uncleaved pro forms of these proteins, as well as "hemi-dimers" wherein one subunit of the dimer is an uncleaved pro form of the protein, and the other subunit comprises the mature form of the protein, including truncated forms thereof, preferably noncovalently associated with a cleaved pro domain peptide.

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As described above, useful pro domains include the full length pro regions, as well as various truncated forms hereof, particularly truncated forms cleaved at proteolytic Arg-Xaa-Xaa-Arg cleavage sites. 20 example, in OP-1, possible pro sequences include sequences defined by residues 30-292 (full length form); 48-292; and 158-292. Soluble OP-1 complex stability is enhanced when the pro region comprises the full length form rather than a truncated form, such as 25 the 48-292 truncated form, in that residues 30-47 show sequence homology to the N-terminal portions of other morphogens, and are believed to have particular utility in enhancing complex stability for all morphogens. Accordingly, currently preferred pro sequences are 30 those encoding the full length form of the pro region for a given morphogen. Other pro sequences

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contemplated to have utility include biosynthetic pro sequences, particularly those that incorporate a sequence derived from the N-terminal portion of one or more morphogen pro sequences.

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As will be appreciated by those having ordinary skill in the art, useful sequences encoding the pro region may be obtained from genetic sequences encoding known morphogens. Alternatively, chimeric pro regions can be constructed from the sequences of one or more known morphogens. Still another option is to create a synthetic sequence variant of one or more known pro region sequences.

In another preferred aspect, useful pro region peptides include polypeptide chains comprising an amino acid sequence encoded by a nucleic acid that hybridizes under stringent conditions with a DNA or RNA sequence encoding at least the N-terminal eighteen amino acids of the pro region sequence for OP1 or OP2, e.g., nucleotides 136-192 and 152-211 of Seq. ID No. 16 and 20, respectively.

# A.1A <u>Isolation of Soluble morphogen complex from</u> conditioned media or body fluid\_

Morphogens are expressed from mammalian cells as soluble complexes. Typically, however the complex is disassociated during purification, generally by

30 exposure to denaturants often added to the purification solutions, such as detergents, alcohols, organic solvents, chaotropic agents and compounds added to reduce the pH of the solution. Provided below is a currently preferred protocol for purifying the soluble

proteins from conditioned media (or, optionally, a body fluid such as serum, cerebro-spinal or peritoneal fluid), under non-denaturing conditions. The method is rapid, reproducible and yields isolated soluble morphogen complexes in substantially pure form.

Soluble morphogen complexes can be isolated from conditioned media using a simple, three step chromatographic protocol performed in the absence of denaturants. The protocol involves running the media (or body fluid) over an affinity column, followed by ion exchange and gel filtration chromatographies. affinity column described below is a Zn-IMAC column. The present protocol has general applicability to the purification of a variety of morphogens, all of which are anticipated to be isolatable using only minor modifications of the protocol described below. alternative protocol also envisioned to have utility an immunoaffinity column, created using standard 20 procedures and, for example, using antibody specific for a given morphogen pro domain (complexed, for example, to a protein A-conjugated Sepharose column.) Protocols for developing immunoaffinity columns are well described in the art, (see, for example, Guide to 25 Protein Purification, M. Deutscher, ed., Academic Press, San Diego, 1990, particularly sections VII and XI.)

In this experiment OP-1 was expressed in mammalian 30 CHO (chinese hamster ovary) cells as described in the art (see, for example, international application US90/05903 (WO91/05802).) The CHO cell conditioned media containing 0.5% FBS was initially purified using Immobilized Metal-Ion Affinity Chromatography (IMAC).

The soluble OP-1 complex from conditioned media binds very selectively to the Zn-IMAC resin and a high concentration of imidazole (50 mM imidazole, pH 8.0) is required for the effective elution of the bound 5 complex. The Zn-IMAC step separates the soluble OP-1 from the bulk of the contaminating serum proteins that elute in the flow through and 35 mM imidazole wash fractions. The Zn-IMAC purified soluble OP-1 is next applied to an S-Sepharose cation-exchange column equilibrated in 20 mM NaPO, (pH 7.0) with 50 mM NaCl. This S-Sepharose step serves to further purify and concentrate the soluble OP-1 complex in preparation for the following gel filtration step. The protein was applied to a Sephacryl S-200HR column equilibrated in 15 TBS. Using substantially the same protocol, soluble morphogens also may be isolated from one or more body fluids, including serum, cerebro-spinal fluid or peritoneal fluid.

IMAC was performed using Chelating-Sepharose (Pharmacia) that had been charged with three column volumes of 0.2 M ZnSO4. The conditioned media was titrated to pH 7.0 and applied directly to the ZN-IMAC resin equilibrated in 20 mM HEPES (pH 7.0) with 500 mM NaCl.

The Zn-IMAC resin was loaded with 80 mL of starting conditioned media per mL of resin. After loading, the column was washed with equilibration buffer and most of the contaminating proteins were eluted with 35 mM imidazole (pH 7.0) in equilibration buffer. The soluble OP-1 complex then is eluted with 50 mM imidazole (pH 8.0) in 20 mM HEPES and 500 mM NaCl.

The 50 mM imidazole eluate containing the soluble OP-1 complex was diluted with nine volumes of 20 mM NaPO<sub>4</sub> (pH 7.0) and applied to an S-Sepharose (Pharmacia) column equilibrated in 20 mM NaPO<sub>4</sub> (pH 7.0) 5 with 50 mM NaCl. The S-Sepharose resin was loaded with an equivalent of 800 mL of starting conditioned media per mL of resin. After loading the S-Sepharose column was washed with equilibration buffer and eluted with 100 mM NaCl followed by 300 mM and 500 mM NaCl in 20 mM  $NaPO_A$  (pH 7.0). The 300 mM NaCl pool was further 10 purified using gel filtration chromatography. mls of the 300 mm NaCl eluate was applied to a 5.0 X 90 cm Sephacryl S-200HR (Pharmacia) equilibrated in Tris buffered saline (TBS), 50 mM Tris, 150 mM NaCl (pH 7.4). The column was eluted at a flow rate of 5 mL/minute collecting 10 mL fractions. The apparent molecular of the soluble OP-1 was determined by comparison to protein molecular weight standards (alcohol dehydrogenase (ADH, 150 kDa), bovine serum 20 albumin (BSA, 68 kDa), carbonic anhydrase (CA, 30 kDa) and cytochrome C (cyt C, 12.5 kDa). The purity of the S-200 column fractions was determined by separation on standard 15% polyacrylamide SDS gels stained with coomassie blue. The identity of the mature OP-1 and 25 the pro-domain was determined by N-terminal sequence analysis after separation of the mature OP-1 from the pro-domain using standard reverse phase C18 HPLC.

The soluble OP-1 complex elutes with an apparent
30 molecular weight of 110 kDa. This agrees well with the
predicted composition of the soluble OP-1 complex with
one mature OP-1 dimer (35-36 kDa) associated with two

pro-domains (39 kDa each). Purity of the final complex can be verified by running the appropriate fraction in a reduced 15% polyacrylamide gel.

The complex components can be verified by running 5 the complex-containing fraction from the S-200 or S-200HR columns over a reverse phase C18 HPLC column and eluting in an acetonitrile gradient (in 0.1% TFA), using standard procedures. The complex is dissociated 10 by this step, and the pro domain and mature species elute as separate species. These separate species then can be subjected to N-terminal sequencing using standard procedures (see, for example, Guide to Protein Purification, M. Deutscher, ed., Academic 15 Press, San Diego, 1990, particularly pp. 602-613), and the identity of the isolated 36kD, 39kDa proteins confirmed as mature morphogen and isolated, cleaved pro domain, respectively. N-terminal sequencing of the isolated pro domain from mammalian cell produced OP-1 20 revealed 2 forms of the pro region, the intact form (beginning at residue 30 of Seq. ID No. 16) and a truncated form, (beginning at residue 48 of Seq. ID No. 16.) N-terminal sequencing of the polypeptide subunit of the isolated mature species reveals a range of 25 N-termini for the mature sequence, beginning at residues 293, 300, 313, 315, 316, and 318, of Seq. ID No. 16, all of which are active as demonstrated by the standard bone induction assay.

# A.1B. In Vitro Soluble Morphogen Complex Formation

As an alternative to purifying soluble complexes from culture media or a body fluid, soluble complexes 5 may be formulated from purified pro domains and mature dimeric species. Successful complex formation apparently requires association of the components under denaturing conditions sufficient to relax the folded structure of these molecules, without affecting 10 disulfide bonds. Preferably, the denaturing conditions mimic the environment of an intracellular vesicle sufficiently such that the cleaved pro domain has an opportunity to associate with the mature dimeric species under relaxed folding conditions. 15 concentration of denaturant in the solution then is decreased in a controlled, preferably step-wise manner, so as to allow proper refolding of the dimer and pro regions while maintaining the association of the pro domain with the dimer. Useful denaturants include 4-6M 20 urea or guanidine hydrochloride (GuHCl), in buffered solutions of pH 4-10, preferably pH 6-8. The soluble complex then is formed by controlled dialysis or dilution into a solution having a final denaturant concentration of less than 0.1-2M urea or GuHCl, 25 preferably 1-2 M urea of GuHCl, which then preferably can be diluted into a physiological buffer. Protein purification/renaturing procedures and considerations are well described in the art, and details for developing a suitable renaturing protocol readily can 30 be determined by one having ordinary skill in the art.

One useful text one the subject is <u>Guide to Protein</u>

<u>Purification</u>, M. Deutscher, ed., Academic Press, San

Diego, 1990, particularly section V. Complex formation
also may be aided by addition of one or more chaperone

proteins.

### A.1C. Stability of Soluble Morphogen Complexes

The stability of the highly purified soluble 10 morphogen complex in a physiological buffer, e.g., tris-buffered saline (TBS) and phosphate-buffered saline (PBS), can be enhanced by any of a number of means. Currently preferred is by means of a pro region that comprises at least the first 18 amino acids of the pro sequence (e.g., residues 30-47 of Seq. ID NO. 16 for OP-1), and preferably is the full length pro region. Residues 30-47 show sequence homology to the N-terminal portion of other morphogens and are believed to have particular utility in enhancing complex stability for all morphogens. Other useful means for enhancing the stability of soluble morphogen complexes include three classes of additives. These additives include basic amino acids (e.g., L-arginine, lysine and betaine); nonionic detergents (e.g., Tween 80 or NonIdet P-120); and carrier proteins (e.g., serum albumin and casein). Useful concentrations of these additives include 1-100 mM, preferably 10-70 mM, including 50 mM, basic amino acid;, 0.01-1.0%, preferably 0.05-0.2%, including 0.1% (v/v) nonionic detergent;, and 0.01-1.0%, preferably 0.05-0.2%, 30 including 0.1% (w/v) carrier protein.

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#### B. Considerations for Infant and Other Formulas

#### 1. Infant Formulas

In all cases the morphogens of this invention preferably are added to an infant formula that complies with the nutritional guidelines provided by the AAP and ESPGAN. Basic ingredients for infant formulas include cow's milk, protein, whey proteins, casein and its salts (i.e. calcium caseinate). Soy protein isolates may be substituted for milk-derived proteins, and preferably are used in the products made for infants with lactose intolerance and/or cow's protein intolerance. Protein hydrolyzates (i.e. casein and lactalbumin hydrolyzates) with low molecular weight, also may be used for these products.

The proportions of the diverse component nutrients preferably are similar to those of human milk. Thus, the ratio of whey proteins to casein preferably varies from 60:40 to 70:30 in infant formulas based on milk. The mixture of fats employed is made up of edible fats to provide an essential fatty acid profile. Lactose preferably is used as the carbohydrate source for at-term newborns infants, and dextrinmaltose preferably is employed in products used for the treatment of lactose intolerance and malabsorption syndromes in infancy.

Infant formulas according to the invention also preferably contain minerals (including calcium, phosphorus, sodium, potassium, chloride, magnesium, iron, zinc, copper, manganese and iodine) and vitamins (including vitamin A, vitamin D3, vitamin C, vitamin

B1, vitamin B2, vitamin B6, vitamin B12, pantothenic acid, vitamin E, vitamin K1, folic acid, biotin) adequate for the infants' requirements. Also, in the products whose source of proteins is derived from soy or protein isolates or hydrolyzates, carnitine preferably is included to satisfy the nutritional requirements for this compound in infants with malabsorptive syndromes.

10 A typical ready-to-feed morphogen-enriched formulation for infants, when diluted to feeding concentrations, preferably comprises in addition to the added morphogen, from about 1-5% by weight fat, from about 0.01 to about 0.5% by weight immunoglobulins as appropriate, from about 4-10% by weight carbohydrate in a quantity substantially to mimic the carbohydrate content of human mother's milk, from about 0.5 to 4% by weight protein in a quantity substantially to mimic the protein content of human mother's milk, optional vitamins and minerals as required, a total solids content of from about 8 to 17% by weight, and the remainder water.

A typical protein source for use in infant formula
is electrodialyzed whey or electrodialyzed skim milk or
milk whey, although other protein sources are also
available and may be used. Preferred sugars include
food grade substances such as glucose, dextrose,
sucrose, or edible lactose. The following vitamins and
minerals may also be incorporated in the infant
formula: calcium, phosphorus, potassium, sodium,
chloride, magnesium, manganese, iron, copper, zinc,
selenium, iodine, and vitamins A, E, D, and B complex.

These micronutrients are added in the form of commonly accepted nutritional compounds in amounts equivalent to those present in human milk on a per calories basis.

The infant formula according to the present 5 invention also preferably is sterilized and subsequently used on a ready-to-feed basis, or can be The concentrate can be stored as a concentrate. prepared using standard procedures known in the art, 10 and the formula can be reconstituted by rehydrating the concentrate. The infant formula preferably is a stable A more detailed liquid and has a suitable shelf life. description of infant formula considerations, including preferred formulations for newborn, preterm and low 15 birth-weight infants, lactose-intolerant infants, may be found, for example, in US Pat. No. 5,066,500 to Gil et al., the disclosure of which is incorporated herein by reference.

#### 20 2. Other Nutritional Products

The morphogen-enriched dietary products for balanced nutrition (e.g., dietary food formulations) according to the present invention, preferably have, in addition to added morphogen, a composition of nutrients adequate to the specific requirements of not only healthy human in need of a balanced nutritional product, but also those individuals at risk for lost or reduced tissue function due malnutrition-maladsorption disorder, and/or altered metabolism. Individuals particularly affected by an altered metabolic function include postmenopausal women or aged individuals, hypercatabolic individuals, and individuals undergoing periods of rapid growth or physical stress, such as

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developing juveniles, and pregnant, lactating and nursing mothers. Other individuals at risk are those suffering from malnutrition, induced, for example, by starvation and/or an eating disorder, and individuals affected with energy-protein malnutrition and in hypercatablic states derived from traumatic, septic, surgical processes and other clinically-derived malabsorption syndromes.

10 Morphogen-enriched nutritional products according to the present invention preferably also provide mineral elements which include trace elements and vitamins in adequate proportions to satisfy the specific requirements of normal healthy individuals as well as individuals at risk, such as those suffering malabsorption-malnutrition processes and in a hypercatabolic state. The nutritional products also preferably are enriched with amino acids sources, vitamins, nucleosides and/or nucleotides in similar amounts to those present in ordinary foods.

As described above for infant formulas, liquid products may be formulated ready for consumption or as concentrates to be diluted before use. Preferably, liquid dietary compositions have pH values generally ranging from about 6.0 to about 8.0, most preferbly 6.8-7.5.

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Useful dietary compositions and considerations for their formulation are well described in the medical and nutritional arts. Useful compositions for clinical nutrition, also are described in detail in US
5 Pat.No. 5,066,500.

#### III. Examples

# 10 Example 1. Determination of the Presence of Morphogen in Milk

Morphogenically active OP-1 was demonstrated to be present in mammary gland extract, colostrum, and milk, as described below. The discovery that the morphogen 15 naturally is present in milk, together with the known observation that mature, active OP-1 is acid-stable and protease-resistant, indicate that oral administration is a useful route for therapeutic administration of morphogen to a mammal. Oral administration typically 20 is the preferred mode of delivery for extended or prophylactic therapies. In addition, the identification of morphogen in all milk forms, including colostrum, indicates that the protein plays a 25 significant role in tissue development, including skeletal development of juveniles.

Rat mammary gland extract and bovine colostrum and 57 day milk were subjected to purification procedures designed to partially purify OP-1. The partially purified product then was examined for the presence of OP-1 by Western blot analysis using OP-1-specific antisera, and tested for in vivo and in vitro activity.

#### 1.1 Purification

The purification protocol for all three "milk" forms (e.g., mammary gland extract, colostrum and 57-5 day milk), involved three chromatography steps: (1) cation-exchange chromatography (S-Sepharose and followed by Phenyl-Sepharose chromatography); (2) Copper-Immobilized Metal Affinity chromatography (Cu++-IMAC); and finally, (3) C-18 reverse phase 10 chromatography. Fractions were sampled at each step for the presence of OP-1. Fraction samples for testing were dialyzed versus water/0.1% TFA, then against 30% acetonitrile/0.1% TFA for analysis on SDSpolyacrylamide gels and immunoblots, using standard methodologies well described in the art. Unless otherwise stated, the primary antibody used for the immunoblots was made against full length OP-1 produced in E.coli using standard recombinant DNA and antibody production techniques (see, for example, Example 8, below for a general description for producing morphogen-specific antibodies.) Fractions found to contain the morphogen then were applied to the next column step or used in the immunoreactivity or activity assays described below.

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Essentially the same protocol was followed for all three milk sources, except that two alternative cationexchange methodologies were employed for colostrum purification, described in detail below. Unless

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otherwise indicated, all chemicals referenced are standard, commercially available reagents, readily available from a number of sources, including Sigma Chemical, Co., St. Louis; Calbiochem, Corp., San Diego 5 and Aldrich Chemcial Co., Milwaukee.

#### step 1. Cation-Exchange Chromatography

The S-Sepharose purification step was performed as 200ml of cation exchanger (S-Sepharose, Sigma Chemical Corp.) were equilibrated with equilibration buffer (6M urea, 20mM MES, 70mM NaCl, pH 6.5). supernatant from the centrifuged extract was diluted to final concentration of 6M urea, 20mM MES, 70mM NaCl, pH 6.5. After loading, the column was washed to baseline 15 using equilibration buffer, and the bound components were eluted stepwise from the column with 6M urea, 20mM MES, 100mM and 500mM NaCl, pH 6.5. The more tightly bound components then were eluted with 4M guanidine, 20 20mM sodium phosphate, pH 7.0.

The Phenyl-Sepharose purification step was performed as follows. 15ml of Phenyl-Sepharose CL-4B (Sigma) were equilibrated with 6M urea, 20mM HEPES, 1M ammonium sulfate, 300mM NaCl, pH 7.0. The 500mM NaCl eluate from the S-Sepharose step was diluted with 6M urea, 20mM HEPES, 3M ammonium sulfate, 300mM NaCl, pH 7.0, to a final concentration of 1M ammonium sulfate, pH 7.0. After loading, the column was washed to 30 baseline with equilibration buffer. The column was eluted with 6M urea, 20mM HEPES, 0.6M ammonium sulfate, 300mM NaCl, pH 7.0, and then with 4M guanidine, 20mM sodium phosphate, pH 7.0.

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Two alternative cation-exchange chromatography schemes (A and B) were exploited in the purification of OP-1 from colostrum, as follows. For both schemes, 200 ml of S-Sepharose (Sigma) was poured into a 5 X 10 cm 5 Bio-Rad econocolumn (Bio-Rad, Inc. Cambridge.)

Scheme A: The colostrum, which had been diluted to 6M urea, 20mM sodium phosphate, pH 7.0, was loaded onto a column equilibrated with 6M urea, 20mM sodium

10 phosphate, 50mM NaCl, pH 7.0. Elution was stepwise, with 6M urea, 20mM sodium phosphate, 100mM and then 500mM NaCl, pH 7.0; and the final wash was with 4M guanidine, 20mM sodium phosphate, pH 7.0. The Phenyl-Sepharose column was run as described above, except

15 that sodium phosphate was used as the running buffer instead of HEPES. The Phenyl-Sepharose bound fraction (0.0M ammonium sulfate eluate) from scheme A then was dialyzed into 6M urea, 20mM Hepes, 500mM NaCl, pH 7.0, before it was applied to the Cu++-IMAC column, which

Scheme B: The alternative S-Sepharose purification was performed as follows. Ethanol-precipitated protein was loaded onto an S-Sepharose column equilibrated in 6M urea, 20mM MES, 50mM NaCl, pH 6.5. Elution was stepwise with 6M urea, 20mM MES, 100mM NaCl and then 500mM NaCl, and finally 4M guanidine, 20mM sodium phosphate, pH 7.0. The Phenyl-Sepharose column was run as described above, with the 0.0M ammonium sulfate eluate then applied to a Cu++-IMAC column.

### step 2. Cu++IMAC Chromatography

The Cu++IMAC purification step was performed as follows. 10ml of Pharmacia Fast Flow Chelating Resin were charged with 0.2M cupric sulfate, and equilibrated with 6M urea, 20mM HEPES, 0.5M NaCl, pH 7.0. After loading, the column was washed to baseline with equilibration buffer. Elution from the column was stepwise, using equilibration buffer containing 1mM, or 10mM imidazole. The column then was stripped with equilibration buffer containing 10mM EDTA. The 10mM imidazole elution was dialyzed against water/0.1% TFA, then against 30% acetonitirile/0.1% TFA.

## 15 step 3. Reverse Phase Chromatography

The C-18 reverse phase chromatography purification step was performed as follows. A HPLC C-18 semi-prep column was used for the final purification step. The gradient used was 30-50% acetonitrile/0.1% TFA over 60 minutes at 3ml/minutes. After the sample was loaded, the column was washed to baseline with 30% acetonitrile/0.1% TFA before the gradient is started. Fractions collected were 3ml in size. Chromatograms were read at 214 nm.

## (a) OP-1 from Rat Mammary Gland Extract

Mammary glands were obtained from 2 female Long
30 Evan rats (Charles River Labs, Wilmington, MA) one week
post-partum. The excised glands were mildly
homogenized in 6M urea, 20mM methylethansulfonate

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(MES), 0.5M NaCl, pH 6.5 using a polytron homogenizer. The suspension then was centrifuged for 20 minutes at 8,000 RPM, and the supernatant removed for further purification.

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Following S-Sepharose chromatography, fractions containing 6M urea, 20mM MES containing 500mM NaCl, also appeared to contain OP-1 as determined by SDS and immunoblot, and were applied to the Phenyl-Sepharose The eluate from the 6M urea, 20mM HEPES, 300mM NaCl, pH 7.0 elution step from this column were found to contain OP-1. This eluate then was applied to a Cu++-IMAC column. Eluate fractions found to contain OP-1 were then applied to the C-18 column and 15 chromatographed as described.

Figure 1(A) shows the chromatogram and 1(B) the corresponding Western blot for fractions from the C-18 reverse phase chromatography step run under reducing 20 conditions. Lane S of the Western blot is a standard, containing reduced, purified, recombinantly-produced The arrows show molecular weight markers corresponding to 17, 27, and 39 Kd. The reduced monomer run at approximately 16-18 Kd; the oxidized homodimer at approximately 36 Kd. Lanes 13-30 represent the 25 corresponding fractions of the C-18 reverse phase column as numbered in Fig.1(A). As can be seen in Fig.1(B), mammary extract OP-1 elutes primarily in fractions 21-25 from this final chromatography step.

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## (b) OP-1 from Bovine Colostrum

Colostrum is the first milk to be produced by the mother following birth. Approximately 5 gallons of bovine colostrum were obtained from a local dairy farm and delipidated by centrifugation (8000 rpm for approximately 10 min. at 4°C). The supernatant then was filtered through cheese cloth. The filtered supernatant was stored in 500ml aliquots at 70°C.

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urea, 30mM sodium phosphate, pH 7.0. Alternately, 50ml of colostrum was added to 50ml of 8M guanidine-HC1, 50mM Tris, pH 7.2 and precipitated with 40%, then 85% ice cold ethanol. The pellet was washed with 90% cold ethanol and lyophilized overnight. The lyophilized pellet was resuspended in 6M urea, 20mM MES, 500mM NaCl, pH 6.5, stirred overnight at 4°C, and centrifuged at 9,000 RPM for 10 minutes to clarify the suspension before loading onto the column as described in schemes A and B, above.

Following S-Sepharose chromatography by scheme A, both the 100mM and the 500 mM eluate fractions were found to contain OP-1, with the 100mM fraction containing relatively more morphogen. This fraction then was loaded onto the Phenyl-Sepharose column following dilution with an equal volume of 6M urea, 20 mM sodium phosphate, 2M ammonium sulfate, and 300mM NaCl.

Following S-Sepharose chromatography by scheme B, the 500mM NaCl eluate was found to contain OP-1 and was loaded onto a Phenyl-Sepharose column as described above, following dilution with 6M urea, 40mM HEPES, 2M ammonium sulfate, pH 7.0.

Following Cu++IMAC chromatography OP-1 was identified in the 5mM and 10mM imidazole eluates for both purification schemes, and was dialyzed for further purification on the C-18 column.

Both purification schemes produce purified OP-1, as determined by immunoblot. Figure 2 shows the chromatogram (A) and corresponding Western blot (B) for 15 results of purification scheme A (Fig. 2B-1, reduced and Fig. 2B-2, oxidized); and Figure 3 shows the chromatogram (A) and Western blot (B, reduced) for C-18-purified protein from scheme B. As for Fig. 1B, lane S in Figs. 2B and 3B is a standard, containing purified, recombinantly produced OP-1; 17, 27 and 39 20 are molecular weight markers, and lane numbers correspond to fraction numbers in the corresponding chromatograms. OP-1 purified by scheme A appears predominantly in fractions 18-27, and OP-1 purified by scheme B appears predominantly in fractions 18-25. 25

### OP-1 from 57-day milk

Milk was obtained from the same cow from which the colostrum came, 57 days after the birth of the calf. The milk was delipidated by centrifugation at 10,000 RPM for 15 minutes, and the milk was poured off and away from the fat layer.

100ml of milk then were diluted with 200ml of 9M urea, 30mM MES, pH 6.5 and loaded onto a 200ml S-Sepharose column which had been equilibrated with 6M urea, 20mM MES, 50mM NaCl, pH 6.5. Elution was with 6M urea, 20mM MES, 100mM and 500mM NaCl, and 4M guanidine, 20mM sodium phosphate, pH 7.0. The 500mM elution was put over a Phenyl-Sepharose column after being diluted with an equal volume of 6M urea, 20mM MES, 2M ammonium sulfate, pH 7.0.

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The Phenyl-Sepharose column then was run as described above. The Phenyl-Sepharose-bound sample was eluted and applied to a Cu++IMAC column, prepared and run as described above. The 10mM imidazole eluate was found to contain OP-1 and was dialyzed for further purification on the C-18 column.

The C-18 reverse phase chromatography column and gradient were performed as described above. The

20 results are presented in Fig. 4A (chromatogram) and 4B (immunoblot, 10B-1, oxidized; 4B-2, reduced.) As above, lane S is a standard, containing purified, recombinantly produced OP-1; 17, 27, and 39 are molecular weight markers, and the lane numbers

25 correspond to the fractions numbers in Fig. 4A. OP-1 purified from 57-day milk appears predominantly in fractions 18-26.

### 1.2 OP-1 Characterization by immunoreactivity

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OP-1 purified from the different milk sources as described above also were characterized by Western blotting using antibodies raised against OP-1 and BMP2. Antibodies were prepared using standard immunology

protocols well known in the art, and as described in Example 8, below using full-length <u>E. coli</u>-produced OP-1 and BMP2 as the immunogens.

As shown in Fig. 5 OP-1 purified from colostrum 5 reacts with the anti-OP-1 antibody, but not with anti-BMP2 antibody. In Fig. 5A and B, lane 1 contains reduced, purified, recombinantly-produced OP-1; lane 2 contains C-18 purified bovine colostrum, and lane 3 10 contains reduced COP-16, a biosynthetic construct having morphogenic activity and an amino acid sequence modeled on the proteins described herein, but having highest amino acid sequence homology with BMP2 (see US Pat. No. 5,011,691 for the COP-16 amino acid sequence.) 15 In Fig. 5A the gel was probed with anti-OP-1 antibody; in Fig. 5B, the gel was probed with anti-BMP2 antibody. As can be seen in the figure, anti-OP-1 antibody hybridizes with protein in lanes 1 and 2, but not 3; while anti-BMP2 antibody hybridizes with lane 3 only.

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C-18 purified mammary gland extract and 57-day milk also were shown to react with anti-OP-1 antibodies, including antibody raised against the full length <u>E. coli</u> OP-1, full length mammalian-produced OP-1, and the OP-1 Ser-17-Cys peptide (e.g., the OP-1 N-terminal 17 amino acids).

# 1.3 OP-1 Characterization by Activity

30 The morphogenic activity of OP-1 purified from mammary gland extract was evaluated in vivo as follows. 33% of each OP-1 immunoreactive fraction of C-18-purified mammary gland extract was lyophilized and resuspended in  $220\mu l$  of 50% acetonitrile/0.1% TFA. After

vortexing, 25 mg of collagen matrix was added. The samples were lyophilized overnight, and implanted in Long Evans rats (Charles River Laboratories, Wilmington, MA, 28-35 days old). Each fraction was implanted in duplicate. For details of the collagen matrix implantation procedure, see, for example, U.S. Pat. No. 4,968,590, hereby incorporated by reference. After 12 days, the implants were removed and evaluated for new bone formation by histological observation.

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The results are presented in Fig. 6A, where "% activity" refers to the % of bone formation/total area covered by bone in the histology sample. In the figure, solid bars represent implants using mammary extract-derived OP-1, where the fraction numbers correspond to the related fractions eluted from the C-18 reverse phase column (see Fig. 1B), and the hatched bar represents implants using recombinantly produced OP-1 (600 ng). The results demonstrate that the peak bone forming activity of C-18-purified mammary gland extract corresponds with the immunoreactive fraction peaks of Fig. 1B (compare Fig. 6A and 1B.)

Similarly, the morphogenic activity of OP-1

purified from mammary gland extract was evaluated in vitro by measuring alkaline phosphatase activity in vitro using the following assay. Test samples were prepared using 15-20% of individual immunoreactive fractions from the C-18 run which were precipitated and resuspended in a smaller volume of 50% acetonitrile/0.1% TFA. Alkaline phosphatase activity was tested using ROS 17/2.8 cells (Rat Osteosarcoma, e.g., obtained, for example, from Dr. Robert J. Majeska, Mt. Sinai Medical Center, New York, NY, in a

standard alkaline phosphatase activity assay (see, for example, U.S. Pat. No. 4,968,590). The results, presented in Fig. 6B, indicate that the immunoreactive fractions obtained from C-18-purified mammary gland 5 extract correspond with alkaline phosphatase activity in vitro (compare Fig. 6B and Fig. 1B.) In Fig. 6B solid bars represent assays performed with mammary gland-purified OP-1, where the fraction numbers correspond to the related fractions eluted from the 10 C-18 reverse phase column (see Fig. 1B), the hatched bar represents the assay performed with purified, recombinantly-produced OP-1 (100ng/ml), and the cross-hatched bar represents background. As for Fig. 6A, alkaline phosphatase activity corresponds with 15 immunoreactivity of the C-18-purified extract (compare Fig. 6B and 1B.)

# Example 2. Morphogen Identification in Human Serum

OP-1 was detected in human serum using the 20 following assay. A monoclonal antibody raised against mammalian, recombinantly produced OP-1 using standard immunology techniques well described in the art and described generally in Example 8, was immobilized by passing the antibody over an activated agarose gel (e.q., Affi-Gel<sup>™</sup>, from Bio-Rad Laboratories, Richmond, CA, prepared following manufacturer's instructions), and used to purify OP-1 from serum. Human serum then was passed over the column and eluted with 3M 30 K-thiocyanate. K-thiocyanante fractions then were dialyzed in 6M urea, 20mM PO, pH 7.0, applied to a C8 HPLC column, and eluted with a 20 minute, 25-50% acetonitrile/0.1% TFA gradient. Mature, recombinantly produced OP-1 homodimers elute between 20-22 minutes.

Fractions then were collected and tested for the presence of OP-1 by standard immunoblot. Fig. 7 is an immunoblot showing OP-1 in human sera under reducing and oxidized conditions. In the figure, lanes 1 and 4 are OP-1 standards, run under oxidized (lane 1) and reduced (lane 4) conditions. Lane 5 shows molecular weight markers at 17, 27 and 39 kDa. Lanes 2 and 3 are human sera OP-1, run under oxidized (lane 2) and reduced (lane 3) conditions.

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## Example 3. Morphogen-Induced CAM Expression

The morphogens described herein induce CAM
expression as part of their induction of morphogenesis.

15 CAMs are morphoregulatory molecules identified in all
tissues as an essential step in tissue development. NCAMs, which comprise at least 3 isoforms (N-CAM-180, NCAM-140 and N-CAM-120, where "180", "140" and "120"
indicate the apparent molecular weights of the isoforms

20 as measured by polyacrylamide gel electrophoresis) are
expressed at least transiently in developing tissues,
and permanently in nerve tissue. Both the N-CAM-180 and
N-CAM-140 isoforms are expressed in both developing and
adult tissue. The N-CAM-120 isoform is found only in
25 adult tissue. Another neural CAM is L1.

CAMs are implicated in normal tissue development;
N-CAMs are implicated in appropriate neural
development, including appropriate neurulation,
neuronal migration, fasciculation, and synaptogenesis.
Inhibition of N-CAM production, as by complexing the
molecule with an N-CAM-specific antibody, inhibits
retina organization, including retinal axon migration,
and axon regeneration in the peripheral nervous system,

as well as axon synapsis with target muscle cells.

CAMs also have been postulated as part of a
morphoregulatory pathway whose activity is induced by a
to date unidentified molecule (See, for example,

Edelman, G.M. (1986) Ann. Rev. Cell Biol. 2:81-116).

Without being limited to any given theory, the
morphogens described herein may act as the inducer of
this pathway.

The morphogens described herein can stimulate CAM production. As described below, the morphogens stimulate L1 and N-CAM production, including all three isoforms of the N-CAM molecule, in nerve tissue.

In this example NG108-15 cells were cultured for 15 4 days in the presence of increasing concentrations of OP-1 and standard Western blots performed on whole cells extracts. The NG10875 cell line is a hybrid cell line (neuroblastoma x glioma, American Type Culture 20 Collection, Rockville, MD). N-CAM isoforms were detected with an antibody which crossreacts with all three isoforms, mAb H28.123, obtained from Sigma Chemical Co., St. Louis, the different isoforms being distinguishable by their different mobilities on an 25 electrophoresis gel. Control NG108-15 cells (untreated) express both the 140 kDa and the 180 kDa isoforms, but not the 120 kDa, as determined by western blot analyses using up to 100  $\mu g$  of protein. As showning Fig.8, treatment of NG108-15 cells with OP-1 resulted in a dose-dependent increase in the expression of the 30 180 kDa and 140 kDa isoforms, as well as the induction of the 120 kDa isoform. Fig. 8B is a Western blot of OP1-treated NG108-15 cell extracts, probed with mAb H28.123, showing the induction of all three isoforms.

Fig. 8A is a dose response curve of N-CAM-180 and N-CAM-140 induction as a function of morphogen concentration. N-CAM-120 is not shown in the graph as it could not be quantitated in control cells. However, 5 as is clearly evident from the Western blot in Fig. 8A, N-CAM-120 is induced in response to morphogen treatment. The induction of the 120 isoform also indicates that morphogen-induced redifferentiation of transformed cells stimulates not only redifferentiation 10 of these cells from a transformed phenotype, but also differentiation to a phenotype corresponding to a developed cell. The differential induction of N-CAM 180 and 140 isoforms seen may be because constitutive expression of the 140 isoform is close to maximum. addition, the increase in N-CAM expression corresponded in a dose-dependent manner with the morphogen induction of multicellular aggregates.

In addition, the cell aggregation effects of OP-1

20 on NG108-15 cells can be inhibited with anti-N-CAM
antibodies or antisense N-CAM oligonucleotides.
Antisense oligonucleotides can be made synthetically on
a nucleotide synthesizer, using standard means known in
the art. Preferably, phosphorothioate oligonucleotides

25 ("S-oligos") are prepared, to enhance transport of the
nucleotides across cell membranes. Concentrations of
both N-CAM antibodies and N-CAM antisense
oliognucleotides sufficient to inhibit N-CAM induction
also inhibited formation of multilayered cell

30 aggregates. Specifically, incubation of morphogentreated NG108-115 cells with 0.3-3 \(\mu\)M N-CAM antisense
S-oligos, 5-500 \(\mu\)M unmodified N-CAM antisense oligos,

or 10  $\mu$ g/ml mAb H28.123 significantly inhibits cell aggregation. It is likely that morphogen treatment also stimulates other CAMs, as inhibition is not complete.

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The experiments also have been performed with soluble morphogen (e.g., mature OP-1 associated with its pro domain) which also specifically induced CAM expression.

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# Example 4. <u>Effect of Morphogen Neutralization</u> on Embryogenesis

As described in Example 7, below, at least one
15 morphogen, OP2, is found principally in early
developing embryos (8-day embryos). As described
below, morphogen neutralization with morphogen-specific
antibodies inhibits embryogenesis.

Morphogen inhibition in developing embryos inhibits 20 tissue and organ development. Specifically, 9-day mouse embryo cells, cultured in vitro under standard culturing conditions, were incubated in the presence and absence of an OP-1-specific monoclonal antibody prepared using recombinantly produced, purified mature 25 OP-1 as the immunogen. The antibody was prepared using standard antibody production means well known in the art and essentially as described for Example 9, below. After two days, the effect of the antibody on the 30 developing embryo was evaluated by histology using standard histology procedures well known in the art. As determined by histological examination, the OP-1-specific antibody specifically inhibits eye lobe formation in the developing embryo. In particular, the

diencephalon outgrowth does not develop. In addition, the heart is malformed and enlarged. Moreover, in separate immunolocalization studies on embryo sections with labelled OP-1 specific antibody, the OP-1-specific antibody localizes to neural epithelia.

Similarly, morphogen activity may be demonstated in fetal development in the mouse model using the following assay. Single lip injections comprising  $10-100\mu g/injection$  of morphogen-specific antibody are administered to pregnant female mice during each day of the gestation period and tissue development (e.g., bone development) in treated and control new mice evaluated by standard histomorphometric analysis at birth.

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Finally, stimulation of endogenous morphogen antibody production in egg-laying hens interferes with shell formation in the developing eggs.

20 All of these data demonstate that inhibition of morphogen activity significantly interferes with tissue development during embryogenesis.

# Example 5. Effect of Morphogen Neutralization on Juvenile Tissue Development

The effect of the morphogens described herein on tissue development in developing mammals also may be demonstrated using neutralizing antibodies specific for particular morphogens and assessing the effect of these antibodies on tissue development as described below. Specifically, anti-morphogen monoclonal and/or polyclonal antibodies may be prepared using standard methodologies including, for example, the protocol

provided in Example 8, below, and provided to juveniles to inhibit the activity of endogenous morphogens.

Generally, purified antibodies are provided

regularly to new born mice, e.g., 10100µg/injection/day for 10-15 days. At 10 or 21 days,
the mice are sacrificed and the effect of morphogen on
bone development assessed by body weight, gross visual
examination and histology. In this example, anti-OP-1
antibodies were used in 10µg injections/day for 14
days, and the mice were sacrificed at 21 days. As is
dramatically demonstrated in Fig. 9, mice treated with
OP-1 specific antibody show consistent and significant
stunted growth, including reduced body length and body
weight, (9B) as compared with untreated mice (9A).
Histological examination showed reduced bone growth as
evidenced by reduced bone size in the treated mice.

In a variation on this protocol, single lip injections also may be provided to older juveniles and adult mice (e.g.,  $10-100~\mu g$ ) over a prolonged time (e.g., 10-15~days) to evaluate the effect or morphogen neutralization on bone growth and bone integrity and to evaluate the onset of osteoporosis.

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#### Example 6. Morphogen Treatment of Osteoporsis

## 6.1 <u>Effect of Morphogen on Trabecular Bone in</u> Ovariectomized (OVX) <u>Rats</u>

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Aged individuals, and particularly postmenopausal women are particularly at risk for osteoporosis. Provided below is an animal osteoposis model demonstrating the ability of morphogens to subtantially

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inhibit and/or reduce the tissue damage effects associated with osteoporosis, wherein osteoporosis is induced by ovary removal in rats. Bone growth is evaluated in these animals by measuring serum alkaline phosphataseans osteocalcin levels in treated an untreated rats.

Forty Long-Evans rats (Charles River Laboratories, Wilmington) weighing about 200g each are ovariectomized (OVX) using standard surgical procedures, and ten rats 10 are sham-operated. The ovariectomization of the rats produces an osteoporotic condition within the rats as a result of decreased estrogen production. Food and water are provided ad libitum. Eight days after ovariectomy, the rats, prepared as described above, were divided into five groups: (A), 10 sham-operated rats; (B), 10 ovariectomized rats receiving 1 ml of phosphate-buffered saline (PBS) i.v. in the tail vein; (C) 10 ovariectomized rats receiving about 1 mg of  $17\beta E_2$  (17- $\beta$ -estradiol  $E_2$ ) by intravenous injection 20 through the tail vein; (D) 9 ovariectomized rats receiving daily injections of approximately  $2\mu g$  of morphogen by tail vein for 22 days; and (E) 9 ovariectomized rats receiving daily injections of approximately 20  $\mu$ g of morphogen by tail vein for 25 22 days. In this example, OP-1 was the morphogen tested.

On the 15th and 21st day of the study, each rat was injected with 5 mg of tetracycline, and on day 22, the rats were sacrificed. The body weights, uterine

weights, serum alkaline phosphate levels, serum calcium levels and serum osteocalcin levels then were determined for each rat. The results are shown in Tables III and IV.

5 <u>Table III</u>

# Body Weights, Uterine Weights and Alkaline Phosphatase

	Group	Body Weights	Uterine Weights	Alk. Phosphatase
10		(g)	(g)	(U/L)
	A-SHAM	$250.90 \pm 17.04$	$0.4192 \pm 0.10$	$43.25 \pm 6.11$
	B-OVX+PBS	$273.40 \pm 16.81$	$0.1650 \pm 0.04$	$56.22 \pm 6.21$
	C-OVX+E2	241.66 <u>+</u> 21.54	$0.3081 \pm 0.03$	$62.66 \pm 4.11$
	D-0VX+0P-1	$266.67 \pm 10.43$	$0.1416 \pm 0.03$	$58.09 \pm 12.97$
15	(2µg)			
	E-0VX+0P-1	$272.40 \pm 20.48$	$0.1481 \pm 0.05$	66.24 <u>+</u> 15.74
	(20 µg)			

### TABLE IV

20

# Serum Calcium and Serum Osteocalcin Levels

	Group	Serum Calcium	Serum Osteocalcin
	-	(ng/dl)	(ng/ml)
25			
	A-SHAM	$8.82 \pm 1.65$	64.66 <u>+</u> 14.77
	B-OVX+PBS	8.95 <u>+</u> 1.25	$69.01 \pm 10.20$
	C-OVX+E2	9.20 <u>+</u> 1.39	67.13 <u>+</u> 17.33
	D-0VX+0P-1	$8.77 \pm 0.95$	$148.50 \pm 84.11$
30	(2µg)		
	E-OVX+OP-1	8.67 <u>+</u> 1.94	182.42 <u>+</u> 52.11
	(20µg)		

The results presented in Table III and IV show that intravenous injection of morphogen into ovariectomized rats produces a significant increase in serum alkaline phosphatase and serum osteocalcin levels and demonstrates that systemic administration of the morphogen stimulates bone formation in osteoporotic bone.

# 6.2 <u>Histomorphometric Analysis of Morphogen on the</u> 10 <u>Tibia Diaphysis in Ovariectomized(OVX) Rats</u>

Fifteen female Long-Evans rats weighing about 160 g were ovariectomized (OVX) to produce an osteoporotic condition and five rats were sham operated (Charles River Laboratories, Wilmington, MA.) as described for Example 8. Food and water were provided ad libitum. Twenty-two days after ovariectomy, the rats were divided into four groups: (A) sham-operated (1 ml of PBS by intravenous injection through tail vein (5 rats); (B) OVX, into which nothing was injected (5 rats); (C) OVX, receiving about 1 mg of 17βE<sub>2</sub> by intravenous injection through the tail vein (5 rats), and (D) OVX, receiving about 1 μg of morphogen by intravenous injection through the tail vein (5 rats). In this example, OP-1 was morphogen tested.

The rats were injected daily as described for seven days, except no injections were given on the thirteenth day. The rats then were sacrificed on the nineteenth day. The tibial diaphyseal long bones then were removed and fixed in ethanol and histomorphometric analysis was carried out using standard procedures well known in the art. The results are shown in Table V.

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	Table V														
	5 Longitudinal Growth 20.2 ± 0.3 19.4 ± 0.2 4.9 ± 0.5 17.9 ± 0														
	MEASUREMENT	• •		OVX + E <sub>2</sub>	OVX + OP-1										
5	Longitudinal Growth Rate (µm/day)	20.2 <u>+</u> 0.3	19.4 ± 0.2	4.9 <u>+</u> 0.5	$17.9 \pm 0.9$										
	Cancellous Bone Volume (BV/TV,	<del>-</del>	13.0 ± 1.6	13.7 <u>+</u> 2.1	16.6 +_1.8										
10	bone vol/total vol	)													
	Cancellous Bone Perimeter (mm)	16.2 <u>+</u> 1.8	9.6 <u>+</u> 0.9	11.5 ± 1.1	12.2 ± 0.7										
15	Labeled Cancellous Perimeter (%)	35.5 <u>+</u> 1.5	51.9 <u>+</u> 5.6	58.0 <u>+</u> 4.2	39.2 <u>+</u> 1.9										
	Mineral Apposition Rate (µm/day)	1.76 ± 0.14	2.25 <u>+</u> 0.16	1.87 ± 0.08	1.86 ± 0.20										

20

The results presented in Table V confirm the results of Example 6.1, namely that intravenous injection of OP-1 into ovariectomized rats stimulates bone growth for bone which had been lost due to the drop in estrogen within the individual rat. Specifically, the inhibition of cancellous bone volume in OVX rats is repaired by the systemically provided morphogen. In addition, in morphogen-treated rats the labelled cancellous perimeter and mineral apposition rate now return to levels measured in the control, sham-operated rats. Moreover, morphogen treatment does not inhibit longitudinal bone growth, unlike estrogen

treatment, which appears to inhibit bone growth significantly. Accordingly, systemic administration of a morphogen in therapeutically effective concentations effectively inhibits loss of bone mass in a mammal without inhibiting natural bone formation.

# Example 7. <u>Identification of Morphogen-Expressing</u> Tissue

Determining the tissue distribution of morphogens 10 may be used to identify different morphogens expressed in a given tissue, as well as to identify new, related Tissue distribution also may be used to morphogens. identify useful morphogen-producing tissue for use in 15 screening and identifying candidate morphogenstimulating agents. The morphogens (or their mRNA transcripts) readily are identified in different tissues using standard methodologies and minor modifications thereof in tissues where expression may 20 be low. For example, protein distribution may be determined using standard Western blot analysis or immunofluorescent techniques, and antibodies specific to the morphogen or morphogens of interest. Similarly, the distribution of morphogen transcripts may be 25 determined using standard Northern hybridization protocols and transcript-specific probes.

Any probe capable of hybridizing specifically to a transcript, and distinguishing the transcript of

30 interest from other, related transcripts may be used.

Because the morphogens described herein share such high sequence homology in their active, C-terminal domains, the tissue distribution of a specific morphogen transcript may best be determined using a probe

specific for the pro region of the immature protein and/or the N-terminal region of the mature protein. Another useful sequence is the 3' non-coding region flanking and immediately following the stop codon. 5 These portions of the sequence vary substantially among the morphogens of this invention, and accordingly, are specific for each protein. For example, a particularly useful Vgr-1-specific probe sequence is the PvuII-SacI fragment, a 265 bp fragment encoding both a portion of 10 the untranslated pro region and the N-terminus of the mature sequence (see Lyons et al. (1989) PNAS 86:4554-4558 for a description of the cDNA sequence). Similarly, particularly useful mOP-1-specific probe sequences are the BstX1-BglI fragment, a 0.68 Kb sequence that covers approximately two-thirds of the 15 mOP-1 pro region; a StuI-StuI fragment, a 0.2 Kb sequence immediately upstream of the 7-cysteine domain; and the Earl-Pstl fragment, an 0.3 Kb fragment containing a portion of the 3'untranslated sequence 20 (See Seq. ID No. 18, where the pro region is defined essentially by residues 30-291.) Similar approaches may be used, for example, with hOP-1 (Seq. ID No. 16) or human or mouse OP-2 (Seq. ID Nos. 20 and 22.)

Using these morphogen-specific probes, which may be synthetically engineered or obtained from cloned sequences, morphogen transcripts can be identified in mammalian tissue, using standard methodologies well known to those having ordinary skill in the art.

30 Briefly, total RNA is prepared from various adult murine tissues (e.g., liver, kidney, testis, heart, brain, thymus and stomach) by a standard methodology such as by the method of Chomczyaski et al. ((1987) Anal. Biochem 162:156-159) and described below. Poly

(A)+ RNA is prepared by using oligo (dT)-cellulose chromatography (e.g., Type 7, from Pharmacia LKB Poly (A)+ RNA (generally 15  $\mu$ g) Biotechnology, Inc.). from each tissue is fractionated on a 1% agarose/formaldehyde gel and transferred onto a Nytran membrane (Schleicher & Schuell). Following the transfer, the membrane is baked at 80°C and the RNA is cross-linked under UV light (generally 30 seconds at 1 mW/cm2). Prior to hybridization, the appropriate probe is denatured by heating. The hybridization is carried 10 out in a lucite cylinder rotating in a roller bottle apparatus at approximately 1 rev/min for approximately 15 hours at 37°C using a hybridization mix of 40% formamide, 5 x Denhardts, 5 x SSPE, and 0.1% SDS. 15 Following hybridization, the non-specific counts are washed off the filters in 0.1 x SSPE, 0.1% SDS at 50°C.

Examples demonstrating the tissue distribution of various morphogens, including Vgr-1, OP-1, BMP2, BMP3, BMP4, BMP5, GDF-1, and OP-2 in developing and adult 20 tissue are disclosed in international application US92/01968 (WO92/15323), and in Ozkaynak, et al., (1991) Biochem. Biophys. Res. Commn. 179:116-123, and Ozkaynak, et al. (1992) (<u>J. Biol. Chem.</u> <u>267</u>: 25220-25 25227), the disclosures of which are incorporated herein by reference. Using the general probing methodology described herein, northern blot hybridizations using probes specific for these morphogens to probe brain, spleen, lung, heart, liver 30 and kidney tissue indicate that kidney-related tissue appears to be the primary expression source for OP-1, with brain, heart and lung tissues being secondary sources. Lung tissue appears to be the primary tissue expression source for Vgr-1, BMP5, BMP4 and BMP3. Lower

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levels of Vgr-1 also are seen in kidney and heart tissue, while the liver appears to be a secondary expression source for BMP5, and the spleen appears to be a secondary expression source for BMP4. GDF-1 appears to be expressed primarily in brain tissue.

Of particular relevance to the present application, OP-1 also is detected in adult rat stomach and gut tissue. Moreover, OP-2 appears to be expressed primarily in early embryonic tissue. Specifically, northern blots of murine embryos and 6-day post-natal animals shows abundant OP2 expression in 8-day embryos. Expression is reduced significantly in 17-day embryos and is not detected in post-natal animals.

15

In addition, labelled soluble OP-1 (iodinated with 125 I, using standard labelling procedures well known in the art) and injected into the rat tail vein also is localized to the stomach tissue within 30 minutes of injection.

# Example 8. <u>Detecting Morphogenic Protein in Solution</u> <u>by Immunoassay</u>

Morphogens are readily detected in solution with a standard immunoassay, using a polyclonal or monoclonal antibody specific for that protein and standard Western blot, ELISA (enzyme-linked immunoabsorbant assay) or other immunoassay technique well known in the art. A currently preferred, exemplary protocol for an ELISA assay, as well as means for generating morphogen-specific antibody are presented below. Standard protocols for antibody production, Western blot and other immunoassays also are described, for example, in

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Molecular Cloning A Laboratory Manual, Sambrook et al., eds. 1989, Cold Spring Harbor Press, Cold Spring Harbor, NY. Standard ELISA technique is described, for example, by Engvall (1980) Methods Enzymol. 70:419-439.

8.1 Morphogen-Specific Antiserum

Polyclonal antibody was prepared as follows. Each rabbit was given a primary immunization of 100 ug/500 10 μl E. coli-produced OP-1 monomer (amino acids 328-431 in SEQ ID NO:5) in 0.1% SDS mixed with 500 μl Complete Freund's Adjuvant. The antigen was injected subcutaneously at multiple sites on the back and flanks of the animal. The rabbit was boosted after a month in the same manner using incomplete Freund's Adjuvant. Test bleeds are taken from the ear vein seven days later. Two additional boosts and test bleeds were performed at monthly intervals until antibody against OP-1 was detected in the serum using an ELISA assay.

20 Then, the rabbit was boosted monthly with 100 μg of antigen and bled (15 ml per bleed) at days seven and ten after boosting.

## 8.2 Morphogen-Specific Antibody

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Monoclonal antibody specific for a given morphogen was prepared as follows. A mouse was given two injections of <u>E. coli</u> produced OP-1 monomer. The first injection contains  $100\mu g$  of OP-1 in complete Freund's adjuvant and was given subcutaneously. The second injection contained 50  $\mu g$  of OP-1 in incomplete adjuvant and was given intraperitoneally. The mouse then received a total of 230  $\mu g$  of OP-1 (amino acids 307-431 in SEQ ID NO:5) in four intraperitoneal

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injections at various times over an eight month period. One week prior to fusion, both mice were boosted intraperitoneally with 100  $\mu g$  of OP-1 (307-431) and 30  $\mu$ g of the N-terminal peptide (Ser293-Asn309-Cys) 5 conjugated through the added cysteine to bovine serum albumin with SMCC crosslinking agent. This boost was repeated five days (IP), four days (IP), three days (IP) and one day (IV) prior to fusion. The mouse spleen cells then were fused to commercially available 10 myeloma cells at a ratio of 1:1 using PEG 1500 (Boeringer Mannheim, Germany), and the cell fusion plated and screened for OP-1-specific antibodies using OP-1 (307-431) as antigen. The cell fusion and monoclonal screening then were according to standard 15 procedures well described in standard texts widely available in the art e.g., Maniatis et al. Molecular Cloning A Laboratory Manual, Cold Spring Harbor Press.

### 8.3 Morphogen ELISA

20

1 μg/100 μl of affinity-purified polyclonal rabbit
IgG specific for OP-1 was added to each well of a
96-well plate and incubated at 37°C for an hour. The
wells were washed four times with 0.167M sodium borate
25 buffer with 0.15 M NaCl (BSB), pH 8.2, containing 0.1%
Tween 20. To minimize non-specific binding, the wells
are blocked by filling completely with 1% bovine serum
albumin (BSA) in BSB and incubating for 1 hour at 37°C.
The wells are then washed four times with BSB
30 containing 0.1% Tween 20. A 100 μl aliquot of an
appropriate dilution of each of the test samples of
cell culture supernatant was added to each well in
triplicate and incubated at 37°C for 30 min. After
incubation, 100 μl biotinylated rabbit anti-OP-1 serum

(stock solution is about 1 mg/ml and diluted 1:400 in BSB containing 1% BSA before use) are added to each well and incubated at 37°C for 30 min. The wells were then washed four times with BSB containing 0.1% Tween 100  $\mu$ l strepavidin-alkaline (Southern Biotechnology Associates, Inc. Birmingham, Alabama, diluted 1:2000 in BSB containing 0.1% Tween 20 before use) was added to each well and incubated at 37°C for The plates were washed four times with 0.5M Tris buffered Saline (TBS), pH 7.2.  $50\mu$ l substrate (ELISA Amplification System Kit, Life Technologies, Inc., Bethesda, MD) was added to each well incubated at room temperature for 15 min. Then, 50  $\mu$ l amplifier (from the same amplification system kit) is added and incubated for another 15 min at room temperature. 15 reaction was stopped by the addition of 50  $\mu$ l 0.3 M sulphuric acid. The OD at 490 nm of the solution in each well was recorded. To quantitate OP-1 in culture media, an OP-1 standard curve was performed in parallel 20 with the test samples.

The invention may be embodied in other specific forms without departing from the spirit or essential characteristics thereof. The present embodiments are therefore to be considered in all respects as illustrative and not restrictive, the scope of the invention being indicated by the appended claims rather than by the foregoing description, and all changes which come within the meaning and range of equivalency of the claims are therefore intended to be embraced therein.

- 95 -

## SEQUENCE LISTING

5	(1) GENERAL INFORMATION:	
10 15	(i) APPLICANT:  (A) NAME: CREATIVE BIOMOLECULES, INC.  (B) STREET: 35 SOUTH STREET  (C) CITY: HOPKINTON  (D) STATE: MA  (E) COUNTRY: USA  (F) POSTAL CODE (ZIP): 01748  (G) TELEPHONE: 1-508-435-9001  (H) TELEFAX: 1-508-435-0454  (I) TELEX:	
	(ii) TITLE OF INVENTION: MORPHOGEN-ENRICHED DIETARY COMP	OSITION
20	(iii) NUMBER OF SEQUENCES: 33	
25	(iv) CORRESPONDENCE ADDRESS:  (A) ADDRESSEE: CREATIVE BIOMOLECULES, INC.  (B) STREET: 35 SOUTH STREET  (C) CITY: HOPKINTON  (D) STATE: MA  (E) COUNTRY: USA  (F) ZIP: 01748	
30	<ul> <li>(v) COMPUTER READABLE FORM:</li> <li>(A) MEDIUM TYPE: Floppy disk</li> <li>(B) COMPUTER: IBM PC compatible</li> <li>(C) OPERATING SYSTEM: PC-DOS/MS-DOS</li> <li>(D) SOFTWARE: PatentIn Release #1.0, Version #1.25</li> </ul>	
35	(viii) ATTORNEY/AGENT INFORMATION:  (A) NAME: KELLEY, ROBIN D.  (B) REGISTRATION NUMBER: 34,637  (C) REFERENCE/DOCKET NUMBER: CRP-071	
40	(ix) TELECOMMUNICATION INFORMATION:  (A) TELEPHONE: 617/248-7000  (B) TELEFAX: 617/248-7100	
45	(2) INFORMATION FOR SEQ ID NO:1:	
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(ii)	MOLECULE	TYPE:	protein
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5 10	(ix)		NAI LO	IE/KI CATI( IER I /not ONE	ON: 1 INFOI te= ' OF 1	L97 RMATI WHEE	ON: REIN ON	EACH TURA	I XAA	A INI -OCCU	EPEN JRIN(	IDENT	CLY ]	INDIO	CATES	S CNO
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35				70D 4		- N										

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  (B) TYPE: amino acid

  (C) STRANDEDNESS: single

  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein 45

- 97 -

5	(	ix)	(A)	LOC	IE/KI CATIO IER I /not ONE	EY: H ON: 1 INFOR Ce= ' OF T	L97 RMATI WHER THE 2	7 EON: REIN 20 NA	EACI TURA	I XAA	A INI OCCI	)EPEN JRIN(	IDENT	LTA 1	INDIC ER A-	CATES	10 2
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5		(D	, 011	/not	ce= ' 1 A (	'WHEI GROUI	REIN P OF	EAC! ONE	AAX I	A IS	SPE	EPENI CIFII	DENTI	LY SI	ACII	ED S
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40		(ii)	` '														
45		(vi)	ORIO	GINAI OR(	L SOI	_	: Homo	sap:		JS							

5	(ix)	(B	) NAI ) LO(	IE/KI CATI(	ON: 1	113	39	/lal	oel=	hOP1	L-MAT	TURE				
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	(1-)	(A	) OR	GANI	SM:	MURI		n								

5	(	(ix)	(A) (B)	NAM LOC	E/KE CATIONER I	N: 1	13	19	/laì	el=	HOP 1	L-MAT	TURE				
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45		(ii) (vi)	ORI	GINA ) OR		URCE SM:	: HOMO	SAP	IENS Camp	IIS.							
			(r	, 11	SOUP	TIL	r: U	TT I O	OUIL	JU							

5		(A) NAME/KEY: Protein (B) LOCATION: 1139 (D) OTHER INFORMATION: /label= HOP2-MATURE															
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45		(vi)	(A	) OR	GANI	URCE SM: 1	MURI		0	٠							

5		(ix)	(A) (B)	TURE: ) NAI ) LOG ) OTI	ME/KI	ON: C	113	39	/lal	bel=	MOP	2-MA'	rure				
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		(vi)		GINA:				nae									

5	(i	x)	(A)	LOC	IE/KI	EY: F ON: 1 INFOI	110	)1	/lal	oel=	СВИІ	?-2A-	-FX					
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- 105 -

	(xi	) SE	QUENC	E DE	SCRI	PTIO	N: S	EQ II	ONO:	:10:						
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30	(ii	•	LECUI													
35	`	.) OR	IGINA A) OF	L SO	URCE	:		LA M	ELAN	OGAS'	TER					
40	Ki)	(	ATURE A) NA B) LO D) OI	ME/K CATI	ON:	11	01	/la	bel=	DPP	-FX					
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25	(ii) MOLECULE TYPE: protein																
25	(vi) ORIGINAL SOURCE: (A) ORGANISM: XENOPUS																
30	<pre>(ix) FEATURE:     (A) NAME/KEY: Protein     (B) LOCATION: 1102     (D) OTHER INFORMATION: /label= VGL-FX</pre>																
35		(xi)	SEQ	UENC	E DE	SCRI	PTIO	N: S	EQ I	D NO	:12:					•	
		Cys 1	Lys	Lys	Arg	His 5	Leu	Tyr	Val	Glu	Phe 10	Lys	Asp	Val	Gly	Trp 15	Gln
40		Asn	Trp	Val	Ile 20	Ala	Pro	Gln	Gly	Tyr 25	Het	Ala	Asn	Tyr	Cys 30	Tyr	Gly
45		Glu	Cys	Pro 35	Tyr	Pro	Leu	Thr	Glu 40	Ile	Leu	Asn	Gly	Ser 45	Asn	His	Ala
45		Ile	Leu 50	Gln	Thr	Leu	Val	His 55	Ser	Ile	Glu	Pro	Glu 60	Asp	Ile	Pro	Let
EΛ		Pro	Cys	Cys	Val	Pro	Thr	Lys	Met	Ser	Pro	Ile	Ser	Met	Leu	Phe	Ту: 80

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Asp Asn Asn Asp Asn Val Val Leu Arg His Tyr Glu Asn Met Ala Val Asp Glu Cys Gly Cys Arg 100 5 (2) INFORMATION FOR SEQ ID NO:13: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 102 amino acids 10 (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 15 (ii) MOLECULE TYPE: protein (vi) ORIGINAL SOURCE: (A) ORGANISM: MURIDAE (ix) FEATURE: 20 (A) NAME/KEY: Protein (B) LOCATION: 1..102 (D) OTHER INFORMATION: /label= VGR-1-FX 25 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13: Cys Lys Lys His Glu Leu Tyr Val Ser Phe Gln Asp Val Gly Trp Gln 30 Glu Cys Ser Phe Pro Leu Asn Ala His Met Asn Ala Thr Asn His Ala 35 Ile Val Gln Thr Leu Val His Val Met Asn Pro Glu Tyr Val Pro Lys 50 Pro Cys Cys Ala Pro Thr Lys Val Asn Ala Ile Ser Val Leu Tyr Phe 40 Asp Asp Asn Ser Asn Val Ile Leu Lys Lys Tyr Arg Asn Met Val Val 45 Arg Ala Cys Gly Cys His 100

	(2) INFOR	MATION 1	FOR S	EQ I	D NO	:14:									
5	(i)	SEQUENCE (A) LEI (B) TYI (C) STI (D) TOI	NGTH: PE: a: RANDE	106 mino DNES	ami aci S: s	no a d ingl	.cids								
10	(ii)	MOLECUL	E TYP	E: p	rote	in									
10	(iii)	HYPOTHE	TICAL	: NO											
	(i∇)	ANTI-SE	NSE:	NO											
15	(vi)	ORIGINA (A) OR (F) TI	GANIS	M: H	omo	sapi ain	.ens								
20	(ix)	FEATURE (A) NA (B) LO (D) OT	ME/KE CATIO	N: 1	10	)6	/not	:e= "	'GDF-	-1 (f	(x)"				
25	(xi)	SEQUENC	E DES	CRIP	TION	l: SI	EQ II	) NO:	14:						
	Cys 1	Arg Ala	Arg	Arg 5	Leu	Tyr	Val	Ser	Phe 10	Arg	Glu	Val	Gly	Trp 15	His
30	Arg	Trp Val	Ile 20	Ala	Pro	Arg	Gly	Phe 25	Leu	Ala	Asn	Tyr	Cys 30	Gln	Gly
	Gln	Cys Ala	Leu	Pro	Val	Ala	Leu 40	Ser	Gly	Ser	Gly	Gly 45	Pro	Pro	Ala
35	Leu	Asn His	Ala	Val	Leu	Arg 55	Ala	Leu	Met	His	Ala 60	Ala	Ala	Pro	Gly
40	Ala 65	Ala Asp	Leu	Pro	Cys 70	Cys	Val	Pro	Ala	Arg 75	Leu	Ser	Pro	Ile	Ser 80
	Val	Leu Phe	Phe	Asp 85	Asn	Ser	Asp	Asn	Val 90	Val	Leu	Arg	Gln	Tyr 95	Glu
45	Asp	Met Val	Val 100	Asp	Glu	Cys	Gly	Cys 105	Arg						

(2) INFORMATION FOR SEQ ID NO:15:

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(i) SEQUENCE CHARACTERISTICS:
               (A) LENGTH: 5 amino acids
               (B) TYPE: amino acid
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               (C) STRANDEDNESS: single
               (D) TOPOLOGY: linear
        (ii) MOLECULE TYPE: peptide
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        (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:
         Cys Xaa Xaa Xaa Xaa
15
    (2) INFORMATION FOR SEQ ID NO:16:
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         (i) SEQUENCE CHARACTERISTICS:
               (A) LENGTH: 1822 base pairs
               (B) TYPE: nucleic acid
               (C) STRANDEDNESS: single (D) TOPOLOGY: linear
25
        (ii) MOLECULE TYPE: cDNA
       (iii) HYPOTHETICAL: NO
30
        (iv) ANTI-SENSE: NO
        (vi) ORIGINAL SOURCE:
               (A) ORGANISM: HOMO SAPIENS
               (F) TISSUE TYPE: HIPPOCAMPUS
35
        _(ix) FEATURE:
               (A) NAME/KEY: CDS
               (B) LOCATION: 49..1341
               (C) IDENTIFICATION METHOD: experimental
               (D) OTHER INFORMATION: /function= "OSTEOGENIC PROTEIN"
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                      /product= "OP1"
                      /evidence= EXPERIMENTAL
                      /standard name= "OP1"
45
         (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:
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	GGT	CGGC	CC (	CGGAC	CCCG	G AC	CCC	GGTA	GCG	CGTA	LGAG	CCGG	CGCG		: His	GTG Val	57
5	CGC Arg	TCA Ser 5	CTG Leu	CGA Arg	GCT Ala	GCG Ala	GCG Ala 10	CCG Pro	CAC His	AGC Ser	TTC Phe	GTG Val 15	GCG Ala	CTC Leu	TGG Trp	GCA Ala	105
10	CCC Pro 20	CTG Leu	TTC Phe	CTG Leu	CTG Leu	CGC Arg 25	TCC Ser	GCC Ala	CTG Leu	GCC Ala	GAC Asp 30	TTC Phe	AGC Ser	CTG Leu	GAC Asp	AAC Asn 35	153
15	GAG Glu	GTG Val	CAC His	TCG Ser	AGC Ser 40	TTC Phe	ATC Ile	CAC His	CGG Arg	CGC Arg 45	CTC Leu	CGC Arg	AGC Ser	CAG Gln	GAG Glu 50	CGG Arg	201
20	CGG Arg	GAG Glu	ATG Met	CAG Gln 55	CGC Arg	GAG Glu	ATC Ile	CTC Leu	TCC Ser 60	ATT Ile	TTG Leu	GGC Gly	TTG Leu	CCC Pro 65	CAC His	CGC Arg	249
20	CCG Pro	CGC Arg	CCG Pro 70	CAC His	CTC Leu	CAG Gln	GGC Gly	.AAG Lys 75	CAC His	AAC Asn	TCG Ser	GCA Ala	CCC Pro 80	ATG Met	TTC Phe	ATG Met	297
25	CTG Leu	GAC Asp 85	CTG Leu	TAC Tyr	AAC Asn	GCC Ala	ATG Met 90	GCG Ala	GTG Val	GAG Glu	GAG Glu	GGC Gly 95	GGC Gly	GGG Gly	CCC Pro	GGC Gly	345
30	GGC Gly 100	CAG Gln	GGC Gly	TTC Phe	TCC Ser	TAC Tyr 105	CCC Pro	TAC Tyr	AAG Lys	GCC Ala	GTC Val 110	TTC Phe	AGT Ser	ACC Thr	CAG Gln	GGC Gly 115	393
35	CCC Pro	CCT Pro	CTG Leu	GCC Ala	AGC Ser 120	CTG Leu	CAA Gln	GAT Asp	AGC Ser	CAT His 125	TTC Phe	CTC Leu	ACC Thr	GAC Asp	GCC Ala 130	GAC Asp	441
40	ATG Met	GTC Val	ATG Met	AGC Ser 135	TTC Phe	GTC Val	AAC Asn	CTC Leu	GTG Val 140	GAA Glu	CAT His	GAC Asp	AAG Lys	GAA Glu 145	TTC Phe	TTC Phe	489
40	CAC His	CCA Pro	CGC Arg 150	TAC Tyr	CAC His	CAT His	CGA Arg	GAG Glu 155	TTC Phe	CGG Arg	TTT Phe	GAT Asp	CTT Leu 160	TCC Ser	AAG Lys	ATC Ile	537
45	CCA Pro	GAA Glu 165	GGG Gly	GAA Glu	GCT Ala	GTC Val	ACG Thr 170	GCA Ala	GCC Ala	GAA Glu	TTC Phe	CGG Arg 175	ATC Ile	TAC Tyr	AAG Lys	GAC Asp	585

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	TAC Tyr 180	ATC Ile	CGG Arg	GAA Glu	CGC Arg	TTC Phe 185	GAC Asp	AAT Asn	GAG Glu	ACG Thr	TTC Phe 190	CGG Arg	ATC Ile	AGC Ser	GTT Val	TAT Tyr 195	633
5	CAG Gln	GTG Val	CTC Leu	CAG Gln	GAG Glu 200	CAC His	TTG Leu	GGC Gly	AGG Arg	GAA Glu 205	TCG Ser	GAT Asp	CTC Leu	TTC Phe	CTG Leu 210	CTC Leu	681
10	GAC Asp	AGC Ser	CGT Arg	ACC Thr 215	CTC Leu	TGG Trp	GCC Ala	TCG Ser	GAG Glu 220	GAG Glu	GGC Gly	TGG Trp	CTG Leu	GTG Val 225	TTT Phe	GAC Asp	729
15	ATC Ile	ACA Thr	GCC Ala 230	ACC Thr	AGC Ser	AAC Asn	CAC His	TGG Trp 235	GTG Val	GTC Val	AAT Asn	CCG Pro	CGG Arg 240	CAC His	AAC Asn	CTG Leu	777
20	GGC Gly	CTG Leu 245	CAG Gln	CTC Leu	TCG Ser	GTG Val	GAG Glu 250	ACG Thr	CTG Leu	GAT Asp	GGG Gly	CAG Gln 255	AGC Ser	ATC Ile	AAC Asn	CCC Pro	825
20	AAG Lys 260	TTG Leu	GCG Ala	GGC Gly	CTG Leu	ATT Ile 265	GGG Gly	CGG Arg	CAC His	GGG Gly	CCC Pro 270	CAG Gln	AAC Asn	AAG Lys	CAG Gln	CCC Pro 275	873
25	TTC Phe	ATG Met	GTG Val	GCT Ala	TTC Phe 280	TTC Phe	AAG Lys	GCC Ala	ACG Thr	GAG Glu 285	GTC Val	CAC His	TTC Phe	CGC Arg	AGC Ser 290	ATC Ile	921
30	CGG Arg	TCC Ser	ACG Thr	GGG Gly 295	AGC Ser	AAA Lys	CAG Gln	CGC Arg	AGC Ser 300	CAG Gln	AAC Asn	CGC Arg	TCC Ser	AAG Lys 305	ACG Thr	CCC Pro	969
35	AAG Lys	AAC Asn	CAG Gln 310	GAA Glu	GCC Ala	CTG Leu	CGG Arg	ATG Met 315	GCC Ala	AAC Asn	GTG Val	GCA Ala	GAG Glu 320	AAC Asn	AGC Ser	AGC Ser	1017
40	AGC Ser	GAC Asp 325	CAG Gln	AGG Arg	CAG Gln	GCC Ala	TGT Cys 330	AAG Lys	AAG Lys	CAC His	GAG Glu	CTG Leu 335	TAT Tyr	GTC Val	AGC Ser	TTC Phe	1065
40	CGA Arg 340	GAC Asp	CTG Leu	GGC Gly	TGG Trp	CAG Gln 345	GAC Asp	TGG Trp	ATC Ile	ATC Ile	GCG Ala 350	CCT Pro	GAA Glu	GGC Gly	TAC Tyr	GCC Ala 355	1113
45	GCC Ala	TAC Tyr	TAC Tyr	TGT Cys	GAG Glu 360	GGG Gly	GAG Glu	TGT Cys	GCC Ala	TTC Phe 365	Pro	CTG Leu	AAC Asn	TCC Ser	TAC Tyr 370	ATG Met	1161

50

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	AAC GCC ACC AAC CAC GCC ATC GTG CAG ACG CTG GTC CAC TTC ATC AAC Asn Ala Thr Asn His Ala Ile Val Gln Thr Leu Val His Phe Ile Asn 375	1209
5	CCG GAA ACG GTG CCC AAG CCC TGC TGT GCG CCC ACG CAG CTC AAT GCC Pro Glu Thr Val Pro Lys Pro Cys Cys Ala Pro Thr Gln Leu Asn Ala 390 395	1257
10	ATC TCC GTC CTC TAC TTC GAT GAC AGC TCC AAC GTC ATC CTG AAG AAA Ile Ser Val Leu Tyr Phe Asp Asp Ser Ser Asn Val Ile Leu Lys Lys' 405	1305
15	TAC AGA AAC ATG GTG GTC CGG GCC TGT GGC TGC CAC TAGCTCCTCC Tyr Arg Asn Met Val Val Arg Ala Cys Gly Cys His 420 425 430	1351
	GAGAATTCAG ACCCTTTGGG GCCAAGTTTT TCTGGATCCT CCATTGCTCG CCTTGGCCAG	1411
	GAACCAGCAG ACCAACTGCC TTTTGTGAGA CCTTCCCCTC CCTATCCCCA ACTTTAAAGG	1471
20	TGTGAGAGTA TTAGGAAACA TGAGCAGCAT ATGGCTTTTG ATCAGTTTTT CAGTGGCAGC	1531
	ATCCAATGAA CAAGATCCTA CAAGCTGTGC AGGCAAAACC TAGCAGGAAA AAAAAACAAC	1591
25	GCATAAAGAA AAATGGCCGG GCCAGGTCAT TGGCTGGGAA GTCTCAGCCA TGCACGGACT	1651
	CGTTTCCAGA GGTAATTATG AGCGCCTACC AGCCAGGCCA CCCAGCCGTG GGAGGAAGGG	1711
	GGCGTGGCAA GGGGTGGGCA CATTGGTGTC TGTGCGAAAG GAAAATTGAC CCGGAAGTTC	1771
30	CTGTAATAAA TGTCACAATA AAACGAATGA ATGAAAAAAA AAAAAAAAA A	1822
	(2) INFORMATION FOR SEQ ID NO:17:	
35	<ul> <li>(i) SEQUENCE CHARACTERISTICS:         <ul> <li>(A) LENGTH: 431 amino acids</li> <li>(B) TYPE: amino acid</li> <li>(D) TOPOLOGY: linear</li> </ul> </li> </ul>	
<b>4</b> 0	(ii) MOLECULE TYPE: protein	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:	
<i>1</i> E	Met His Val Arg Ser Leu Arg Ala Ala Pro His Ser Phe Val Ala	
45	1 5 10 15	
	Leu Trp Ala Pro Leu Phe Leu Leu Arg Ser Ala Leu Ala Asp Phe Ser 20 25 30	

	Leu	Asp	Asn 35	Glu	Val	His	Ser	Ser 40	Phe	Ile	His	Arg	Arg 45	Leu	Arg	Ser
5	Gln	Glu 50	Arg	Arg	Glu	Met	Gln 55	Arg	Glu	Ile	Leu	Ser 60	Ile	Leu	Gly	Leu
	Pro 65	His	Arg	Pro	Arg	Pro 70	His	Leu	Gln	Gly	Lys 75	His	Asn	Ser	Ala	Pro 80
10	Met	Phe	Het	Leu	Asp 85	Leu	Tyr	Asn	Ala	<b>Met</b> 90	Ala	Val	Glu	Glu	<b>Gly</b> 95	Gly
15	Gly	Pro	Gly	Gly 100	Gln	Gly	Phe	Ser	Tyr 105	Pro	Tyr	Lys	Ala	Val 110	Phe	Ser
	Thr	Gln	Gly 115	Pro	Pro	Leu	Ala	Ser 120	Leu	Gln	Asp	Ser	His 125	Phe	Leu	Thr
20	Asp	Ala 130	Asp	Met	Val	Met	Ser 135	Phe	Val	Asn	Leu	Val 140	Glu	His	Asp	Lys
	Glu 145	Phe	Phe	His	Pro	Arg 150	Tyr	His	His	Arg	Glu 155	Phe	Arg	Phe	Asp	Leu 160
25	Ser	Lys	Ile	Pro	Glu 165	Gly	Glu	Ala	Val	Thr 170	Ala	Ala	Glu	Phe	Arg 175	Ile
30	Tyr	Lys	Asp	Tyr 180	Ile	Arg	Glu	Arg	Phe 185	Asp	Asn	Glu	Thr	Phe 190	Arg	Ile
30	Ser	Val	Tyr 195	Gln	Val	Leu	Gln	Glu 200	His	Leu	Gly	Arg	Glu 205	Ser	Asp	Let
35	Phe	Leu 210	Leu	Asp	Ser	Arg	Thr 215	Leu	Trp	Ala	Ser	Glu 220	Glu	Gly	Trp	Leu
	Val 225	Phe	Asp	Ile	Thr	Ala 230	Thr	Ser	Asn	His	Trp 235	Val	Val	Asn	Pro	Arg 240
40	His	Asn	Leu	Gly	Leu 245		Leu	Ser	Val	Glu 250	Thr	Leu	Asp	Gly	Gln 255	Ser
45	Ile	Asn	Pro	Lys 260		Ala	Gly	Leu	11e 265	Gly	Arg	His	Gly	Pro 270	Gln	Asr
<b>4</b> 0	Lys	Gln	Pro 275		Met	Val	Ala	Phe 280		Lys	Ala	Thr	Glu 285	Val	His	Phe
50	Arg	Ser		Arg	Ser	Thr	Gly 295	Ser	Lys	Gln	Arg	Ser 300	Gln	Asn	Arg	Set

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Lys Thr Pro Lys Asn Gln Glu Ala Leu Arg Met Ala Asn Val Ala Glu 305 Asn Ser Ser Ser Asp Gln Arg Gln Ala Cys Lys His Glu Leu Tyr 330 5 Val Ser Phe Arg Asp Leu Gly Trp Gln Asp Trp Ile Ile Ala Pro Glu Gly Tyr Ala Ala Tyr Tyr Cys Glu Gly Glu Cys Ala Phe Pro Leu Asn Ser Tyr Met Asn Ala Thr Asn His Ala Ile Val Gln Thr Leu Val His 380 15 Phe Ile Asn Pro Glu Thr Val Pro Lys Pro Cys Cys Ala Pro Thr Gln 385 Leu Asn Ala Ile Ser Val Leu Tyr Phe Asp Asp Ser Ser Asn Val Ile 20 Leu Lys Lys Tyr Arg Asn Met Val Val Arg Ala Cys Gly Cys His 430 420 (2) INFORMATION FOR SEQ ID NO:18: 25 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 1873 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single 30 (D) TOPOLOGY: linear (ii) MOLECULE TYPE: cDNA 35 (iii) HYPOTHETICAL: NO (iv) ANTI-SENSE: NO (vi) ORIGINAL SOURCE: 40 (A) ORGANISM: MURIDAE (F) TISSUE TYPE: EMBRYO (ix) FEATURE: (A) NAME/KEY: CDS 45 (B) LOCATION: 104..1393 (D) OTHER INFORMATION: /function= "OSTEOGENIC PROTEIN" /product= "MOP1" /note= "MOP1 (CDNA)" 50 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

WO 94/03075 PCT/US93/07190

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5	CGGC	CGCGC	GGC (	CCGGT	rgcco	CC GC	GATCO	GCGCC	TAC	GAGC	CGGC	GCG	ATG Met 1	CAC His	GTG Val	CGC Arg	115
	TCG Ser 5	CTG Leu	CGC Arg	GCT Ala	GCG Ala	GCG Ala 10	CCA Pro	CAC His	AGC Ser	TTC Phe	GTG Val 15	GCG Ala	CTC Leu	TGG Trp	GCG Ala	CCT Pro 20	163
10	CTG Leu	TTC Phe	TTG Leu	CTG Leu	CGC Arg 25	TCC Ser	GCC Ala	CTG Leu	GCC Ala	GAT Asp 30	TTC Phe	AGC Ser	CTG Leu	GAC Asp	AAC Asn 35	GAG Glu	211
15	GTG Val	CAC His	TCC Ser	AGC Ser 40	TTC Phe	ATC Ile	CAC His	CGG Arg	CGC Arg 45	CTC Leu	CGC Arg	AGC Ser	CAG Gln	GAG Glu 50	CGG Arg	CGG Arg	259
20	GAG Glu	ATG Met	CAG Gln 55	CGG Arg	GAG Glu	ATC Ile	CTG Leu	TCC Ser 60	ATC Ile	TTA Leu	GGG Gly	TTG Leu	CCC Pro 65	CAT His	CGC Arg	CCG Pro	307
25				CTC Leu													355
20				AAC Asn													403
30	GGC Gly	TTC Phe	TCC Ser	TAC Tyr	CCC Pro 105	TAC Tyr	AAG Lys	GCC Ala	GTC Val	TTC Phe 110	AGT Ser	ACC Thr	CAG Gln	GGC Gly	CCC Pro 115	CCT Pro	451
35	TTA Leu	GCC Ala	AGC Ser	CTG Leu 120	CAG Gln	GAC Asp	AGC Ser	CAT His	TTC Phe 125	CTC Leu	ACT Thr	GAC Asp	GCC Ala	GAC Asp 130	ATG Met	GTC Val	499
40				GTC Val													547
45	CGA Arg	TAC Tyr 150	CAC His	CAT His	CGG Arg	GAG Glu	TTC Phe 155	CGG Arg	TTT Phe	GAT Asp	CTT Leu	TCC Ser 160	AAG Lys	ATC Ile	CCC Pro	GAG Glu	<b>5</b> 95
50				GTG Val													643

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	CGG Arg	GAG Glu	CGA Arg	TTT Phe	GAC Asp 185	AAC Asn	GAG Glu	ACC Thr	TTC Phe	CAG Gln 190	ATC Ile	ACA Thr	GTC Val	TAT Tyr	CAG Gln 195	GTG Val	691
5	CTC Leu	CAG Gln	GAG Glu	CAC His 200	TCA Ser	GGC Gly	AGG Arg	GAG Glu	TCG Ser 205	GAC Asp	CTC Leu	TTC Phe	TTG Leu	CTG Leu 210	GAC Asp	AGC Ser	739
10	CGC Arg	ACC Thr	ATC Ile 215	TGG Trp	GCT Ala	TCT Ser	GAG Glu	GAG Glu 220	GGC Gly	TGG Trp	TTG Leu	GTG Val	TTT Phe 225	GAT Asp	ATC Ile	ACA Thr	<b>7</b> 87
15	GCC Ala	ACC Thr 230	AGC Ser	AAC Asn	CAC His	TGG Trp	GTG Val 235	GTC Val	AAC Asn	CCT Pro	CGG Arg	CAC His 240	AAC Asn	CTG Leu	GGC Gly	TTA Leu	835
20	CAG Gln 245	CTC Leu	TCT Ser	GTG Val	GAG Glu	ACC Thr 250	CTG Leu	GAT Asp	GGG Gly	CAG Gln	AGC Ser 255	ATC Ile	AAC Asn	CCC Pro	AAG Lys	TTG Leu 260	883
20	GCA Ala	GGC Gly	CTG Leu	ATT Ile	GGA Gly 265	CGG Arg	CAT His	GGA Gly	CCC Pro	CAG Gln 270	AAC Asn	AAG Lys	CAA Gln	CCC Pro	TTC Phe 275	ATG Met	931
25	GTG Val	GCC Ala	TTC Phe	TTC Phe 280	AAG Lys	GCC Ala	ACG Thr	GAA Glu	GTC Val 285	CAT His	CTC Leu	CGT Arg	AGT Ser	ATC Ile 290	CGG Arg	TCC Ser	979
30	ACG Thr	GGG Gly	GGC Gly 295	AAG Lys	CAG Gln	CGC Arg	AGC Ser	CAG Gln 300	AAT Asn	CGC Arg	TCC Ser	AAG Lys	ACG Thr 305	CCA Pro	AAG Lys	AAC Asn	1027
35	CAA Gln	GAG Glu 310	GCC Ala	CTG Leu	AGG Arg	ATG Met	GCC Ala 315	AGT Ser	GTG Val	GCA Ala	GAA Glu	AAC Asn 320	AGC Ser	AGC Ser	AGT Ser	GAC Asp	1075
••	CAG Gln 325	AGG Arg	CAG Gln	GCC Ala	TGC Cys	AAG Lys 330	AAA Lys	CAT His	GAG Glu	CTG Leu	TAC Tyr 335	GTC Val	AGC Ser	TTC Phe	CGA Arg	GAC Asp 340	1123
40	CTT Leu	GGC Gly	TGG Trp	CAG Gln	GAC Asp 345	Trp	ATC Ile	ATT Ile	GCA Ala	CCT Pro 350	GAA Glu	GGC Gly	TAT Tyr	GCT Ala	GCC Ala 355	TAC Tyr	1171
45	TAC Tyr	TGT Cys	GAG Glu	GGA Gly 360		TGC Cys	GCC Ala	TTC Phe	CCT Pro 365	Leu	AAC Asn	TCC Ser	TAC Tyr	ATG Met 370	AAC Asn	GCC Ala	1219

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	ACC AAC CAC GCC ATC GTC CAG ACA CTG GTT CAC TTC ATC AAC CCA GAC Thr Asn His Ala Ile Val Gln Thr Leu Val His Phe Ile Asn Pro Asp 375	1267
5	ACA GTA CCC AAG CCC TGC TGT GCG CCC ACC CAG CTC AAC GCC ATC TCT Thr Val Pro Lys Pro Cys Cys Ala Pro Thr Gln Leu Asn Ala Ile Ser 390 395	1315
10	GTC CTC TAC TTC GAC GAC AGC TCT AAT GTC GAC CTG AAG AAG TAC AGA Val Leu Tyr Phe Asp Asp Ser Ser Asn Val Asp Leu Lys Lys Tyr Arg 410 415	1363
15	AAC ATG GTG GTC CGG GCC TGT GGC TGC CAC TAGCTCTTCC TGAGACCCTG Asn Met Val Val Arg Ala Cys Gly Cys His 425 430	1413
	ACCTTTGCGG GGCCACACCT TTCCAAATCT TCGATGTCTC ACCATCTAAG TCTCTCACTG	1473
	CCCACCTTGG CGAGGAGAAC AGACCAACCT CTCCTGAGCC TTCCCTCACC TCCCAACCGG	1533
20	AAGCATGTAA GGGTTCCAGA AACCTGAGCG TGCAGCAGCT GATGAGCGCC CTTTCCTTCT	1593
	GGCACGTGAC GGACAAGATC CTACCAGCTA CCACAGCAAA CGCCTAAGAG CAGGAAAAAT	1653
25	GTCTGCCAGG AAAGTGTCCA GTGTCCACAT GGCCCCTGGC GCTCTGAGTC TTTGAGGAGT	1713
	AATCGCAAGC CTCGTTCAGC TGCAGCAGAA GGAAGGGCTT AGCCAGGGTG GGCGCTGGCG	1773
	TCTGTGTTGA AGGGAAACCA AGCAGAAGCC ACTGTAATGA TATGTCACAA TAAAACCCAT	1833
30	GAATGAAAAA AAAAAAAAA AAAAAAAAA AAAAGAATTC	1873
	(2) INFORMATION FOR SEQ ID NO:19:	
35	<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 430 amino acids</li><li>(B) TYPE: amino acid</li></ul>	
40	(D) TOPOLOGY: linear	
40	(ii) MOLECULE TYPE: protein	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:	
45	Met His Val Arg Ser Leu Arg Ala Ala Ala Pro His Ser Phe Val Ala 1 5 10 15	
50	Leu Trp Ala Pro Leu Phe Leu Leu Arg Ser Ala Leu Ala Asp Phe Ser 20 25 30	

	Leu	Asp	Asn 35	Glu	Val	His	Ser	Ser 40	Phe	Ile	His	Arg	Arg 45	Leu	Arg	Ser	
5	Gln	Glu 50	Arg	Arg	Glu	Met	Gln 55	Arg	Glu	Ile	Leu	Ser 60	Ile	Leu	Gly	Leu	
	Pro 65	His	Arg	Pro	Arg	Pro 70	His	Leu	Gln	Gly	Lys 75	His	Asn	Ser	Ala	Pro 80	
10	Met	Phe	Met	Leu	Asp 85	Leu	Tyr	Asn	Ala	Met 90	Ala	Val	Glu	Glu	Ser 95	Gly	
	Pro	Asp	Gly	Gln 100	Gly	Phe	Ser	Tyr	Pro 105	Tyr	Lys	Ala	Val	Phe 110	Ser	Thr	
15	Gln	Gly	Pro 115	Pro	Leu	Ala	Ser	Leu 120	Gln	Asp	Ser	His	Phe 125	Leu	Thr	Asp	
20	Ala	Asp 130	Met	Val	Met	Ser	Phe 135	Val	Asn	Leu	Val	Glu 140	His	Asp	Lys	Glu	
	Phe 145	Phe	His	Pro	Arg	Tyr 150	His	His	Arg	Glu	Phe 155	Arg	Phe	Asp	Leu	Ser 160	
25	Lys	Ile	Pro	Glu	Gly 165	Glu	Arg	Val	Thr	Ala 170	Ala	Glu	Phe	Arg	Ile 175	Tyr	
20	Lys	Asp	Tyr	Ile 180	Arg	Glu	Arg	Phe	Asp 185	Asn	Glu	Thr	Phe	Gln 190	Ile	Thr	
30	Val	Tyr	Gln 195	Val	Leu	Gln	Glu	His 200	Ser	Gly	Arg	Glu	Ser 205	Asp	Leu	Phe	
<b>3</b> 5	Leu	Leu 210	Asp	Ser	Arg	Thr	Ile 215	Trp	Ala	Ser	Glu	Glu 220	Gly	Trp	Leu	Val	
	Phe 225	-	Ile	Thr	Ala	Thr 230		Asn	His	Trp	Val 235	Val	Asn	Pro	Arg	His 240	
40	Asn	Leu	Gly	Leu	Gln 245		Ser	Val	Glu	Thr 250	Leu	Asp	Gly	Gln	Ser 255	Ile	
	Asn	Pro	Lys	Leu 260		Gly	Leu	Ile	Gly 265		His	Gly	Pro	Gln 270	Asn	Lys	
45	Gln	Pro	Phe 275		. Val	Ala	Phe	Phe 280		Ala	Thr	Glu	Val 285	His	Leu	Arg	

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	Ser	Ile 290	Arg	Ser	Thr	Gly	Gly 295	Lys	Gln	Arg	Ser	Gln 300	Asn	Arg	Ser	Lys
5	Thr 305	Pro	Lys	Asn	Gln	Glu 310	Ala	Leu	Arg	Met	Ala 315	Ser	Val	Ala	Glu	Asn 320
	Ser	Ser	Ser	Asp	Gln 325	Arg	Gln	Ala	Cys	Lys 330	Lys	His	Glu	Leu	Tyr 335	Val
10	Ser	Phe	Arg	Asp 340	Leu	Gly	Trp	Gln	Asp 345	Trp	Ile	Ile	Ala	Pro 350	Glu	Gly
15	Tyr	Ala	Ala 355	Tyr	Tyr	Cys	Glu	Gly 360	Glu	Cys	Ala	Phe	Pro 365	Leu	Asn	Ser
15	Tyr	Met 370	Asn	Ala	Thr	Asn	His 375	Ala	Ile	Val	Gln	Thr 380	Leu	Val	His	Phe
20	Ile 385	Asn	Pro	Asp	Thr	Val 390	Pro	Lys	Pro	Cys	Cys 395	Ala	Pro	Thr	Gln	Leu 400
	Asn	Ala	Ile	Ser	Val 405	Leu	Tyr	Phe	Asp	Asp 410	Ser	Ser	Asn	Val	Asp 415	Leu
25	Lys	Lys	Tyr	Arg 420	Asn	Met	Val	Val	Arg 425	Ala	Cys	Gly	Cys	His 430		
	(2)	INF	ORMA:	CION	FOR	SEQ	ID 1	10:20	):							
30		(i)		À) LI	ENGTI	IARA( I: 1; nuc.	723 I	oase	pair	rs						
			()	c) s:	[RANI	DEDNI DGY:	ESS:	sing								
35		( <b>ii</b> )	) MO	LECUI	LE T	TPE:	cDNA	A								
		(vi		A) 01	RGAN:	ISM:	Homo									
40			•	•		E TYI	PE: I	HIPP	OCAMI	PUS						
45		(ix	Ò	A) NA B) L(	AME/I OCAT: THER 'p	KEY: ION: INF( roducte=	490. ORMA: ct= '	rion hop:	: /fi 2-PP'	17	ion=	"OS:	reog!	ENIC	PRO	CEIN'
50		(xi	) SE	QUEN	CE DI	ESCR:	IPTI	ON:	SEQ :	ID NO	20:20	:				

#### - 120 -

	GGCGCCGGCA GAGCAGGAGT GGCTGGAGGA GCTGTGGTTG GAGCAGGAGG TGGCACGGCA	60
	GGGCTGGAGG GCTCCCTATG AGTGGCGGAG ACGGCCCAGG AGGCGCTGGA GCAACAGCTC	120
5	CCACACCGCA CCAAGCGGTG GCTGCAGGAG CTCGCCCATC GCCCCTGCGC TGCTCGGACC	180
	GCGGCCACAG CCGGACTGGC GGGTACGGCG GCGACAGAGG CATTGGCCGA GAGTCCCAGT	240
10	CCGCAGAGTA GCCCCGGCCT CGAGGCGGTG GCGTCCCGGT CCTCTCCGTC CAGGAGCCAG	300
10	GACAGGTGTC GCGCGGGGG GCTCCAGGGA CCGCGCCTGA GGCCGGCTGC CCGCCCGTCC	360
	CGCCCCGCCC CGCCCCCCC CGCCCGCCGA GCCCAGCCTC CTTGCCGTCG GGGCGTCCCC	420
15	AGGCCCTGGG TCGGCCGCGG AGCCGATGCG CGCCCGCTGA GCGCCCCAGC TGAGCGCCCC	480
	CGGCCTGCC ATG ACC GCG CTC CCC GGC CCG CTC TGG CTC CTG GGC CTG Met Thr Ala Leu Pro Gly Pro Leu Trp Leu Leu Gly Leu	528
20	1 5 10	
_	GCG CTA TGC GCG CTG GGC GGG GGC CCC GGC CTG CGA CCC CCG CCC Ala Leu Cys Ala Leu Gly Gly Gly Pro Gly Leu Arg Pro Pro 15 20 25	576
25	GGC TGT CCC CAG CGA CGT CTG GGC GCG CGC GAG CGC CGG GAC GTG CAG Gly Cys Pro Gln Arg Arg Leu Gly Ala Arg Glu Arg Arg Asp Val Gln	624
	30 35 40 45	
30	CGC GAG ATC CTG GCG GTG CTC GGG CTG CCT GGG CGG C	672
30	50 55 60	
	GCG CCA CCC GCC GCC TCC CGG CTG CCC GCG TCC GCG CCG C	720
<b>3</b> 5	65 70 75	
	CTG GAC CTG TAC CAC GCC ATG GCC GGC GAC GAC GAC GAC GGC GCG Leu Asp Leu Tyr His Ala Met Ala Gly Asp Asp Glu Asp Gly Ala	. 768
40	80 85 90	
40	CCC GCG GAG CGG CGC CTG GGC CGC GCC GAC CTG GTC ATG AGC TTC GTT	816
	Pro Ala Glu Arg Arg Leu Gly Arg Ala Asp Leu Val Met Ser Phe Val 95 100 105	
45	AAC ATG GTG GAG CGA GAC CGT GCC CTG GGC CAC CAG GAG CCC CAT TGG Asn Met Val Glu Arg Asp Arg Ala Leu Gly His Gln Glu Pro His Trp	864
	110 115 120 125	

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	AAG Lys	GAG Glu	TTC Phe	CGC Arg	TTT Phe 130	GAC Asp	CTG Leu	ACC Thr	CAG Gln	ATC Ile 135	CCG Pro	GCT Ala	GGG Gly	GAG Glu	GCG Ala 140	GTC Val	91:	2
5	ACA Thr	GCT Ala	GCG Ala	GAG Glu 145	TTC Phe	CGG Arg	ATT Ile	TAC Tyr	AAG Lys 150	GTG Val	CCC Pro	AGC Ser	ATC Ile	CAC His 155	CTG Leu	CTC Leu	960	O
10	AAC Asn	AGG Arg	ACC Thr 160	CTC Leu	CAC His	GTC Val	AGC Ser	ATG Met 165	TTC Phe	CAG Gln	GTG Val	GTC Val	CAG Gln 170	GAG Glu	CAG Gln	TCC Ser	100	В
15	AAC Asn	AGG Arg 175	GAG Glu	TCT Ser	GAC Asp	TTG Leu	TTC Phe 180	TTT Phe	TTG Leu	GAT Asp	CTT Leu	CAG Gln 185	ACG Thr	CTC Leu	CGA Arg	GCT Ala	105	6
20	GGA Gly 190	GAC Asp	GAG Glu	GGC Gly	TGG Trp	CTG Leu 195	GTG Val	CTG Leu	GAT Asp	GTC Val	ACA Thr 200	GCA Ala	GCC Ala	AGT Ser	GAC Asp	TGC Cys 205	110	4
20	TGG Trp	TTG Leu	CTG Leu	AAG Lys	CGT Arg 210	CAC His	AAG Lys	GAC Asp	CTG Leu	GGA Gly 215	CTC Leu	CGC Arg	CTC Leu	TAT Tyr	GTG Val 220	GAG Glu	115	2
<b>2</b> 5	ACT Thr	GAG Glu	GAC Asp	GGG Gly 225	CAC His	AGC Ser	GTG Val	GAT Asp	CCT Pro 230	GGC Gly	CTG Leu	GCC Ala	GGC Gly	CTG Leu 235	CTG Leu	GGT Gly	120	0
30	CAA Gln	CGG Arg	GCC Ala 240	CCA Pro	CGC Arg	TCC Ser	CAA Gln	CAG Gln 245	CCT Pro	TTC Phe	GTG Val	GTC Val	ACT Thr 250	TTC Phe	TTC Phe	AGG Arg	124	8
35	GCC Ala	AGT Ser 255	CCG Pro	AGT Ser	CCC Pro	ATC Ile	CGC Arg 260	ACC Thr	CCT Pro	CGG Arg	GCA Ala	GTG Val 265	AGG Arg	CCA Pro	CTG Leu	AGG Arg	129	6
40	AGG Arg 270	AGG Arg	CAG Gln	CCG Pro	AAG Lys	AAA Lys 275	AGC Ser	AAC Asn	GAG Glu	CTG Leu	CCG Pro 280	CAG Gln	GCC Ala	AAC Asn	CGA Arg	CTC Leu 285	134	4
40	CCA Pro	GGG Gly	ATC Ile	TTT Phe	GAT Asp 290	GAC Asp	GTC Val	CAC His	GGC Gly	TCC Ser 295	CAC His	GGC Gly	CGG Arg	CAG Gln	GTC Val 300	TGC Cys	139	2
45	CGT Arg	CGG Arg	CAC His	GAG Glu 305	CTC Leu	TAC Tyr	GTC Val	AGC Ser	TTC Phe 310	CAG Gln	GAC Asp	CTC Leu	GGC Gly	TGG Trp 315	CTG Leu	GAC Asp	144	0

	TGG Trp	GTC Val	ATC Ile 320	GCT Ala	CCC Pro	CAA Gln	GGC Gly	TAC Tyr 325	TCG Ser	GCC Ala	TAT Tyr	TAC Tyr	TGT Cys 330	GAG Glu	GGG Gly	GAG Glu		1488
5	TGC Cys	TCC Ser 335	TTC Phe	CCA Pro	CTG Leu	GAC Asp	TCC Ser 340	TGC Cys	ATG Met	AAT Asn	GCC Ala	ACC Thr 345	AAC Asn	CAC His	GCC Ala	ATC Ile		1536
10	CTG Leu 350	CAG Gln	TCC Ser	CTG Leu	GTG Val	CAC His 355	CTG Leu	ATG Met	AAG Lys	CCA Pro	AAC Asn 360	GCA Ala	GTC Val	CCC Pro	AAG Lys	GCG Ala 365	•	1584
15	TGC Cys	TGT Cys	GCA Ala	CCC Pro	ACC Thr 370	AAG Lys	CTG Leu	AGC Ser	GCC Ala	ACC Thr 375	TCT Ser	GTG Val	CTC Leu	TAC Tyr	TAT Tyr 380	GAC Asp		1632
20	AGC Ser	AGC Ser	AAC Asn	AAC Asn 385	GTC Val	ATC Ile	CTG Leu	CGC Arg	AAA Lys 390	GCC Ala	CGC Arg	AAC Asn	ATG Met	GTG Val 395	GTC Val	AAG Lys		1680
20			GGC Gly 400			T G	AGTC	AGCC	C GC(	CCAG	CCCT	ACT	GCAG					1723
25	(2)	INF	ORMA?	rion	FOR	SEQ	ID 1	NO:2	1:									
30		l	(i) \$	(A (B		NGTH PE: a	: 402 amin	2 am:	ino a id	: acid	s							
		(:	ii) l	MOLE	CULE	TYP	E: p:	rote	in									
35		(:	xi) :	SEQU	ENCE	DES	CRIP'	TION	: SE	Q ID	NO:	21:						
	Met 1	Thr	Ala	Leu	Pro 5	Gly	Pro	Leu	Trp	Leu 10	Leu	Gly	Leu	Ala	Leu 15	Cys		
40	Ala	Leu	Gly	Gly 20	Gly	Gly	Pro	Gly	Leu 25	Arg	Pro	Pro	Pro	Gly 30	Cys	Pro		
	Gln	Arg	Arg 35	Leu	Gly	Ala	Arg	Glu 40		Arg	Asp	Val	Gln 45	Arg	Glu	Ile		
45	Leu	Ala 50		Leu	Gly	Leu	Pro 55		Arg	Pro	Arg	Pro 60	Arg	Ala	Pro	Pro		

	Ala 65	Ala	Ser	Arg	Leu	Pro 70	Ala	Ser	Ala	Pro	Leu 75	Phe	Met	Leu	Asp	Leu 80
5	Tyr	His	Ala	Met	Ala 85	Gly	Asp	Asp	Asp	Glu 90	Asp	Gly	Ala	Pro	Ala 95	Glu
	Arg	Arg	Leu	Gly 100	Arg	Ala	Asp	Leu	Val 105	Met	Ser	Phe	Val	Asn 110	Met	Val
10	Glu	Arg	Asp 115	Arg	Ala	Leu	Gly	His 120	Gln	Glu	Pro	His	Trp 125	Lys	Glu	Phe
15	Arg	Phe 130	Asp	Leu	Thr	Gln	Ile 135	Pro	Ala	Gly	Glu	Ala 140	Val	Thr	Ala	Ala
15	Glu 145	Phe	Arg	Ile	Tyr	Lys 150	Val	Pro	Ser	Ile	His 155	Leu	Leu	Asn	Arg	Thr 160
20	Leu	His	Val	Ser	Met 165	Phe	Gln	Val	Val	Gln 170	Glu	Gln	Ser	Asn	Arg 175	Glu
	Ser	Asp	Leu	Phe 180	Phe	Leu	Asp	Leu	Gln 185	Thr	Leu	Arg	Ala	Gly 190	Asp	Glu
25	Gly	Trp	Leu 195	Val	Leu	Asp	Val	Thr 200	Ala	Ala	Ser	Asp	Cys 205	Trp	Leu	Leu
30	Lys	Arg 210	His	Lys	Asp	Leu	Gly 215	Leu	Arg	Leu	Tyr	Val 220	Glu	Thr	Glu	Asp
<b>J</b> 0	Gly 225	His	Ser	Val	Asp	Pro 230	Gly	Leu	Ala	Gly	Leu 235	Leu	Gly	Gln	Arg	Ala 240
35	Pro	Arg	Ser	Gln	Gln 245	Pro	Phe	Val	Val	Thr 250	Phe	Phe	Arg	Ala	Ser 255	Pro
	Ser	Pro	Ile	Arg 260	Thr	Pro	Arg	Ala	Val 265	Arg	Pro	Leu	Arg	Arg 270	Arg	Glr
40	Pro	Lys	Lys 275	Ser	Asn	Glu	Leu	Pro 280	Gln	Ala	Asn	Arg	Leu 285	Pro	Gly	Ile
45	Phe	Asp 290		Val	His	Gly	Ser 295		Gly	Arg	Gln	Val 300	Cys	Arg	Arg	His
-1-7	Glu 305	Leu	Tyr	Val	Ser	Phe 310		Asp	Leu	Gly	Trp 315	Leu	Asp	Trp	Val	11e 320
50	Ala	Pro	Gln	Gly	Tyr 325	Ser	Ala	Tyr	Tyr	Cys 330	Glu	Gly	Glu	Cys	Ser 335	Phe

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	Pro	Leu	Asp	Ser 340	Cys	Met	Asn	Ala	Thr 345	Asn	His	Ala	Ile	<b>Leu</b> 350	GIn	Ser		
5	Leu	Val	His 355	Leu	Met	Lys	Pro	Asn 360	Ala	Val	Pro	Lys	Ala 365	Cys	Cys	Ala		
	Pro	Thr 370	Lys	Leu	Ser	Ala	Thr 375	Ser	Val	Leu	Tyr	Tyr 380	Asp	Ser	Ser	Asn	÷	
10	Asn 385	Val	Ile	Leu	Arg	Lys 390	Ala	Arg	Asn	Met	Val 395	Val	Lys	Ala	Cys	Gly 400		•
	Cys	His																
15	(2)	INF	ORMA'	TION	FOR	SEQ	ID I	NO: 22	2:									
20		(i	(. (. (.	QUENCA) LIB) T'CO	ENGTI YPE: IRANI	nuc. DEDNI	926   leic ESS:	base acio sin	pai: d	rs								
25		(vi	(.	IGINA A) O F) T	RGAN:	ISM:	MUR	IDAE EMBR	YO									
30		(ix	(	ATUR A) N B) L D) O	AME/I OCAT THER 'p	ION:	93. ORMA ct=	TION "mOP	: /f 2-PP	unct	ion=	"OS	TEOG	ENIC	PRO	TEIN"		
35		(xi	) SE	QUEN	CE D	ESCR	IPTI	ON:	SEQ	ID N	0:22	:				·		
	GCC	AGGC	ACA	GGTG	CGCC	GT C	TGGT	CCTC	с сс	GTCT	GGCG	TCA	GCCG	AGC	CCGA	CCAGCT	•	60
40	ACC	AGTG	GAT	GCGC	GCCG	GC I	GAAA	GTCC	G AG	ATG Met	GCT Ala	ATG Met	CGT	CCC Pro	GGG Gly	CCA		113
<b>4</b> 5	CTC Leu	TGG	CTA Leu 10	Leu	GGC Gly	CTT Leu	GCT Ala	CTG Leu 15	Cys	GCG Ala	CTG Leu	GGA Gly	GGC Gly 20	GIA	CAC His	GGT Gly		161
50	CCG Pro	CGT Arg	Pro	CCG Pro	CAC His	ACC	TGT Cys	Pro	CAG Gln	CGT	CGC Arg	CTG Leu 35	Gly	GCG Ala	CGC	GAG Glu		209

	CGC Arg 40	CGC Arg	GAC Asp	ATG Met	CAG Gln	CGT Arg 45	GAA Glu	ATC Ile	CTG Leu	GCG Ala	GTG Val 50	CTC Leu	GGG Gly	CTA Leu	CCG Pro	GGA Gly 55	257
5	CGG Arg	CCC Pro	CGA Arg	CCC Pro	CGT Arg 60	GCA Ala	CAA Gln	CCC Pro	GCC Ala	GCT Ala 65	GCC Ala	CGG Arg	CAG Gln	CCA Pro	GCG Ala 70	TCC Ser	305
10	GCG Ala	CCC Pro	CTC Leu	TTC Phe 75	ATG Met	TTG Leu	GAC Asp	CTA Leu	TAC Tyr 80	CAC His	GCC Ala	ATG Met	ACC Thr	GAT Asp 85	GAC Asp	GAC Asp	<b>35</b> 3
15	GAC Asp	GGC Gly	GGG Gly 90	CCA Pro	CCA Pro	CAG Gln	GCT Ala	CAC His 95	TTA Leu	GGC Gly	CGT Arg	GCC Ala	GAC Asp 100	CTG Leu	GTC Val	ATG Met	401
20	AGC Ser	TTC Phe 105	GTC Val	AAC Asn	ATG Met	GTG Val	GAA Glu 110	CGC Arg	GAC Asp	CGT Arg	ACC Thr	CTG Leu 115	GGC Gly	TAC Tyr	CAG Gln	GAG Glu	449
20	CCA Pro 120	CAC His	TGG Trp	AAG Lys	GAA Glu	TTC Phe 125	CAC His	TTT Phe	GAC Asp	CTA Leu	ACC Thr 130	CAG Gln	ATC Ile	CCT Pro	GCT Ala	GGG Gly 135	497
25	GAG Glu	GCT Ala	GTC Val	ACA Thr	GCT Ala 140	GCT Ala	GAG Glu	TTC Phe	CGG Arg	ATC Ile 145	TAC Tyr	AAA Lys	GAA Glu	CCC Pro	AGC Ser 150	ACC Thr	545
30	CAC His	CCG Pro	CTC Leu	AAC Asn 155	ACA Thr	ACC Thr	CTC Leu	CAC His	ATC Ile 160	AGC Ser	ATG Met	TTC Phe	GAA Glu	GTG Val 165	GTC Val	CAA Gln	593
35	GAG Glu	CAC His	TCC Ser 170	AAC Asn	AGG Arg	GAG Glu	TCT Ser	GAC Asp 175	TTG Leu	TTC Phe	TTT Phe	TTG Leu	GAT Asp 180	CTT Leu	CAG Gln	ACG Thr	641
40	CTC Leu	CGA Arg 185		GGG Gly	GAC Asp	GAG Glu	GGC Gly 190	TGG Trp	CTG Leu	GTG Val	CTG Leu	GAC Asp 195	ATC Ile	ACA Thr	GCA Ala	GCC Ala	689
40	AGT Ser 200	Asp	CGA Arg	TGG Trp	CTG Leu	CTG Leu 205	Asn	CAT His	CAC His	AAG Lys	GAC Asp 210	Leu	GGA Gly	CTC Leu	CGC Arg	CTC Leu 215	737
45	TAT Tyr	GTG Val	GAA Glu	ACC Thr	GCG Ala 220	Asp	GGG Gly	CAC His	AGC	ATG Met 225	GAT Asp	CCT Pro	GGC Gly	CTG Leu	GCT Ala 230	Gly	785

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	CTG Leu	CTT Leu	GGA Gly	CGA Arg 235	CAA Gln	GCA Ala	CCA Pro	CGC Arg	TCC Ser 240	AGA Arg	CAG Gln	CCT Pro	TTC Phe	ATG Met 245	GTA Val	ACC Thr	833
5	TTC Phe	TTC Phe	AGG Arg 250	GCC Ala	AGC Ser	CAG Gln	AGT Ser	CCT Pro 255	GTG Val	CGG Arg	GCC Ala	CCT Pro	CGG Arg 260	GCA Ala	GCG Ala	AGA Arg	881
10	CCA Pro	CTG Leu 265	AAG Lys	AGG Arg	AGG Arg	CAG Gln	CCA Pro 270	AAG Lys	AAA Lys	ACG Thr	AAC Asn	GAG Glu 275	CTT Leu	CCG Pro	CAC His	CCC Pro	929
15	AAC Asn 280	AAA Lys	CTC Leu	CCA Pro	GGG Gly	ATC Ile 285	TTT Phe	GAT Asp	GAT Asp	GGC Gly	CAC His 290	GGT Gly	TCC Ser	CGC Arg	GGC Gly	AGA Arg 295	977
	GAG Glu	GTT Val	TGC Cys	CGC Arg	AGG Arg 300	CAT His	GAG Glu	CTC Leu	TAC Tyr	GTC Val 305	AGC Ser	TTC Phe	CGT Arg	GAC Asp	CTT Leu 310	GGC Gly	1025
20	TGG Trp	CTG Leu	GAC Asp	TGG Trp 315	GTC Val	ATC Ile	GCC Ala	CCC Pro	CAG Gln 320	GGC Gly	TAC Tyr	TCT Ser	GCC Ala	TAT Tyr 325	TAC Tyr	TGT Cys	1073
25	GAG Glu	GGG Gly	GAG Glu 330	Cys	GCT Ala	TTC Phe	CCA Pro	CTG Leu 335	GAC Asp	TCC Ser	TGT Cys	ATG Met	AAC Asn 340	Ala	ACC Thr	AAC Asn	1121
30	CAT His	GCC Ala 345	Ile	TTG Leu	CAG Gln	TCT Ser	CTG Leu 350	GTG Val	CAC His	CTG Leu	ATG Met	AAG Lys 355	CCA Pro	GAT Asp	GTT Val	GTC Val	1169
35	CCC Pro 360	Lys	GCA Ala	TGC Cys	TGT Cys	GCA Ala 365	CCC Pro	ACC Thr	AAA Lys	CTG Leu	AGT Ser 370	Ala	ACC	TCT	GTG Val	CTG Leu 375	1217
	TAC Tyr	TAT Tyr	GAC Asp	AGC Ser	AGC Ser 380	Asn	AAT Asn	GTC Val	ATC Ile	CTG Leu 385	Arg	AAA Lys	CAC	CGT	AAC Asn 390	ATG Met	1265
40	GTG Val	GTC Val	AAG Lys	GCC Ala 395	Cys	GGC Gly	TGC Cys	CAC His	TGA	GGCC	CCG	CCCA	GCAT	CC I	GCTT	CTACT	1319
45	ACC	TTAC	CAT	CTGG	CCGG	GC C	CCTC	TCCA	.G A.G	GCAG	AAAC	CCI	TCTA	TGT	TATC	ATAGCT	1379
	CAG	ACAG	GGG	CAAT	'GGGA	.GG C	CCTT	CACI	T CC	CCTG	GCCA	CTI	CCTC	CTA	TAAA	TCTGGT	1439
50	CTT	TCCC	AGT	TCCI	CTGT	CC T	TCAT	GGGG	T TI	CGGG	GCTA	TCA	cccc	GCC	CTCI	CCATCC	<b>149</b> 9

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	TCCT	ACCC	CA A	GCAT	'AGAC	T GA	ATGU	ACAU	AGU	ATCC	CAG	AGCI	AIGU	TH I	1CIGA	GAGGI
	CTGG	GGTC	AG C	CACTO	AAGG	c co	CACAI	GAGG	AAG	ACTG	ATC	CTTC	GCCA	TC (	CTCAG	CCCAC
5	AATG	GCAA	I TA	CTG	ATGG	T CI	TAAGA	AGGC	CC1	GGAA	TTC	TAAA	CTAC	AT (	GATCI	GGGCT
	CTCT	GCAC	CA I	TCAI	TGTG	G CA	GTTG	GGAC	TTA:	TTTA	GGT	ATAA	CAGA	CA (	CATAC	ACTTA
••	GATC	AATG	CA I	CGCI	GTAC	T CO	CTTGA	AATC	AGA	GCTA	GCT	TGTI	AGAA	AA A	AGAAT	CAGAG
10	CCAG	GTAI	AG C	CGGTC	CATO	T CA	ATTA	TCCC	AGC	CGCTA	LAAG	AGAC	AGAC	GAC A	AGGAG	SAATCT
	CTGT	'GAG'I	TC A	AGGC	CCACA	A TA	GAAAG	SAGCO	TGI	CTC	GGA	GCAG	GAAA	AAA	AAAAA	AAAAC
15	GGAA	TTC														
	(2)	TNFO	DM A T	ידחא	FOR	SEO	TD N	in•23	<b>!</b> •							
20	(2)				ENCE											
20		(	, <b>1</b> ) 2	(A)	LEN TYF	IGTH:	399	ami	no a		5					
				(D)	TOE	OLO	3Y: ]	linea	ir							
25		(i	.i) ł	OLE	CULE	TYP	E: p1	otei	in							
		(x	(i)	SEQUI	ENCE	DES	CRIPT	CION:	SEC	Q ID	NO:	23:				
30	Met 1	Ala	Met	Arg	Pro 5	Gly	Pro	Leu	Trp	Leu 10	Leu	Gly	Leu	Ala	Leu 15	Cys
	Ala	Leu	Gly	Gly 20	Gly	His	Gly	Pro	Arg 25	Pro	Pro	His	Thr	Cys 30	Pro	Gln
35	Arg	Arg	Leu 35	Gly	Ala	Arg	Glu	Arg 40	Arg	Asp	Met	Gln	Arg 45	Glu	Ile	Leu
••	Ala	Val 50	Leu	Gly	Leu	Pro	Gly 55	Arg	Pro	Arg	Pro	Arg 60	Ala	Gln	Pro	Ala
40	Ala 65	Ala	Arg	Gln	Pro	Ala 70	Ser	Ala	Pro	Leu	Phe 75	Met	Leu	Asp	Leu	<b>Tyr</b> 80
<b>4</b> 5	His	Ala	Met	Thr	Asp 85	Asp	Asp	Asp	Gly	Gly 90	Pro	Pro	Gln	Ala	His 95	Leu
	Gly	Arg	Ala	Asp 100	Leu	Val	Met	Ser	Phe 105	Val	Asn	Met	Val	Glu 110	Arg	Asp

PCT/US93/07190

	Arg	Thr	<b>Leu</b> 115	Gly	Tyr	Gln	Glu	Pro 120	His	Trp	Lys	Glu	Phe 125	His	Phe	Asp
5	Leu	Thr 130	Gln	Ile	Pro	Ala	Gly 135	Glu	Ala	Val	Thr	Ala 140	Ala	Glu	Phe	Arg
	Ile 145	Tyr	Lys	Glu	Pro	Ser 150	Thr	His	Pro	Leu	Asn 155	Thr	Thr	Leu	His	Ile 160
10	Ser	Met	Phe	Glu	Val 165	Val	Gln	Glu	His	Ser 170	Asn	Arg	Glu	Ser	Asp 175	Leu
<b>1</b> F	Phe	Phe	Leu	Asp 180	Leu	Gln	Thr	Leu	Arg 185	Ser	Gly	Asp	Glu	Gly 190	Trp	Leu
15	Val	Leu	Asp 195	Ile	Thr	Ala	Ala	Ser 200	Asp	Arg	Trp	Leu	Leu 205	Asn	His	His
20	Lys	Asp 210	Leu	Gly	Leu	Arg	Leu 215	Tyr	Val	Glu	Thr	Ala 220	Asp	Gly	His	Ser
	Met 225	Asp	Pro	Gly	Leu	Ala 230	Gly	Leu	Leu	Gly	Arg 235	Gln	Ala	Pro	Arg	Ser 240
25	Arg	Gln	Pro	Phe	Met 245	Val	Thr	Phe	Phe	Arg 250	Ala	Ser	Gln	Ser	Pro 255	Va]
20	Arg	Ala	Pro	Arg 260	Ala	Ala	Arg	Pro	Leu 265	Lys	Arg	Arg	Gln	Pro 270	Lys	Lys
30	Thr	Asn	Glu 275		Pro	His	Pro	Asn 280	Lys	Leu	Pro	Gly	Ile 285	Phe	Asp	Ası
35	Gly	His 290	Gly	Ser	Arg	Gly	Arg 295	Glu	Val	Cys	Arg	Arg 300	His	Glu	Leu	Ту
	Val 305		Phe	Arg	Asp	Leu 310		Trp	Leu	Asp	Trp 315	Val	Ile	Ala	Pro	Gl: 320
40	Gly	Tyr	Ser	Ala	Tyr 325	Tyr	Cys	Glu	Gly	Glu 330	Cys	Ala	Phe	Pro	Leu 335	As
45	Ser	Cys	Met	340		Thr	Asn	His	Ala 345	Ile	Leu	Gln	Ser	Leu 350	Val	Hi

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	Leu	Met	Lys 355	Pro	Asp	Val	Val	Pro 360	Lys	Ala	Cys	Cys	Ala 365	Pro	Thr	Lys		
5	Leu	Ser 370	Ala	Thr	Ser	Val	Leu 375	Tyr	Tyr	Asp	Ser	Ser 380	Asn	Asn	Val	Ile		
	Leu 385	Arg	Lys	His	Arg	Asn 390	Met	Val	Val	Lys	Ala 395	Cys	Gly	Cys	His			
LO	(2)	INF	ORMAI	NOI	FOR	SEQ	ID 1	10:24	<b>:</b> :							,		
15		(i)	(E	i) LI 3) Ti 3) Si	ENGTI PE: PRANI	I: 13 nuc] EDNI	TER 368 l leic ESS: line	ase acio sing	pain 1	:s								
		(ii)	) MOI	LECUI	LE TY	PE:	cDNA	A										
20		(ix)	) FE# (# (I	A) NA	ME/I	KEY:	CDS 1	L368										
25		(xi	) SEC	QUENC	CE DI	ESCR:	IPTI(	ON: S	SEQ :	ID NO	0:24:	:						
30	ATG Met 1	TCG Ser	GGA Gly	CTG Leu	CGA Arg 5	AAC Asn	ACC Thr	TCG Ser	GAG Glu	GCC Ala 10	GTT Val	GCA Ala	GTG Val	CTC Leu	GCC Ala 15	TCC Ser	4	8
	CTG Leu	GGA Gly	CTC Leu	GGA Gly 20	ATG Met	GTT Val	CTG Leu	CTC Leu	ATG Met 25	TTC Phe	GTG Val	GCG Ala	ACC Thr	ACG Thr 30	CCG Pro	CCG Pro	9	6
35	GCC Ala	GTT Val	GAG Glu 35	GCC Ala	ACC Thr	CAG Gln	TCG Ser	GGG Gly 40	ATT Ile	TAC Tyr	ATA Ile	GAC Asp	AAC Asn 45	GGC Gly	AAG Lys	GAC Asp	14	4
40	CAG Gln	ACG Thr 50	ATC Ile	ATG Met	CAC His	AGA Arg	GTG Val 55	CTG Leu	AGC Ser	GÁG Glu	GAC Asp	GAC Asp 60	AAG Lys	CTG Leu	GAC Asp	GTC Val	19	)2
45	TCG Ser 65	TAC Tyr	GAG Glu	ATC Ile	CTC Leu	GAG Glu 70	Phe	CTG Leu	GGC Gly	ATC Ile	GCC Ala 75	GAA Glu	CGG Arg	CCG Pro	ACG Thr	CAC His 80	24	ŧ0

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	CTG Leu	AGC Ser	AGC Ser	CAC His	CAG Gln 85	TTG Leu	TCG Ser	CTG Leu	AGG Arg	AAG Lys 90	TCG Ser	GCT Ala	CCC Pro	AAG Lys	TTC Phe 95	CTG Leu	2	88
5	CTG Leu	GAC Asp	GTC Val	TAC Tyr 100	CAC His	CGC Arg	ATC Ile	ACG Thr	GCG Ala 105	GAG Glu	GAG Glu	GGT Gly	CTC Leu	AGC Ser 110	GAT Asp	CAG Gln	3	336
10	GAT Asp	GAG Glu	GAC Asp 115	GAC Asp	GAC Asp	TAC Tyr	GAA Glu	CGC Arg 120	GGC Gly	CAT His	CGG Arg	TCC Ser	AGG Arg 125	AGG Arg	AGC Ser	GCC Ala	3	384
15	GAC Asp	CTC Leu 130	GAG Glu	GAG Glu	GAT Asp	GAG Glu	GGC Gly 135	GAG Glu	CAG Gln	CAG Gln	AAG Lys	AAC Asn 140	TTC Phe	ATC Ile	ACC Thr	GAC Asp	4	432
20	CTG Leu 145	GAC Asp	AAG Lys	CGG Arg	GCC Ala	ATC Ile 150	GAC Asp	GAG Glu	AGC Ser	GAC Asp	ATC Ile 155	ATC Ile	ATG Met	ACC Thr	TTC Phe	CTG Leu 160	1	¥80
20	AAC Asn	AAG Lys	CGC Arg	CAC His	CAC His 165	AAT Asn	GTG Val	GAC Asp	GAA Glu	CTG Leu 170	CGT Arg	CAC His	GAG Glu	CAC His	GGC Gly 175	CGT Arg	<u>.</u>	528
25	CGC Arg	CTG Leu	TGG Trp	TTC Phe 180	GAC Asp	GTC Val	TCC Ser	AAC Asn	GTG Val 185	CCC Pro	AAC Asn	GAC Asp	AAC Asn	TAC Tyr 190	CTG Leu	GTG Val		576
30	ATG Met	GCC Ala	GAG Glu 195	CTG Leu	CGC Arg	ATC Ile	TAT Tyr	CAG Gln 200	AAC Asn	GCC Ala	AAC Asn	GAG Glu	GGC Gly 205	AAG Lys	TGG Trp	CTG Leu	Ć	624
35	ACC Thr	GCC Ala 210	AAC Asn	AGG Arg	GAG Glu	TTC Phe	ACC Thr 215	ATC Ile	ACG Thr	GTA Val	TAC Tyr	GCC Ala 220	ATT Ile	GGC Gly	ACC Thr	GGC Gly		672
40	ACG Thr 225	CTG Leu	GGC Gly	CAG Gln	CAC His	ACC Thr 230	ATG Met	GAG Glu	CCG Pro	CTG Leu	TCC Ser 235	TCG Ser	GTG Val	AAC Asn	ACC Thr	ACC Thr 240		720
40	GGG Gly	GAC Asp	TAC Tyr	GTG Val	GGC Gly 245	Trp	TTG Leu	GAG Glu	CTC Leu	AAC Asn 250	Val	ACC Thr	GAG Glu	GGC Gly	CTG Leu 255	CAC His	•	768
45	GAG Glu	TGG Trp	CTG Leu	GTC Val 260	Lys	TCG Ser	AAG Lys	GAC Asp	AAT Asn 265	His	GGC Gly	ATC Ile	TAC Tyr	ATT Ile 270	GGA Gly	GCA Ala	;	816

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	CAC His	GCT Ala	GTC Val 275	AAC Asn	CGA Arg	CCC Pro	GAC Asp	CGC Arg 280	GAG Glu	GTG Val	AAG Lys	CTG Leu	GAC Asp 285	GAC Asp	ATT Ile	GGA Gly	864
5	CTG Leu	ATC Ile 290	CAC His	CGC Arg	AAG Lys	GTG Val	GAC Asp 295	GAC Asp	GAG Glu	TTC Phe	CAG Gln	CCC Pro 300	TTC Phe	ATG Met	ATC Ile	GGC Gly	912
10	TTC Phe 305	TTC Phe	CGC Arg	GGA Gly	CCG Pro	GAG Glu 310	CTG Leu	ATC Ile	AAG Lys	GCG Ala	ACG Thr 315	GCC Ala	CAC His	AGC Ser	AGC Ser	CAC His 320	960
15	CAC His	AGG Arg	AGC Ser	AAG Lys	CGA Arg 325	AGC Ser	GCC Ala	AGC Ser	CAT His	CCA Pro 330	CGC Arg	AAG Lys	CGC Arg	AAG Lys	AAG Lys 335	TCG Ser	1008
20	GTG Val	TCG Ser	CCC Pro	AAC Asn 340	AAC Asn	GTG Val	CCG Pro	CTG Leu	CTG Leu 345	GAA Glu	CCG Pro	ATG Met	GAG Glu	AGC Ser 350	ACG Thr	CGC Arg	1056
20	AGC Ser	TGC Cys	CAG Gln 355	ATG Met	CAG Gln	ACC Thr	CTG Leu	TAC Tyr 360	ATA Ile	GAC Asp	TTC Phe	AAG Lys	GAT Asp 365	CTG Leu	GGC Gly	TGG Trp	1104
25	CAT His						CCA Pro 375										1152
30							CTC Leu										1200
35							GTC Val										1248
40	AAG Lys	CCC Pro	TGC Cys	TGC Cys 420	GCT Ala	CCG Pro	ACC Thr	AGG Arg	CTG Leu 425	GGA Gly	GCA Ala	CTA Leu	CCC Pro	GTT Val 430	CTG Leu	TAC Tyr	1296
40							GTG Val										1344
45						TGC Cys	CAT His 455	TGA			٠						1368

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#### (2) INFORMATION FOR SEQ ID NO:25:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 455 amino acids(B) TYPE: amino acid

  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:25: 10

Met Ser Gly Leu Arg Asn Thr Ser Glu Ala Val Ala Val Leu Ala Ser

- 15 Leu Gly Leu Gly Met Val Leu Leu Met Phe Val Ala Thr Thr Pro Pro
  - Ala Val Glu Ala Thr Gln Ser Gly Ile Tyr Ile Asp Asn Gly Lys Asp 35
- 20 Gln Thr Ile Met His Arg Val Leu Ser Glu Asp Asp Lys Leu Asp Val
- Ser Tyr Glu Ile Leu Glu Phe Leu Gly Ile Ala Glu Arg Pro Thr His 25

Leu Ser Ser His Gln Leu Ser Leu Arg Lys Ser Ala Pro Lys Phe Leu

- Leu Asp Val Tyr His Arg Ile Thr Ala Glu Glu Gly Leu Ser Asp Gln
  - Asp Glu Asp Asp Asp Tyr Glu Arg Gly His Arg Ser Arg Arg Ser Ala
- 35 Asp Leu Glu Glu Asp Glu Gly Glu Gln Gln Lys Asn Phe Ile Thr Asp
- Leu Asp Lys Arg Ala Ile Asp Glu Ser Asp Ile Ile Met Thr Phe Leu 40 145
  - Asn Lys Arg His His Asn Val Asp Glu Leu Arg His Glu His Gly Arg 170
- Arg Leu Trp Phe Asp Val Ser Asn Val Pro Asn Asp Asn Tyr Leu Val
  - Met Ala Glu Leu Arg Ile Tyr Gln Asn Ala Asn Glu Gly Lys Trp Leu 200 195

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	Thr	Ala 210	Asn	Arg	Glu	Phe	Thr 215	Ile	Thr	Val	Tyr	Ala 220	Ile	Gly	Thr	Gly
5	Thr 225	Leu	Gly	Gln	His	Thr 230	Met	Glu	Pro	Leu	Ser 235	Ser	Val	Asn	Thr	Thr 240
	Gly	Asp	Tyr	Val	Gly 245	Trp	Leu	Glu	Leu	Asn 250	Val	Thr	Glu	Gly	Leu 255	His
10	Glu	Trp	Leu	Val 260	Lys	Ser	Lys	Asp	Asn 265	His	Gly	Ile	Tyr	Ile 270	Gly	Ala
<b>.</b>	His	Ala	<b>Val</b> 275	Asn	Arg	Pro	Asp	Arg 280	Glu	Val	Lys	Leu	Asp 285	Asp	Ile	Gly
15	Leu	Ile 290	His	Arg	Lys	Val	Asp 295	Asp	Glu	Phe	Gln	Pro 300	Phe	Met	Ile	Gly
20	Phe 305	Phe	Arg	Gly	Pro	Glu 310	Leu	Ile	Lys	Ala	Thr 315	Ala	His	Ser	Ser	His 320
	His	Arg	Ser	Lys	Arg 325	Ser	Ala	Ser	His	Pro 330	Arg	Lys	Arg	Lys	Lys 335	Ser
25	Val	Ser	Pro	Asn 340	Asn	Val	Pro	Leu	Leu 345	Glu	Pro	Met	Glu	Ser 350	Thr	Arg
20	Ser	Cys	Gln 355	Met	Gln	Thr	Leu	Tyr 360	Ile	Asp	Phe	Lys	Asp 365	Leu	Gly	Trp
30	His	Asp 370	Trp	Ile	Ile	Ala	Pro 375	Glu	Gly	Tyr	Gly	Ala 380	Phe	Tyr	Cys	Ser
35	Gly 385	Glu	Cys	Asn	Phe	Pro 390	Leu	Asn	Ala	His	Met 395	Asn	Ala	Thr	Asn	His 400
	Ala	Ile	Val	Gln	Thr 405	Leu	Val	His	Leu	Leu 410	Glu	Pro	Lys	Lys	Val 415	Pro
40	Lys	Pro	Cys	Cys 420	Ala	Pro	Thr	Arg	Leu 425	Gly	Ala	Leu	Pro	Val 430	Leu	Tyr
4.5	His	Leu	Asn 435		Glu	Asn	Val	Asn 440	Leu	Lys	Lys	Tyr	Arg 445	Asn	Met	Ile
45	Val	Lys 450	Ser	Cys	Gly	Cys	His 455									

5		(i)	(A) (B) (C)	LENCE TYP STF TOP	GTH: E: a KANDE	104 mino DNES	ami aci S: s	no a .d :ingl	cids	•							
10		(ii)	MOLE	CULE	TYF	e: p	rote	in									
15		(ix)	(A) (B) (D)	NAM LOC OTH	IE/KI CATI( IER ]	N: 1 INFOR	L10	)4 [ON:				3 11					
		(xi)	SEQU	JENCE	E DES	CRIE	OITS	I: SI	EQ II	NO:	26:						
20		Cys 1	Ala	Arg	Arg	Tyr 5	Leu	Lys	Val	Asp	Phe 10	Ala	Asp	Ile	Gly	Trp 15	Ser
		Glu	Trp	Ile	<b>Ile</b> 20	Ser	Pro	Lys	Ser	Phe 25	Asp	Ala	Tyr	Tyr	Cys 30	Ser	Gly
25		Ala	Cys	Gln 35	Phe	Pro	Met	Pro	Lys 40	Ser	Leu	Lys	Pro	Ser 45	Asn	His	Ala
30		Thr	Ile 50	Gln	Ser	Ile	Val	Ala 55	Arg	Ala	Val	Gly	Val 60	Val	Pro	Gly	Ile
		Pro 65	Glu	Pro	Cys	Cys	<b>Val</b> 70	Pro	Glu	Lys	Met	Ser 75	Ser	Leu	Ser	Ile	Leu 80
<b>3</b> 5		Phe	Phe	Asp	Glu	Asn 85	Lys	Asn	Val	Val	Leu 90	Lys	Val	Tyr	Pro	Asn 95	Met
		Thr	Val	Glu	Ser 100	Cys	Ala	Cys	Arg								•
40	(2)	INFO	RMAT:	ION I	FOR :	SEQ :	ID N	D: 27	:								
45		(i)	(A (B (C	UENCI ) LEI ) TY! ) STI ) TO!	NGTH PE: ( RAND)	: 10: amin EDNE:	2 am: o ac: SS:	ino a id sing	acid	S							

(ii) MOLECULE TYPE: protein

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PCT/US93/07190

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	(VI)	(A)			SM: I		SAP	IENS								
5	(ix)	(B	) NAI ) LO	ME/K	EY: 1 ON: 1 INFOI	110	)2	/not	te= '	"BMP!	5"					
10	(xi)	SEQ	UENC	E DE	SCRII	PTIO	N: S	EQ II	ON O	:27:						
	Cys 1	Lys	Lys	His	Glu 5	Leu	Tyr	Val	Ser	Phe 10	Arg	Asp	Leu	Gly	Trp 15	Gln
15	Asp	Trp	Ile	Ile 20	Ala	Pro	Glu	Gly	Tyr 25	Ala	Ala	Phe	Tyr	Cys 30	Asp	Gly
20	Glu	Cys	Ser 35	Phe	Pro	Leu	Asn	Ala 40	His	Met	Asn	Ala	Thr 45	Asn	His	Ala
20	Ile	Val 50	Gln	Thr	Leu	Val	His 55	Leu	Met	Phe	Pro	Asp 60	His	Val	Pro	Lys
<b>2</b> 5	Pro 65	Cys	Cys	Ala	Pro	Thr 70	Lys	Leu	Asn	Ala	Ile 75	Ser	Val	Leu	Tyr	Phe 80
	Asp	Asp	Ser	Ser	Asn 85	Val	Ile	Leu	Lys	Lys 90	Tyr	Arg	Asn	Met	Val 95	Val
30	Arg	Ser	Cys	Gly 100	Cys	His										
	(2) INFO	RMAT:	ION 1	FOR :	SEQ :	ID N	0:28	:							٠	
35	(i)	(B (C	) LEI ) TYI ) STI	NGTH PE: RAND	ARAC: 102 amine EDNE:	2 am: o ac: SS:	ino a id sing:	acid	s							
40	(ii)	HOL	•													
	(vi)	ORI (A			URCE		SAP	IENS								
<b>45</b> <b>50</b>	(ix)	(B	) NA	ME/K CATI	EY: 1 ON: 1	11	02	/no	te=	"BMP	6"					
J-U																

	(xi)	SEQU	JENCE	E DES	CRIE	PTION	l: SI	EQ II	NO:	:28:						
_	Cys 1	Arg	Lys	His	Glu 5	Leu	Tyr	Val	Ser	Phe 10	Gln	Asp	Leu	Gly	Trp 15	Gln
5	Asp	Trp	Ile	Ile 20	Ala	Pro	Lys	Gly	Tyr 25	Ala	Ala	Asn	Tyr	Cys 30	Asp	Gly
10	Glu	Cys	Ser 35	Phe	Pro	Leu	Asn	Ala 40	His	Met	Asn	Ala	Thr 45	Asn	His	Ala
	Ile	<b>Val</b> 50	Gln	Thr	Leu	Val	His 55	Leu	Met	Asn	Pro	Glu 60	Tyr	Val	Pro	Lys
15	Pro 65	Cys	Cys	Ala	Pro	Thr 70	Lys	Leu	Asn	Ala	Ile 75	Ser	Val	Leu	Tyr	Phe 80
20	Asp	Asp	Asn	Ser	Asn 85	Val	Ile	Leu	Lys	Lys 90	Tyr	Arg	Trp	Met	Val 95	Val
20	Arg	Ala	Cys	Gly 100	Cys	His										
25	(2) INFO	RMAT:	ION I	FOR S	SEQ :	ED NO	0:29	:								
25	(i)	(A)	JENCI ) LEI ) TYI ) TOI	NGTH:	: 102 amino	2 am:	ino a id		s							
30	(ii)															
35	(ix)	(A)	) NAI ) LO	ME/KI CATIO HER	ON:	110	02 ION:	/lai	bel=	OPX	TND	FDFN	ጉፑህጥ	LY S	F1 FC'	ren
40				FROI	A M	GROU	P OF	ONE	OR 3	MORE	SPE	CIFI	ED A	MINO N II	ACI	DS
	(xi)	SEQ	UENC	E DE	SCRI	PTIO	N: S	EQ I	D NO	:29:						
45	Cys 1	Xaa	Xaa	His	Glu 5	Leu	Tyr	Val	Xaa	Phe 10	Xaa	Asp	Leu	Gly	Trp 15	Xaa
	Asp	Trp	Xaa	Ile 20	Ala	Pro	Xaa	Gly	Tyr 25	Xaa	Ala	Tyr	Tyr	Cys 30	Glu	Gly
50				20												

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	Glu	Cys	Xaa 35	Phe	Pro	Leu	Xaa	Ser 40	Xaa	Met	Asn	Ala	Thr 45	Asn	His	Ala
5	Ile	Xaa 50	Gln	Xaa	Leu	Val	His 55	Xaa	Xaa	Xaa	Pro	<b>Xa</b> a 60	Xaa	Val	Pro	Lys
	Xaa 65	Cys	Cys	Ala	Pro	Thr 70	Xaa	Leu	Xaa	Ala	Xaa 75	Ser	Val	Leu	Tyr	<b>Xaa</b> 80
10	Asp	Xaa	Ser	Xaa	Asn 85	Val	Xaa	Leu	Xaa	Lys 90	Xaa	Arg	Asn	Met	Val 95	Val
15	Xaa	Ala	Cys	Gly 100	Cys	His										
	(2) INFO	RMAT:	ION :	FOR S	SEQ I	ED NO	30:30	:								
20	(i)	(B (C	) LEI ) TY! ) ST!	E CHANGTH: PE: a RANDI POLOG	: 97 amino EDNES	amin ac: SS: 8	no ad id sing:	cids								
25	(ii)	MOL	ECUL	E TYI	PE: 1	prot	ein									
30	(ix)	(B	) NA	ME/KI CATION HER I /not FROI	ON: INFO te= M A (	19 RMAT "WHE GROU	7 ION: REIN	EAC! ONE	H XA.	A IS MORE	IND SPE	EPEN CIFI	5 DENT: ED Al	LY S MINO	ELEC'	red DS
<b>35</b>	(xi)	SEQ	UENC	E DE	SCRI	PTIO	N: S	EQ I	D NO	:30:						
40	Leu 1	Xaa	Xaa	Xaa	Phe 5	Xaa	Xaa	Xaa	Gly	Trp 10	Xaa	Xaa	Trp	Xaa	Xaa 15	Xaa
40	Pro	Xaa	Xaa	<b>Xaa</b> 20	Xaa	Ala	Xaa	Tyr	Cys 25	Xaa	Gly	Xaa	Cys	<b>Xa</b> a 30	Xaa	Pro
<b>4</b> 5	Xaa	Xaa	Xaa 35	Xaa	Xaa	Xaa	Xaa	Xaa 40	Asn	His	Ala	Xaa	Xaa 45	Xaa	Xaa	Xaa

- 138 -

	2		Xaa 50	Xaa	Xaa	Xaa	Xaa	Xaa 55	Xaa	Xaa	Xaa	Xaa	<b>Xaa</b> 60	Cys	Cys	Xaa	Pro
5		Xaa 65	Xaa	Xaa	Xaa	Xaa	<b>Xaa</b> 70	Xaa	Xaa	Leu	Xaa	<b>Xaa</b> 75	Xaa	Xaa	Xaa	Xaa	<b>X</b> aa 80
	,	Val	Xaa	Leu	Xaa	Xaa 85	Xaa	Xaa	Xaa	Met	<b>Xaa</b> 90	Val	Xaa	Xaa	Cys	<b>Xaa</b> 95	Cys
LO	:	Х́аа															
	(2) I	NFOR	ITAM	ON F	OR S	SEQ 1	D NO	31:	:								
15		(i)	(A) (B)	LEN TYP	E CHA NGTH: PE: a	: 102 imino	ami aci	ino a id	cids	5							
20	(	ii)	• •		OLOC TYP												
	(	,	11011	.0011		. <b></b> .	,,,,,,										
25	(	ix)	(A) (B)	LOC	IE/KI CATIO IER ]	ON: 1 INFOR	L10	)2 [ON:	/lal	oel=	GENI	ERIC-	-SEQ6	ó Senti	LY SI	71.EC1	ren
30					FRON	e= A A ( DEFIN	ROUI	OF	ONE	OR 1	MORE	SPE	CIFIE	ED Al	INO	ACII	DS .
	(	xi)	SEQU	JENCI	E DES	SCRII	PTIO	N: S1	EQ II	) NO:	:31:						
35		Cys 1	Xaa	Xaa	Xaa	Xaa 5	Leu	Xaa	Xaa	Xaa	Phe 10	Xaa	Xaa	Xaa	Gly	Trp 15	Xaa
40		Xaa	Trp	Xaa	Xaa 20	Xaa	Pro	Xaa	Xaa	<b>Xaa</b> 25	Xaa	Ala	Xaa	Tyr	Cys 30	Xaa	Gly
40	•	Xaa	Cys	Xaa 35	Xaa	Pro	Xaa	Xaa	<b>X</b> aa 40	Xaa	Xaa	Xaa	Xaa	Xaa 45	Asn	His	Ala
45		Xaa	<b>Xaa</b> 50	Xaa	Xaa	Xaa	Xaa	Xaa 55	Xaa	Xaa	Xaa	Xaa	<b>Xaa</b> 60	Xaa	Xaa	Xaa	Xaa

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	Xaa 65	Cys (	Cys Xaa	Pro	Xaa 70	Xaa	Xaa	Xaa	Xaa	Xaa 75	Xaa	Xaa	Leu	Xaa	<b>Xaa</b> 80	
5	Xaa	Xaa X	Xaa Xaa	Xaa 85	Val	Xaa	Leu	Xaa	Xaa 90	Xaa	Xaa	Xaa	Met	Xaa 95	Val	
	Xaa	Xaa (	Cys Xaa 100		Xaa											
10	(2) INFO	RMATI	ON FOR	SEQ 1	D NO	:32:	:									
<b>15</b> ,	(i)	(A) (B) (C)	ENCE CH LENGTH TYPE: STRAND TOPOLO	: 124 nucle EDNES	7 ba eic a SS: s	se p cid ingl	airs	5								
	(ii)	MOLE	CULE TY	PE: c	DNA											
20	(vi)	(A)	INAL SO ORGANI TISSUE	SM: F	OMO		ENS									
25	(ix)	(B)	NAME/K LOCATI OTHER	ON: .8	341 RMATI	ON:		oduc <sup>.</sup>	t= "(	GDF-	1"					
30	(xi)	SEQU	ENCE DE	SCRII	PTION	I: SI	EQ II	D NO	:32:							
	GGGGACAC	CG GC	ccccccc	T CAC	GCCCA	CTG	GTC	CCGG	GCC	GCCG	CGGA	CC C	rgcg	CACT	C	60
35	TCTGGTCA	TC GC	CTGGGAG	G AAC		: Pro			o Gl	G CAA n Gli 5						110
40	GGC CAC Gly His 10	His L	eu Leu	Leu I	Leu I	Leu A	Ala 1	Leu 🗆	Leu	Leu l	CCC :	rcg ( Ser )	CTG Leu	CCC Pro 25		158
45	CTG ACC Leu Thr	CGC G	CC CCC la Pro 30	GTG ( Val 1	CCC C	CCA (	GGC (	CCA Pro .	GCC Ala	GCC (	GCC (	CTG ( Leu :	CTC Leu 40	CAG Gln		206
	GCT CTA Ala Leu	Gly L	TG CGC eu Arg 45	GAT ( Asp (	GAG C Glu F	CCC (	CAG ( Gln (	GGT Gly	GCC Ala	CCC A	AGG ( Arg ]	CTC ( Leu ,	CGG Arg	CCG Pro		254

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	GTT Val	CCC Pro	CCG Pro 60	GTC Val	ATG Met	TGG Trp	CGC Arg	CTG Leu 65	TTT Phe	CGA Arg	CGC Arg	CGG Arg	GAC Asp 70	CCC Pro	CAG Gln	GAG Glu	302
5	ACC Thr	AGG Arg 75	TCT Ser	GGC Gly	TCG Ser	CGG Arg	CGG Arg 80	ACG Thr	TCC Ser	CCA Pro	GGG Gly	GTC Val 85	ACC Thr	CTG Leu	CAA Gln	CCG Pro	350
10	TGC Cys 90	CAC His	GTG Val	GAG Glu	GAG Glu	CTG Leu 95	GGG Gly	GTC Val	GCC Ala	GGA Gly	AAC Asn 100	ATC Ile	GTG Val	CGC Arg	CAC His	ATC Ile 105	398
15	CCG Pro	GAC Asp	CGC Arg	GGT Gly	GCG Ala 110	CCC Pro	ACC Thr	CGG Arg	GCC Ala	TCG Ser 115	GAG Glu	CCT Pro	GTC Val	TCG Ser	GCC Ala 120	GCG Ala	446
20	GGG Gly	CAT His	TGC Cys	CCT Pro 125	GAG Glu	TGG Trp	ACA Thr	GTC Val	GTC Val 130	TTC Phe	GAC Asp	CTG Leu	TCG Ser	GCT Ala 135	GTG Val	GAA Glu	494
20	CCC Pro	GCT Ala	GAG Glu 140	CGC Arg	CCG Pro	AGC Ser	CGG Arg	GCC Ala 145	CGC Arg	CTG Leu	GAG Glu	CTG Leu	CGT Arg 150	TTC Phe	GCG Ala	GCG Ala	542
25	GCG Ala	GCG Ala 155	GCG Ala	GCA Ala	GCC Ala	CCG Pro	GAG Glu 160	GGC Gly	GGC Gly	TGG Trp	GAG Glu	CTG Leu 165	AGC Ser	GTG Val	GCG Ala	CAA Gln	590
30	GCG Ala 170	GGC Gly	CAG Gln	GGC Gly	GCG Ala	GGC Gly 175	GCG Ala	GAC Asp	CCC Pro	GGG Gly	CCG Pro 180	GTG Val	CTG Leu	CTC Leu	CGC Arg	CAG Gln 185	638
35	TTG Leu	GTG Val	CCC Pro	GCC Ala	CTG Leu 190	GGG Gly	CCG Pro	CCA Pro	GTG Val	CGC Arg 195	GCG Ala	GAG Glu	CTG Leu	CTG Leu	GGC Gly 200	GCC Ala	<b>6</b> 86
	GCT Ala	TGG Trp	GCT Ala	CGC Arg 205	AAC Asn	GCC Ala	TCA Ser	TGG Trp	CCG Pro 210	CGC Arg	AGC Ser	CTC Leu	CGC Arg	CTG Leu 215	GCG Ala	CTG Leu	734
40	GCG Ala	CTA Leu	CGC Arg 220	CCC Pro	CGG Arg	GCC Ala	CCT Pro	GCC Ala 225	GCC Ala	TGC Cys	GCG Ala	CGC Arg	CTG Leu 230	GCC Ala	GAG Glu	GCC Ala	<b>7</b> 82
45	TCG Ser	CTG Leu 235	CTG Leu	CTG Leu	GTG Val	ACC Thr	CTC Leu 240	GAC Asp	CCG Pro	CGC Arg	CTG Leu	TGC Cys 245	CAC	CCC Pro	CTG Leu	GCC Ala	830

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	CGG Arg 250	CCG Pro	CGG Arg	CGC Arg	GAC Asp	GCC Ala 255	GAA Glu	CCC Pro	GTG Val	TTG Leu	GGC Gly 260	GGC Gly	GGC Gly	CCC Pro	GGG Gly	GGC Gly 265	878
5	GCT Ala	TGT Cys	CGC Arg	GCG Ala	CGG Arg 270	CGG Arg	CTG Leu	TAC Tyr	GTG Val	AGC Ser 275	TTC Phe	CGC Arg	GAG Glu	GTG Val	GGC Gly 280	TGG Trp	926
10	CAC His	CGC Arg	TGG Trp	GTC Val 285	ATC Ile	GCG Ala	CCG Pro	CGC Arg	GGC Gly 290	TTC Phe	CTG Leu	GCC Ala	AAC Asn	TAC Tyr 295	TGC Cys	CAG Gln	974
15	GGT Gly	CAG Gln	TGC Cys 300	GCG Ala	CTG Leu	CCC Pro	GTC Val	GCG Ala 305	CTG Leu	TCG Ser	GGG Gly	TCC Ser	GGG Gly 310	GGG Gly	CCG Pro	CCG Pro	1022
20	GCG Ala	CTC Leu 315	AAC Asn	CAC His	GCT Ala	GTG Val	CTG Leu 320	CGC Arg	GCG Ala	CTC Leu	ATG Met	CAC His 325	GCG Ala	GCC Ala	GCC Ala	CCG Pro	1070
20	GGA Gly 330	GCC Ala	GCC Ala	GAC Asp	CTG Leu	CCC Pro 335	TGC Cys	TGC Cys	GTG Val	CCC Pro	GCG Ala 340	CGC Arg	CTG Leu	TCG Ser	CCC Pro	ATC Ile 345	1118
25	TCC Ser	GTG Val	CTC Leu	TTC Phe	TTT Phe 350	GAC Asp	AAC Asn	AGC Ser	GAC Asp	AAC Asn 355	GTG Val	GTG Val	CTG Leu	CGG Arg	CAG Gln 360	TAT Tyr	1166
30			ATG Met									TAAC	CCCG(	GGG (	CGGG(	CAGGGA	1219
	CCC	GGC	CCA A	ACAA'	CAAA'	rg c	CGCG:	rgg									1247
35	(2)		ORMA!														
40		•	(i) :	(A (B	) LEI	CHAINGTH:	: 372 amin	2 am: o ac:	ino a id		5						
		(:	ii) l	MOLE	CULE	TYP	E: p	rote	in								
45		(:	xi) S	SEQU	ENCE	DES	CRIP:	rion:	: SE	Q ID	NO:	33:					
	Met 1	Pro	Pro	Pro	Gln 5	Gln	Gly	Pro	Cys	Gly 10	His	His	Leu	Leu	Leu 15	Leu	

Leu Ala Leu Leu Pro Ser Leu Pro Leu Thr Arg Ala Pro Val Pro 20 Pro Gly Pro Ala Ala Ala Leu Leu Gln Ala Leu Gly Leu Arg Asp Glu 5 Pro Gln Gly Ala Pro Arg Leu Arg Pro Val Pro Pro Val Met Trp Arg Leu Phe Arg Arg Arg Asp Pro Gln Glu Thr Arg Ser Gly Ser Arg Arg Thr Ser Pro Gly Val Thr Leu Gln Pro Cys His Val Glu Glu Leu Gly 15 Val Ala Gly Asn Ile Val Arg His Ile Pro Asp Arg Gly Ala Pro Thr 105 Arg Ala Ser Glu Pro Val Ser Ala Ala Gly His Cys Pro Glu Trp Thr 20 Val Val Phe Asp Leu Ser Ala Val Glu Pro Ala Glu Arg Pro Ser Arg 130 Ala Arg Leu Glu Leu Arg Phe Ala Ala Ala Ala Ala Ala Pro Glu 155 Gly Gly Trp Glu Leu Ser Val Ala Gln Ala Gly Gln Gly Ala Gly Ala 30 Asp Pro Gly Pro Val Leu Leu Arg Gln Leu Val Pro Ala Leu Gly Pro 185 Pro Val Arg Ala Glu Leu Leu Gly Ala Ala Trp Ala Arg Asn Ala Ser 35 Trp Pro Arg Ser Leu Arg Leu Ala Leu Ala Leu Arg Pro Arg Ala Pro Ala Ala Cys Ala Arg Leu Ala Glu Ala Ser Leu Leu Val Thr Leu 40 235 Asp Pro Arg Leu Cys His Pro Leu Ala Arg Pro Arg Arg Asp Ala Glu 45 Pro Val Leu Gly Gly Gly Pro Gly Gly Ala Cys Arg Ala Arg Arg Leu

	Tyr	Val	Ser 275	Phe	Arg	Glu	Val	Gly 280	Trp	His	Arg	Trp	Val 285	Ile	Ala	Pro
5	Arg	Gly 290	Phe	Leu	Ala	Asn	Tyr 295	Cys	Gln	Gly	Gln	Cys 300	Ala	Leu	Pro	Val
	Ala 305	Leu	Ser	Gly	Ser	Gly 310	Gly	Pro	Pro	Ala	Leu 315	Asn	His	Ala	Val	Leu 320
10	Arg	Ala	Leu	Met	His 325	Ala	Ala	Ala	Pro	Gly 330	Ala	Ala	Asp	Leu	Pro 335	Cys
•	Cys	Val	Pro	Ala 340	Arg	Leu	Ser	Pro	Ile 345	Ser	Val	Leu	Phe	Phe 350	Asp	Asn
15	Ser	Asp	Asn 355	Val	Val	Leu	Arg	Gln 360	Tyr	Glu	Asp	Het	Val 365	Val	Asp	Glu
20	Cys	Gly 370	Cys	Arg												

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### What is claimed is:

- 1. A morphogen-enriched infant formula composition.
- 5 2. The morphogen-enriched infant formula of claim 1 wherein said formula is milk-based.
  - 3. The morphogen-enriched infant formula of claim 1 wherein said formula is soy-based.
- The morphogen-enriched infant formula of claim 1 wherein said formula is adapted for preterm infants.
- 5. The morphogen-enriched infant formula of claim 1 wherein said formula is adapted for low birth weight infants.
  - 6. A morphogen-enriched dietary composition for individuals at risk for tissue damage due to protein-energy malnutrition.
- 7. The morphogen-enriched dietary composition of claim 6 wherein said individual is at risk for tissue damage from starvation, dehydration, anorexia nervosa or trauma.
- The morphogen-enriched dietary composition of claim 6
  wherein said individual is at risk for tissue damage
  from a malabsorption-malnutrition disorder.
- 30 9. The morphogen-enriched dietary composition of claim 8 wherein malabsorption-malnutrition disorder results from a digestive or intestinal fistula, short bowel, gastrointestinal disorder or hypercatabolism.

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10. The morphogen-enriched dietary composition of claim 6 wherein said individual is at risk for malnutrition-malabsorption induced tissue damage following radiotherapy, chemotherapy or surgery.

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- 11. A morphogen-enriched dietary composition for individuals at risk for tissue damage due to altered metabolism function.
- 10 12. The morphogen-enriched dietary composition of claim 11 wherein said individual at risk is a pregnant, lactacting or post-menopausal female.
- 13. The morphogen-enriched dietary composition of claim 11 wherein said individual at risk is an aged individual.
  - 14. The composition of claim 1, 6 or 11 wherein said morphogen is associated with a controlled release componen, adapted such that the morphogen is released in a controlled manner lower in the gastrointestinal tract.
  - 15. The composition of claim 1, 6 or 11 adapted for enteral administration.
- 25 16. The composition of claim 1, 6 or 11 adapted for aerosol administration.
  - 17. The composition of claim 1, 6 or 11 adapted for nasal administration.

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18. The composition of claim 6 or 11 formulated as a solid.

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- 19. The composition of claim 18 wherein said solid is a tablet, troche or lozenge.
- 20. The composition of claim 1, 6 or 11 formulated as a liquid.
  - 21. The composition of claim 20 wherein said liquid is a beverage or a syrup.
- 10 22. The composition of claim 1, 6 or 11 wherein said morphogen is associated with a morphogen-solubilizing molecule.
- 23. The composition of claim 22 wherein said molecule is casein or a derivative, salt or analog thereof.
  - 24. The composition of claim 22 wherein said molecule comprises part or all of a morphogen pro domain.
- 25. The composition of claim 1, 6 or 11 wherein said morphogen comprises an amino acid sequence sharing at least 70% homology with one of the sequences selected from the group consisting of: OP-1, OP-2, CBMP2, BMP3(fx), Vg1(fx), Vgr(fx), DPP(fx), GDF-1(fx) and 60A(fx).
- 26. The composition of claim 25 wherein said morphogen comprises an amino acid sequence sharing a last 80% homology with one of the sequences selected from the group consisting of: OP-1, OP-2, CBMP2, BMP3(fx), BMP5(fx), BMP6(fx), Vg1(fx), Vgr(fx), DPP(fx), GDF-1(fx) and 60A(fx).

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27. The composition of claim 1, 6 or 11 wherein said morphogen comprises an amino acid sequence having greater than 60% amino acid identity with the sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1).

28. The composition of claim 27 wherein said morphogen comprises an amino acid sequence having greater than 65% amino acid identity with the sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1).

- 29. The composition of claim 28 wherein said morphogen comprises an amino acid sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1), including allelic and species variants thereof.
- 30. The composition of claim 1, 6 or 11 wherein said morphogen comprises an amino acid sequence defined by Generic Sequences 1, 2, 3, 4, 5 or 6 (Seq. ID Nos. 1, 2, 3, 4, 30 or 31).
  - 31. The composition of claim 1, 6 or 11 wherein said morphogen comprises an amino acid sequence defined by OPX (Seq. ID No. 29).
- 25 32. The composition of claim 1, 6 or 11 wherein said morphogen comprises an amino acid sequence encoded by a nucleic acid that hybridizes under stringent conditions with a DNA defined by nucleotides 1036-1341 of Seq. ID No. 16 or nucleotides 1390-1695 of Seq. ID No. 20.

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- 33. A method for improving the human milk mimetic characteristics of an infant formula, the method comprising the step of adding a morphogenically effective concentration of a morphogen to said formula prior to providing said formula to an infant.
- 34. A method for enhancing tissue viability in a malnourished individual, the method comprising the step of providing to said individual a morphogenically effective concentration of a morphogen.
- 35. A method for enhancing tissue viability in an individual having altered metabolic function, the method comprising the step of providing to said individual a morphogenically effective concentration of a morphogen.
  - 36. The method of claim 35 wherein said individual is a pregnant, lactating or postmenopausal female.
- 20 37. The method of claim 35 wherein said individual is an aged individual.
- 38. The method of claim 34 or 35 wherein said morphogen is provided to said individual as part of a food formulation.
  - 39. The method of claim 34 or 35 wherein said morphogen is provided to said individual as a dietary supplement.

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- 40. The method of claim 33, 34 or 35 wherein said morphogen comprises an amino acid sequence sharing at least 70% homology with one of the sequences selected from the group consisting of: OP-1, OP-2, CBMP2, BMP3(fx), Vg1(fx), Vgr(fx), DPP(fx), GDF-1(fx) and 60A(fx).
- 41. The method of claim 40 wherein said morphogen comprises an amino acid sequence sharing a last 80% homology with one of the sequences selected from the group consisting of: OP-1, OP-2, CBMP2, BMP3(fx), BMP5(fx), BMP6(fx), Vg1(fx), Vgr(fx), DPP(fx), GDF-1(fx) and 60A(fx).
- 42. The method of claim 33, 34 or 35 wherein said morphogen comprises an amino acid sequence having greater than 60% amino acid identity with the sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1).
  - 43. The method of claim 42 wherein said morphogen comprises an amino acid sequence having greater than 65% amino acid identity with the sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1).
  - 44. The method of claim 43 wherein said morphogen comprises an amino acid sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1), including allelic and species variants thereof.
- 45. The method of claim 33, 34 or 35 wherein said morphogen comprises an amino acid sequence defined by Generic Sequences 1, 2, 3, 4, 5 or 6 (Seq. ID Nos. 1, 2, 3, 4, 30 or 31).

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- 46. The method of claim 33, 34 or 35 wherein said morphogen comprises an amino acid sequence defined by OPX (Seq. ID No. 29).
- 5 47. The method of claim 33, 34 or 35 wherein said morphogen comprises an amino acid sequence encoded by a nucleic acid that hybridizes under strigent conditions with a DNA defined by nucleotides 1036-1341 of Seq. ID No. 16 or nucleotides 1390-1695 of Seq. ID No. 20.
- 48. The invention of claim 1, 6, 11, 33, 34 or 35 wherein said morphogen comprises a dimeric protein species complexed with a peptide comprising a pro region of a member of the morphogen family, or an allelic, species or other sequence variant thereof.
  - 49. The invention of claim 48 wherein said dimeric morphogen species is noncovalently complexed with said peptide.
- 20 50. The invention of claim 48 or 49 wherein said dimeric morphogen species is complexed with two said peptides.
- 51. The invention of claim 48 or 49 wherein said peptide comprises at least the first eighteen amino acids of a sequence defining said pro region.
  - 52. The invention of claim 51 wherein said peptide comprises the full length of said pro region.

- 53. The invention of claim 48 or 49 wherein said peptide comprises a nucleic acid that hybridizes under stringent conditions with a DNA defined by nucleotides 136-192 of Seq. ID No. 16, or nucleotides 157-211 of Seq. ID No. 20.
- 54. The invention of claim 48 or 49 wherein said complex is further stabilized by exposure to a basic amino acid, a detergent or a carrier protein.

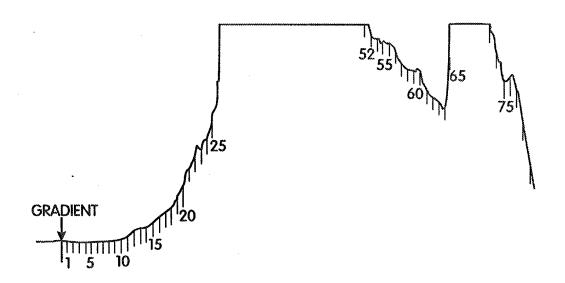
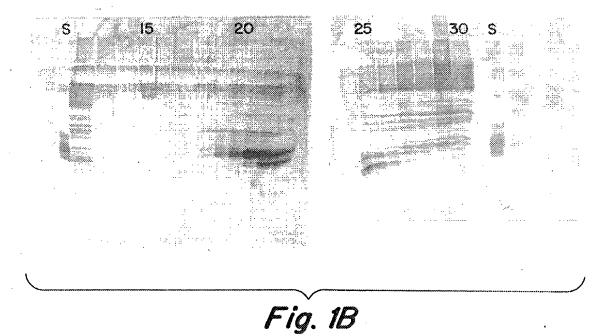


Fig. 14



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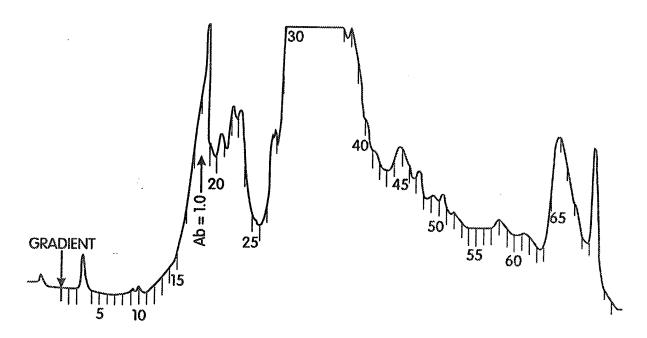
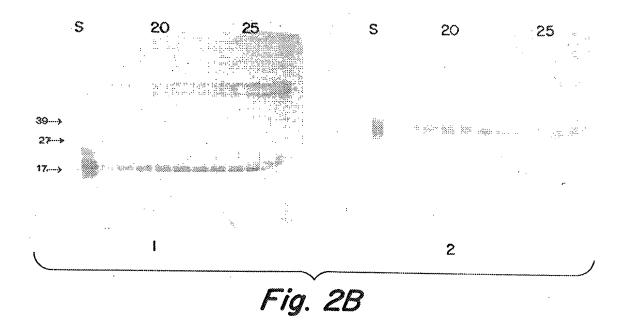


Fig. 2A



**SUBSTITUTE SHEET** 

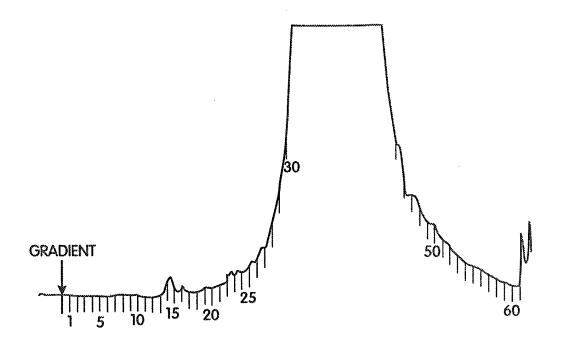
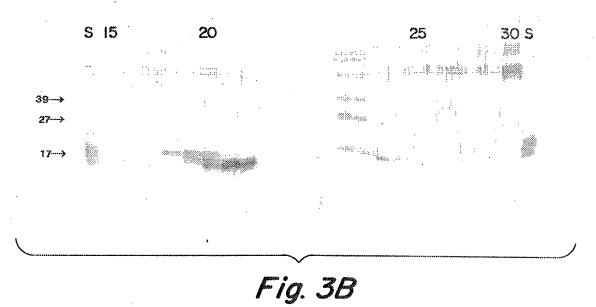


Fig. 3A



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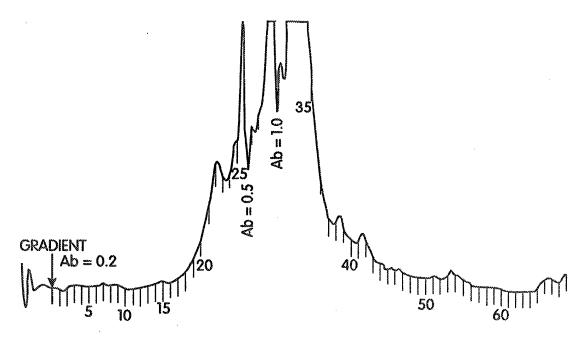
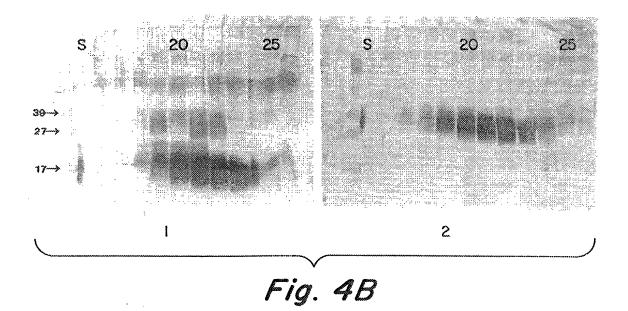


Fig. 4A



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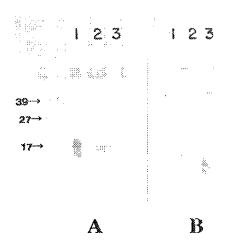


Fig. 5

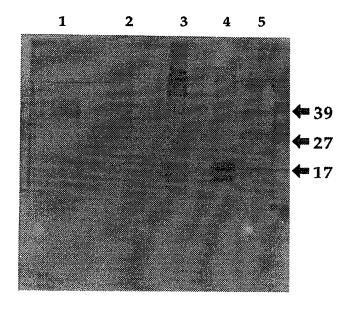
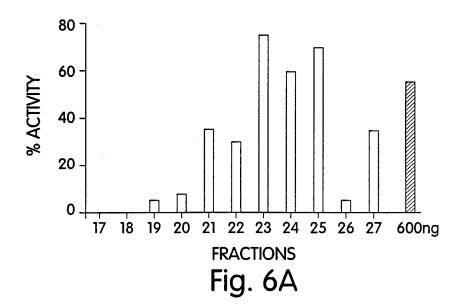
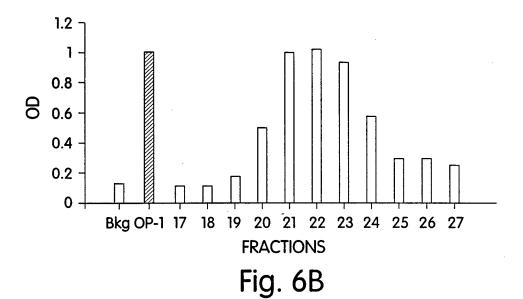


Fig. 7

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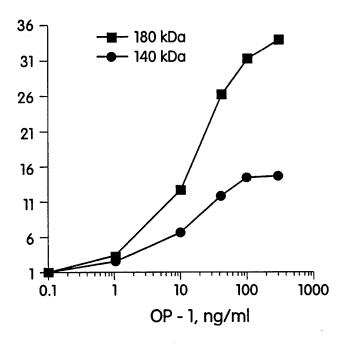


Fig. 8A

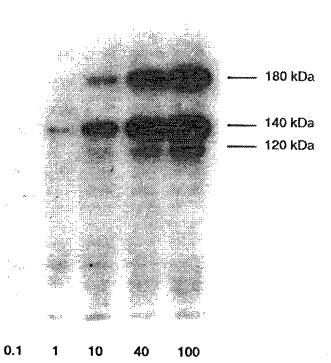


Fig. 8B

OP-1, ng/ml

## PCT

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### INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 5:

. . | ```'

(11) International Publication Number:

WO 94/03075

A23L 1/305, A61K 37/02

**A3** 

(43) International Publication Date:

17 February 1994 (17.02.94)

(21) International Application Number:

PCT/US93/07190

(22) International Filing Date:

29 July 1993 (29.07.93)

(30) Priority data:

 923,780
 31 July 1992 (31.07.92)
 US

 946,235
 16 September 1992 (16.09.92)
 US

 029,335
 4 March 1993 (04.03.93)
 US

 040,510
 31 March 1993 (31.03.93)
 US

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(74) Agent: KELLEY, Robin, D.; Testa, Hurwitz & Thibeault, Exchange Place, 53 State Street, Boston, MA 02109 (US).

(81) Designated States: AT, AU, BB, BG, BR, CA, CH, CZ, DE, DK, ES, FI, GB, HU, JP, KP, KR, LK, LU, MG, MN, MW, NL, NO, NZ, PL, PT, RO, RU, SD, SE, SK, UA, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).

#### Published

With international search report.

Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.

(88) Date of publication of the international search report: 10 November 1994 (10.11.94)

(54) Title: MORPHOGEN-ENRICHED DIETARY COMPOSITION

#### (57) Abstract

Disclosed are methods and compositions useful in dietary applications and capable of enhancing tissue morphogenesis, including tissue development and viability in a mammal, particularly a human. The methods and compositions include a morphogen which, when provided to an individual as a food formulation or supplement, is capable of enhancing tissue development and viability in the individual.

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Interna al Application No PCT/US 93/07190

A. CLASSIFICATION OF SUBJECT MATTER IPC 5 A23L1/305 A61K37/02

According to International Patent Classification (IPC) or to both national classification and IPC

#### **B. FIELDS SEARCHED**

Minimum documentation searched (classification system followed by classification symbols)  $IPC \ 5 \ A23L \ A61K$ 

THO S AZSE AUTK

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

I C.	DOCUMENTS	CONSIDERED	TO	BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Х	US,A,4 440 860 (M.KLAGSBURN) 30 August 1983	1-5, 14-17, 20-33
A	see column 1, line 63 - line 65 see column 4, line 40 - line 44	14-17, 20-32, 40-54
X	EP,A,O 295 009 (BAYLOR COLLEGE OF MEDECINE) 14 December 1988	1-5, 14-17, 20-33
A	see page 2, line 19 - line 23 see page 4, line 12 - line 20 see page 4, line 34 - page 5, line 3; claims 1-25	40-54
	-/	

X Patent family members are listed in annex.

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Date of mailing of the international search report

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Date of the actual completion of the international search

1 0, 10, 94

### 15 July 1994

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## INTERNATIONAL SEARCH REPORT

Interna. .1 Application No
PCT/US 93/07190

	ion) DOCUMENTS CONSIDERED TO BE RELEVANT	
ategory °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
-	EP,A,O 313 515 (CIBA-GEIGY) 26 April 1989	1-5, 14-17, 20-33
	see page 2, line 7-11 see page 10, line 31 - line 38; claim 19	40-54
	•	

### INTERNATIONAL SEARCH REPORT

In ational application No.
PCT/US 93/07190

Box I	Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This inte	ernational search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
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3.	Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II	Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This Int	ernational Searching Authority found multiple inventions in this international application, as follows:
C1	aims: 1-5, 33, and partially 14-17, 20-32 and 40-54 aims: 6-10, 34 and partially 14-32 and 38-54 aims: 11-13, 35-37 and partially 14-32 and 38-54
1.	As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2.	As all searchable claims could be searches without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3.	As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. X	No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remark	on Protest  The additional search fees were accompanied by the applicant's protest.  No protest accompanied the payment of additional search fees.

## INTERNATIONAL SEARCH REPORT

Information on patent family members

Interna I Application No
PCT/US 93/07190

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